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(54) Title: RECOMBINANT ANTI-VLA4 ANTIBODY MOLECULES

#### (57) Abstract

The present invention discloses recombinant anti-VLA4 antibody molecules, including humanized recombinant anti-VLA4 antibody molecules. These antibodies are useful in the treatment of specific and non-specific inflammation, including asthma and inflammatory bowel disease. In addition, the humanized recombinant anti-VLA4 antibodies disclosed can be useful in methods of diagnosing and localizing sites of inflammation.

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WO 94/16094 PCT/US94/00266

#### RECOMBINANT ANTI-VLA4 ANTIBODY MOLECULES

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#### FIELD OF THE INVENTION

The present invention relates to recombinant anti-VLA4 antibody molecules, including humanized recombinant anti-VLA4 antibody molecules.

10 BACKGROUND OF THE INVENTION

### A. Immunoglobulins and Monoclonal Antibodies

Natural immunoglobulins have been known for many years, as have the various fragments thereof, such as the Fab, (Fab')<sub>2</sub> and Fc fragments, which can be derived by enzymatic cleavage. Natural immunoglobulins comprise generally a Y-shaped molecule having an antigen-binding site towards the free end of each upper arm. The remainder of the structure, and particularly the stem of the Y, mediates the effector functions associated with immunoglobulins.

Specifically, immunoglobulin molecules are comprised of two heavy (H) and two light (L) polypeptide chains, held together by disulfide bonds. Each chain of an immunoglobulin chain is divided into regions or domains, each being approximately 110 amino acids. chain has two such domains while the heavy chain has four The amino acid sequence of the amino-terminal domains. domain of each polypeptide chain is highly variable (V region), while the sequences of the remaining domains are conserved or constant (C regions). A light chain is therefore composed of one variable  $(V_L)$  and one constant domain (C<sub>L</sub>) while a heavy chain contains one variable  $(V_H)$  and three constant domains  $(CH_1,\ CH_2\ and\ CH_3)$ . An arm of the Y-shap d molecule consists of a light chain (V +  $C_L$ ) and the variable domain  $(V_H)$  and one constant domain (CH<sub>1</sub>) of a heavy chain. The tail of the Y is composed of

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the remaining heavy chain constant d mains (CH, + CH<sub>3</sub>). The C-terminal ends of the heavy chains associate to form the Fc portion. Within each variable region are three hypervariable regions. These hypervariable regions are also described as the complementarity determining regions (CDRs) because of their importance in binding of antigen. The four more conserved regions of the variable domains are described as the framework regions (FRs). domain of an immunoglobulin consists of two beta-sheets held together by a disulfide bridge, hydrophobic faces packed together. The individual beta . strands are linked together by loops. The overall appearance can be described as a beta barrel having loops at the ends. The CDRs form the loops at one end of the beta barrel of the variable region.

Natural immunoglobulins have been used in assay, to a more limited extent, therapy. diagnosis and, However, such uses, especially in therapy, have been hindered the polyclonal by nature of natural immunoglobulins. A significant step towards realization of the potential of immunoglobulins therapeutic agents was the discovery of techniques for the preparation of monoclonal antibodies (MAbs) defined specificity, Kohler et al., 1975 [1]. most MAbs are produced by fusions of rodent (i.e., mouse, rat) spleen cells with rodent myeloma cells. They are therefore essentially rodent proteins.

By 1990, over 100 murine monoclonal antibodies were in clinical trials, particularly in the U.S. and especially for application in the treatment of cancer. However, by this time it was recognized that rejection of murine monoclonal antibodies by the undesirable immune response in humans termed the HAMA (Human Anti-Mouse Antibody) response was a severe limitation, especially for th treatment of chronic disease. Therefore, the use of rodent MAbs as therapeutic agents in humans is

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inherently limited by the fact that the human subject will mount an immunological response to the MAb and either remove the MAb entirely or at least reduce its effectiveness. In practice MAbs of rodent origin may not be used in a patient for more than one or a few treatments as a HAMA response soon develops rendering the MAb ineffective as well as giving rise to undesirable reactions. In fact, a HAMA response has been observed in the majority of patients following a single injection of mouse antibody, Schroff et al., 1985 [2]. A solution to the problem of HAMA is to administer immunologically compatible human monoclonal antibodies. However, the technology for development of human monoclonal antibodies has lagged well behind that of murine antibodies (Borrebaeck et al., 1990 [3] such that very few human antibodies have proved useful for clinical study.

Proposals have therefore been made for making nonhuman MAbs less antigenic in humans. Such techniques can be generically termed "humanization" techniques. techniques generally involve the use of recombinant DNA technology to manipulate DNA sequences encoding the polypeptide chains of the antibody molecule. recombinant DNA technology to clone antibody genes has provided an alternative whereby a murine monoclonal antibody can be converted to a predominantly human-form humanized) with the same antigen properties (Riechmann et al., 1988 [4]). Generally, the goal of the humanizing technology is to develop humanized antibodies with very little or virtually no murine component apart from the CDRs (see, e.g., Tempest et al., to reduce or [5]) so as eliminate immunogenicity in humans.

Early methods for humanizing MAbs involved production of chimeric antibodies in which an antigen binding site comprising the complete variable domains of one antibody is linked to constant domains derived from

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another antibody. Methods for carrying out such chimerizati n procedures have been described, example, in EP 120694 [6], EP 125023 [7], and WO 86/01533 Generally disclosed are processes for preparing antibody molecules having the variable domains from a non-human MAb such as a mouse MAb and the constant domains from a human immunoglobulin. Such chimeric antibodies are not truly humanized because they still contain a significant proportion of non-human amino acid sequence, i.e., the complete non-human variable domains, still elicit may some HAMA response, particularly if administered over a prolonged period, Begent et al., 1990 [9]. In addition, it is believed that these methods in some cases (e.g., EP 120694 [6]; EP 125023 [7] and U.S. Patent No. 4,816,567 [10] did not lead to the expression of any significant quantities of Ig polypeptide chains, nor the production of Ig activity without in vitro solubilization and chain reconstitution, nor to the secretion and assembly of the chains into the desired chimeric recombinant antibodies. These same problems may be noted for the initial production of nonchimeric recombinant antibodies (e.g., U.S. Patent No. 4,816,397 [11].

## B. Humanized Recombinant Antibodies and CDR-Grafting Technology

Following the early methods for the preparation of chimeric antibodies, a new approach was described in EP 0239400 [12] whereby antibodies are altered by substitution of their complementarity determining regions (CDRs) for one species with those from another. This process may be used, for example, to substitute the CDRs from human heavy and light chain Ig variable region domains with alternative CDRs from murine variable region domains. These altered Ig variable regions may subsequently be combined with human Ig constant regions to created antibodies which are totally human in composition except for the substituted murine CDRs. Such

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murine CDR-substituted antibodi s would be predicted to be less likely to elicit a considerably reduced immune response in humans compared to chimeric antibodies because they contain considerably less murine components.

The process for humanizing monoclonal antibodies via CDR grafting has been termed "reshaping". (Riechmann 1988 [4]; Verhoeyen et al., 1988 Typically, complementarity determining regions (CDRs) of a murine antibody are transplanted onto the corresponding regions in a human antibody, since it is the CDRs (three in antibody heavy chains, three in light chains) that are the regions of the mouse antibody which bind to a specific antigen. Transplantation of CDRs is achieved by genetic engineering whereby CDR DNA sequences determined by cloning of murine heavy and light chain region gene segments, (V) and are then transferred to corresponding human V regions by sitedirected mutagenesis. In the final stage of the process, human constant region gene segments of the desired isotype (usually gamma 1 for  $C_{\rm H}$  and kappa for  $C_{\rm L}$ ) are added and the humanized heavy and light chain genes are coexpressed in mammalian cells to produce soluble humanized antibody.

The transfer of these CDRs to a human antibody confers on this antibody the antigen binding properties of the original murine antibody. The six CDRs in the murine antibody are mounted structurally on a V region "framework" region. The reason that CDR-grafting is successful is that framework regions between mouse and human antibodies may have very similar 3-D structures with similar points of attachment for CDRs, such that CDRs can be interchanged. Nonetheless, certain amino acids within framework regions are thought to interact with CDRs and to influence overall antigen binding The direct transfer of CDRs from a murine affinity. antibody to produce a recombinant humanized antibody

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without any modifications f the human V region frameworks often results in a partial or complete loss of binding affinity.

In Riechmann et al., 1988 [4] and WO 89/07454 [14], it was found that transfer of the CDR regions alone (as defined by Kabat et al., 1991 [15] and Wu et al., 1970 [16] was not sufficient to provide satisfactory antigen binding activity in the CDR-grafted product. Riechmann et al. 1988 [4] found that it was necessary to convert a serine residue at position 27 of the human sequence to the corresponding rat phenylalanine residue to obtain a CDR-grafted product having satisfactory antigen binding activity. This residue at position 27 of the heavy chain is within the structural loop adjacent to CDR1. further construct which additionally contained a human serine to rat tyrosine change at position 30 of the heavy chain did not have a significantly altered binding activity over the humanized antibody with the serine to phenylalanine change at position 27 alone. These results indicate that changes to residues of the human sequence outside the CDR regions, for example, in the loop adjacent to CDR1, may be necessary to obtain effective antigen binding activity for CDR-grafted antibodies which recognize more complex antigens. Even so, the binding affinity of the best CDR-grafted antibodies obtained was still significantly less than the original MAb.

More recently, Queen et al., 1989 [17] and WO 90/07861 [18] have described the preparation of a humanized antibody that binds to the interleukin 2 receptor, by combining the CDRs of a murine MAb (anti-Tac) with human immunoglobulin framework and constant regions. They have demonstrated one solution to the problem of the loss of binding affinity that often results from direct CDR transfer without any m difications of the human V region framework residues; their s lution involves two key steps. First, the human

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V framework regions are chosen by c mputer analysis for optimal protein sequence homology to the V region framework of the original murine antibody, in this case, In the second step, the tertiary the anti-Tac MAb. structure of the murine V region is modelled by computer in order to visualize framework amino acid residues which are likely to interact with the murine CDRs and these murine amino acid residues are then superimposed on the homologous human framework. Their approach of employing homologous human frameworks with putative murine contact residues resulted in humanized antibodies with similar binding affinities to the original murine antibody with respect to antibodies specific for the interleukin 2 receptor (Queen et al., 1989 [17]) and antibodies specific for herpes simplex virus (HSV) (Co. However, the reintroduction of et al., 1991 [19]). murine residues into human frameworks (at least 9 for anti-interleukin 2 receptor antibodies, at least 9 and 7 for each of two anti-HSV antibodies) may increase the prospect of HAMA response to the framework region in the humanized antibody. Bruggemann et al., 1989 [20] have V region frameworks human demonstrated that recognized as foreign in mouse, and so, conversely, murine modified human frameworks might give rise to an immune reaction in humans.

According to the above described two step approach in WO 90/07861 [18], Queen et al. outlined four criteria for designing humanized immunoglobulins. The first criterion is to use as the human acceptor the framework from a particular human immunoglobulin that is usually homologous to the non-human donor immunoglobulin to be humanized, or to use a consensus framework from many human antibodies. The second criterion is to use the donor amino acid rather than the acceptor if the human acc ptor residue is unusual and the d nor residue is typical for human sequences at a specific residue of the

WO 94/16094 PCT/US94/00266

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framework. The third criterion is to use the donor framework amin acid residue rather than the acceptor at positions immediately adjacent to the CDRs. The fourth criterion is to use the donor amino acid residue at framework positions at which the amino acid is predicted to have a side chain atom within about 3 Å of the CDRs in a three-dimensional immunoglobulin model and to be capable of interacting with the antigen or with the CDRs of the humanized immunoglobulin. It is proposed that criteria two, three or four may be applied in addition or alternatively to criterion one, or each criteria may be applied singly or in any combination.

addition. In WO 90/07861 [18] details the preparation of a single CDR-grafted humanized antibody, a humanized antibody specificity for the p55 Tac protein of the IL-2 receptor, by employing the combination of all four criteria, as above, in designing this humanized antibody. The variable region frameworks of the human antibody EU (see, Kabat et al., 1991 [15]) were used as acceptor. In the resultant humanized antibody, the donor CDRs were as defined by Kabat et al., 1991 [15] and Wu et al., 1970 [16] and, in addition, the mouse donor residues were used in place of the human acceptor residues, at positions 27, 30, 48, 66, 67, 89, 91, 94, 103, 104, 105 and 107 in heavy chain and at positions 48, 60 and 63 in the light chain, of the variable region frameworks. The humanized anti-Tac antibody obtained was reported to have an affinity for p55 of 3 x 109 M1, about one-third of that of the murine MAb.

Several other groups have demonstrated that Queen et al.'s approach of first choosing homologous frameworks followed by reintroduction of mouse residues may not be necessary to achieve humanized antibodies with similar binding affinities to the original mouse antibodies (Riechmann et al., 1988 [4]; Tempest t al., 1991 [5]; Verhoeyen, et al. 1991 [21]). Moreover, these groups

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have used a different approach and have demonstrated that it is possible to utilize, as standard, the V region frameworks derived from NEWM and REI heavy and light chains respectively for CDR-grafting without radical introduction of mouse residues. However, the determination which mouse residues should of introduced to produce antibodies with binding efficiencies similar to the original murine MAb can be difficult to predict, being largely empirical and not taught by available prior art. In the case of the humanized CAMPATH-IH antibody, the substitution of a phenylalanine for a serine residue at position 27 was the only substitution required to achieve а binding efficiency similar to that of the original murine antibody (Riechmann, et al., 1988 [4]; WO92/04381 [22]). In the case of a humanized (reshaped) antibody specific for respiratory syncytial virus (RSV) for the inhibition of RSV infection in vivo, substitution of a block of 3 residues adjacent to CDR3 in the CDR-grafted NEWM heavy chain was required to produce biological activity equivalent to the original mouse antibody (Tempest et al., 1991 [5]; WO 92/04381 [22]). The reshaped antibody in which only the mouse CDRs were transferred to the human framework showed poor binding for RSV. advantage of using the Tempest et al., 1991 [5] approach to construct NEWM and REI based humanized antibodies is that the 3-dimensional structures of NEWM and REI variable regions are known from x-ray crystallography and thus specific interactions between CDRs and V region framework residues can be modelled.

Regardless of the approach taken, the examples of the initial humanized antibodies prepared to date have shown that it is not a straightforward process to obtain humanized antibodies with the characteristics, in particular, the binding affinity, as well as other desirable properties, of the original murine MAD from

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which the humanized antib dy is derived. Regardless of the approach to CDR grafting taken, it is often not sufficient merely to graft the CDRs from a donor Ig onto the framework regions of an acceptor Ig (see, e.g., Tempest et al., 1991 [5], Riechmann et al., 1988 [4], etc., cited herein). In a number of cases, it appears to be critical to alter residues in the framework regions of the acceptor antibody in order to obtain binding activity. However, even acknowledging that framework changes may be necessary, it is not possible to predict, on the basis of the available prior art, which, if any, framework residues will need to be altered to obtain functional humanized recombinant antibodies of the desired specificity. Results thus far indicate that changes necessary to preserve specificity and/or affinity are for the most part unique to a given antibody and cannot be predicted based on the humanization of a different antibody.

In particular, the sets of residues in the framework region which are herein disclosed as being of critical importance to the activity of the recombinant humanized anti-VLA4 antibodies constructed in accordance with the teachings of the present invention do not generally coincide with residues previously identified as critical to the activity of other humanized antibodies and were not discovered based on the prior art.

# C. Therapeutic Applications of Humanized Antibodies

To date, humanized recombinant antibodies have been developed mainly for therapeutic application in acute disease situations (Tempest, et al., 1991 [5]) or for diagnostic imaging (Verhoeyen, et al., 1991 [21]). Recently, clinical studies have begun with at least two humanized antibodies with NEWM and REI V region frameworks, CAMPATH-IH (Riechmann et al., 1988 [4]) and

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humanized anti-placental alkaline phosphatase (PLAP) (Verhoeyen et al., 1991 [21]) and these studies have initially indicated the absence of any marked immune reaction to these antibodies. A course of treatment with CAMPATH-IH provided remission for two patients with non-Hodgkins lymphoma thus demonstrating efficacy in a chronic disease situation (Hale et al., 1988 [23]). In addition, the lack of immunogeneicity of CAMPATH-IH was demonstrated after daily treatment of the two patients for 30 and 43 days. Since good tolerance to humanized antibodies has been initially observed with CAMPATH-IH, treatment with humanized antibody holds promise for the prevention of acute disease and to treatment of diseases with low mortality.

# D. The VCAM-VLA4 Adhesion Pathway and Antibodies to VLA4

Vascular endothelial cells constitute the lining of blood vessels and normally exhibit a low affinity for circulating leukocytes (Harlan, 1985 [24]). The release of cytokines at sites of inflammation, and in response to immune reactions, causes their activation and results in the increased expression of a host of surface antigens. (Collins et al., 1986 [25]; Pober et al., 1986 [26]; Bevilacqua et al., 1987 [27]; Leeuwenberg et al., 1989 [28]). These include the adhesion proteins ELAM-1, which binds neutrophils (Bevilacqua et al., 1989 [29], ICAM-1 which interacts with all leukocytes (Dustin et al., 1986 [30]; Pober et al. 1986, [26]; Boyd et al., 1988 [31]; Dustin and Springer, 1988 [32]), and VCAM-1 which binds lymphocytes (Osborn et al., 1989 [33]). These cytokineinduced adhesion molecules appear to play an important role in leukocyte recruitment to extravascular tissues.

The integrins are a group of cell-extracellular matrix and cell-cell adhesion receptors exhibiting an alpha-beta heterodimeric structure, with a widespread cell distribution and a high degree of c nservation throughout evolution (Hynes, 1987 [34]; Marcantonio and

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Hynes, 1988 [35]). The integrins have been subdivided into three major subgroups; the  $\beta_2$  subfamily of integrins (LFA-1, Mac-1, and p150,95) is mostly involved in cellcell interactions within the immune system (Kishimoto et al., 1989 [36]), whereas members of the  $\beta_1$  and  $\beta_3$  integrin subfamilies predominantly mediate cell attachment to the extracellular matrix (Hynes, 1987 [34]; Ruoslahti, 1988 In particular, the  $\beta_1$  integrin family, also termed VLA proteins, includes at least six receptors that specifically interact with fibronectin, collagen, and/or laminin (Hemler, 1990 [38]). Within the VLA family, VLA4 is atypical because it is mostly restricted to lymphoid and myeloid cells (Hemler et al., 1987 [39]), and indirect evidence had suggested that it might be involved in various cell-cell interactions (Clayberger et al., 1987 [40]; Takada et al., 1989 [41]; Holtzmann et al., 1989 [42]; Bendarczyk and McIntyre, 1990 [43]). addition, VLA4 has been shown to mediate T and B lymphocyte attachment to the heparin II binding fragment of human plasma fibronectin (FN) (Wayner et al., 1989 [44]).

VCAM-1. like ICAM-1, is a member of the immunoglobulin gene superfamily (Osborn et al., [33]). VCAM-1 and VLA4 were demonstrated to be a ligandreceptor pair that allows attachment of lymphocytes to activated endothelium by Elices et al., 1990 [45]. Thus, VLA4 represents a singular example of a  $\beta_1$  integrin receptor participating in both cell-cell and cellextracellular matrix adhesion functions by means of the defined ligands VCAM-1 and FN.

VCAM1 (also known as INCAM-110) was first identified as an adhesion molecule induced on endothelial cells by inflammatory cytokines (TNF and IL-1) and LPS (Rice et al., 1989 [46]; Osborn et al., 1989 [33]). Because VCAM1 binds to cells exhibiting the integrin VLA4  $(\alpha_4\beta_1)$ , including T and B lymphocyts, monocytes, and

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eosinophils, but not neutrophils, it is thought to participate in recruitm nt of these cells from the bloodstream to areas of infection and inflammation (Elices et al, 1990 [45]; Osborn, 1990 [33]). The VCAM1/VLA4 adhesion pathway has been associated with a number of physiological and pathological processes. Although VLA4 is normally restricted to hematopoietic lineages, it is found on melanoma cell lines, and thus it has been suggested that VCAM1 may participate in metastasis of such tumors (Rice et al., 1989 [46]).

In vivo, VCAM1 is found on areas of arterial endothelium representing early atherosclerotic plaques in a rabbit model system (Cybulsky and Gimbrone, 1991 [47]). VCAM1 is also found on follicular dendritic cells in human lymph nodes (Freedman et al., 1990 [48]). It is also present on bone marrow stromal cells in the mouse (Miyake et al., 1991 [49]), thus VCAM1 appears to play a role in B-cell development.

The major form of VCAM1 in vivo on endothelial cells, has been referred to as VCAM-7D, and has seven Ig 20 homology units or domains; domains 4, 5 and 6 are similar amino acid sequence to domains 1, 2 respectively, suggesting an intergenic duplication event in the evolutionary history of the gene (Osborn et al., 25 1989 [33]; Polte et al. 1990 [50]; Hession et al., 1991 [51]; Osborn and Benjamin, U.S. Ser. No. 07/821,712 filed September 30, 1991, [52]). A 6-domain form (referred to as VCAM-6D herein) is generated by alternative splicing, in which the fourth domain is deleted (Osborn et al., 1989 [33]; Hession et al. 1991 [51], Cybulsky et al., 30 1991 [47]; Osborn and Benjamin, U.S. Ser. No. 07/821,712 filed September 30, 1991 [52]). The VCAM-6D, was the first sequenced of these alternate forms, however, later in vivo studies showed that the VCAM-7D form was dominant 35 in vivo. The biological significance of the alternate splicing is not known, however as shown by Osb rn and

WO 94/16094 . PCT/US94/00266

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Benjamin, U.S. Ser. No. 07/821,712 filed September 30, 1991 [52], VCAM-6D can bind VLA4-expressing cells and thus clearly has potential functionality in vivo.

The apparent involvement of the VCAM1/VLA4 adhesion pathway in infection, inflammation and possibly atherosclerosis has led to continuing intensive research to understand the mechanisms of cell-cell adhesion on a molecular level and has led investigators to propose intervention in this adhesion pathway as a treatment for diseases, particularly inflammation (Osborn et al., 1989 [33]). One method of intervention in this pathway could involve the use of anti-VLA4 antibodies.

Monoclonal antibodies that inhibit VCAM1 binding to VLA4 are known. For example, anti-VLA4 MAbs HP2/1 and HP1/3 have been shown to block attachment of VLA4-expressing Ramos cells to human umbilical vein cells and VCAM1-transfected COS cells (Elices et al., 1990 [45]). Also, anti-VCAM1 antibodies such as the monoclonal antibody 4B9 (Carlos et al., 1990 [53]) have been shown to inhibit adhesion of Ramos (B-cell-like), Jurkat (T-cell-like) and HL60 (granulocyte-like) cells to COS cells transfected to express VCAM-6D and VCAM-7D (Hession et al., 1991 [51]).

The monoclonal antibodies to VLA4 that have been described to date fall into several categories based on epitope mapping studies (Pulido, et al., 1991 [54]). Importantly one particular group of antibodies, t epitope "B", are effective blockers of all VLA4-dependent adhesive functions (Pulido et al., 1991, [54]). The preparation of such monoclonal antibodies to epitope B of VLA 4, including, for example the HP1/2 MAb, have been described by Sanchez-Madrid et al., 1986, [55]. Antibodies having similar specificity and having high binding affinities to VLA4 comparable to that of HP1/2, would be particularly promising candidates f r the

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preparation of humanized recombinant anti-VLA4 antibodies us ful as assay reagents, diagnostics and therapeutics.

stated above, inflammatory leukocytes recruited to sites of inflammation by cell adhesion molecules that expressed are on the surface cells and which act as receptors leukocyte surface proteins or protein complexes. In particular, eosinophils have recently been found to participate in three distinct cell adhesion pathways to vascular endothelium, binding to cells intercellular adhesion molecule-1 (ICAM-1), endothelial cell adhesion molecule-1 (ELAM-1), and vascular cell adhesion molecule-1 (VCAM-1) (Weller et al., 1991 [56]; Walsh et al., 1991 [57]; Bochner et al., 1991 [58]; and Dobrina et al., 1991 [59]). That eosinophils express VLA4 differentiates them from other inflammatory cells such as neutrophils, which bind to ELAM-1 and ICAM-1 but not VCAM-1.

VLA4-mediated adhesion pathway investigated in an asthma model to examine the possible role of VLA4 in leukocyte recruitment to inflamed lung tissue (Lobb, U.S. Ser. No. 07/821,768 filed January 13, 1992 [60]). Administering anti-VLA4 antibody inhibited both the late phase response and hyperresponsiveness in allergic sheep. Surprisingly, administration of anti-VLA4 led to a reduction in the number of both neutrophils and eosinophils in the lung at 4 hours after allergen challenge, even though both cells have alternate adhesion pathways by which they can be recruited to lung tissues. Also surprisingly, inhibition of hyperresponsiveness in the treated sheep was observed which continued to 1 week, even though infiltration of leukocytes, including neutrophils and eosinophils, was not significantly reduced over time.

The VLA4-mediated adhesion model has also been inv stigated in a primate mod 1 of inflammatory bowel

WO 94/16094

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disease (IBD) (Lobb, U.S. Ser. No, 07/835,139 filed February 12, 1992 [61]). The administration of anti-VLA4 antibody surprisingly and significantly reduced acute inflammation in that model, which is comparable to ulcerative colitis in humans.

More recently, anti-VLA4 antibodies have been used in methods for the peripheralizing of CD34<sup>+</sup> cells, including hematopoietic stem cells as described in Papyannopoulou, U.S. Ser. No. 07/977,702, filed November 13, 1992 [62].

Thus, anti-VLA4 antibodies having certain epitopic specificities and certain binding affinities may be therapeutically useful in a variety of inflammatory conditions, including asthma and IBD. In particular, humanized recombinant versions of such anti-VLA4 antibodies, if they could be constructed, might be especially useful for administration in humans. humanized antibodies would have the desired potency and specificity, while avoiding or minimizing immunological response which would render the antibody ineffective and/or give rise to undesirable side effects.

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### SUMMARY F THE INVENTION

The present inventi n provides a method of constructing a recombinant anti-VLA4 antibody molecule. Specifically, recombinant antibodies according to the present invention comprise the antigen binding regions derived from the heavy and/or light chain variable regions of an anti-VLA4 antibody.

The present invention provides a method for the construction of humanized recombinant antibody molecule using as a first step CDR grafting or "reshaping" technology. Specifically, the humanized antibodies according to the present invention have specificity for VLA4 and have an antigen binding site wherein at least one or more of the complementarity determining regions (CDRs) of the variable domains are derived from a donor non-human anti-VLA4 antibody, and in which there may or may not have been minimal alteration of the acceptor antibody heavy and/or light variable framework region in order to retain donor antibody binding specificity. Preferably, the antigen binding regions of the CDRgrafted heavy chain variable domain comprise the CDRs corresponding to positions 31-35 (CDR1), 50-65 (CDR2) and 95-102 (CDR3). Preferably, the antigen binding regions of the CDR-grafted light chain variable domain comprise CDRs corresponding to positions 24-34 (CDR1), (CDR2) and 89-97 (CDR3). These residue designations are numbered according to the Kabat numbering (Kabat et al., 1991 [15]). Thus, the residue/position designations do not always correspond directly with the linear numbering of the amino acid residues shown in the sequence listing. In the case of the humanized  $V_{\boldsymbol{K}}$  sequence disclosed herein, the Kabat numbering does actually correspond to the linear numbering of amino acid residues shown in the sequence listing. In contrast, in the case of the humanized  $V_H$  sequences disclosed herein, the Kabat numbering does not correspond to the linear numbering of

PCT/US94/00266

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amino acid residues shown in the sequence listing (e.g., f r the humanized  $V_{\rm H}$  regi ns disclosed in the sequence listing, CDR2 = 50-66, CDR3 = 99-110).

The invention further provides the recombinant and humanized anti-VLA4 antibodies which may be detectably labelled.

The invention additionally provides a recombinant DNA molecule capable of expressing the recombinant and humanized anti-VLA4 antibodies of the present invention.

The invention further provides host cells capable of producing the recombinant and humanized anti-VLA4 antibodies of the present invention.

The invention additionally relates to diagnostic and therapeutic uses for the recombinant and humanized anti-VLA4 antibodies of the present invention.

The invention further provides a method for treating inflammation resulting from a response of the specific defense system in a mammalian subject, including humans, which comprises providing to a subject in need of such treatment an amount of an anti-inflammatory agent sufficient to suppress the inflammation wherein the anti-inflammatory agent is a recombinant and humanized anti-VLA4 antibody of the present invention.

The invention further provides a method for treating non-specific inflammation in a mammalian subject, including humans using the recombinant and humanized anti-VLA4 antibodies.

The invention further concerns the embodiment of the above-described methods wherein the recombinant and humanized anti-VLA4 antibodies of the present invention are derived from the murine monoclonal antibody HP1/2.

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# DETAILED DESCRIPTI N OF SPECIFIC EMBODIMENTS OF THE INVENTION

The technology for producing monoclonal antibodies is well known. Briefly, an immortal cell line (typically myeloma cells) is fused to lymphocytes (typically splenocytes) from a mammal immunized with whole cells expressing a given antigen, e.g., VLA4, and the culture supernatants of the resulting hybridoma cells are screened for antibodies against the antigen (see, generally, Kohler et al., 1975 [1]).

Immunization may be accomplished using standard procedures. The unit dose and immunization regimen depend on the species of mammal immunized, its immune status, the body weight of the mammal, etc. Typically, the immunized mammals are bled and the serum from each blood sample is assayed for particular antibodies using appropriate screening assays. For example, anti-VLA4 antibodies may be identified by immunoprecipitation of 125I-labeled cell lysates from VLA4-expressing cells (see, Sanchez-Madrid et al., 1986 [55] and Hemler et al., 1987 [39]). Anti-VLA-4 antibodies may also be identified by flow cytometry, e.g., by measuring fluorescent staining of Ramos cells incubated with an antibody believed to recognize VLA4 (see, Elices et al., 1990 The lymphocytes used in the production of hybridoma cells typically are isolated from immunized mammals whose sera have already tested positive for the presence of anti-VLA4 antibodies using such screening assays.

Typically, the immortal cell line (e.g., a myeloma cell line) is derived from the same mammalian species as the lymphocytes. Preferred immortal cell lines are mouse myeloma cell lines that are sensitive to culture medium containing hypoxanthine, aminopterin and thymidine ("HAT medium").

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Typically, HAT-sensitive m use myeloma cells are fused t mous splenocytes using 1500 m lecular weight polyethylene glycol ("PEG 1500"). Hybridoma cells resulting from the fusion are then selected using HAT medium, which kills unfused and unproductively fused myeloma cells (unfused splenocytes die after several days because they are not transformed). Hybridomas producing a desired antibody are detected by screening the hybridoma culture supernatants. For example, hybridomas prepared to produce anti-VLA4 antibodies may be screened by testing the hybridoma culture supernatant for secreted antibodies having the ability to bind to a recombinant  $\alpha_4$ -subunit-expressing cell line, such as transfected K-562 cells (see, e.g., Elices et al., 1990 [45]).

To produce anti VLA4-antibodies, hybridoma cells that tested positive in such screening assays are cultured in a nutrient medium under conditions and for a time sufficient to allow the hybridoma cells to secrete the monoclonal antibodies into the culture medium. Tissue culture techniques and culture media suitable for hybridoma cells are well known. The conditioned hybridoma culture supernatant may be collected and the anti-VLA4 antibodies optionally further purified by well-known methods.

Alternatively, the desired antibody may be produced by injecting the hybridoma cells into the peritoneal cavity of an unimmunized mouse. The hybridoma cells proliferate in the peritoneal cavity, secreting the antibody which accumulates as ascites fluid. The antibody may be harvested by withdrawing the ascites fluid from the peritoneal cavity with a syringe.

Several anti-VLA4 monoclonal antibodies have been previously described (see, e.g., Sanchez-Madrid et al., 1986 [55]; Hemler et al., 1987 [39]; Pulido et al., 1991 [54]). HP1/2, for example, is one such murine monoclonal antibody which recognizes VLA4. VLA4 acts as a leukocyte

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receptor for plasma fibr nectin and VCAM-1. Other mon clonal antibodies, such as HP2/1, HP2/4, L25 and P4C2, have been described that also recognize VLA4.

Recombinant antibodies have been constructed and are described herein in which the CDRs of the variable domains of both heavy and light chains were derived from the murine HP1/2 sequence. Preferred starting materials constructing recombinant humanized according to the present invention antibodies are anti-VLA4 antibodies, such as HP1/2, that block the interaction of VLA4 with both VCAM1 and fibronectin. preferred are those antibodies, such as HP1/2, which in Particularly addition, do not cause cell aggregation. Some anti-VLA4 blocking antibodies have been observed to cause such aggregation. The HP1/2 MAb (Sanchez-Madrid et al., 1986 a particularly excellent humanization since it has an extremely high potency, candidate blocks VLA4 interaction with both VCAM1 and fibronectin, but does not cause cell aggregation, and has specificity for epitope B on VLA4. In the initial experiments,  $V_{\text{H}}$  and  $V_{\text{K}}$  DNA were isolated and cloned from an HP1/2-producing hybridoma cell line. The variable domain frameworks and constant domains for humanization were initially derived from human antibody sequences.

The three CDRs that lie on both heavy and light chains are composed of those residues which structural studies have shown to be involved in antigen binding. Theoretically, if the CDRs of the murine HP1/2 antibody were grafted onto human frameworks to form a CDR-grafted variable domain, and this variable domain were attached to human constant domains, the resulting CDR-grafted antibody would essentially be a human antibody with the specificity of murine HP1/2 to bind human VLA4. Given the highly "human" nature of this antibody, it would be expected to be far less immunogenic than murine HP1/2 when administered to patients.

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However, following testing f r antigen binding of a CDR-grafted HP1/2 antibody in which only the CDRs were grafted onto the human framework, it was shown that this did not produce a CDR-grafted antibody having reasonable affinity for the VLA4 antigen. It was therefore decided that additional residues adjacent to some of the CDRs and critical framework residues needed to be substituted from the human to the corresponding murine HP1/2 residues in order to generate an antibody with binding affinity in the range of 10% to 100% of the binding affinity of the murine HP1/2 MAb. Empirically, changes of one or more residues in the framework regions of  $V_H$  and  $V_V$  were made to prepare antibodies of the desired specificity and potency, but without making so many changes in the human framework so as to compromise the essentially human nature of the humanized  $V_{\text{H}}$  and  $V_{\text{K}}$  region sequences.

Furthermore, VLA4-binding fragments may be prepared from the recombinant anti-VLA4 antibodies described herein, such as Fab, Fab', F(ab')2, and F(v) fragments; heavy chain monomers or dimers; light chain monomers or dimers; and dimers consisting of one heavy chain and one light chain are also contemplated herein. Such antibody fragments may be produced by chemical methods, e.g., by cleaving an intact antibody with a protease, such as pepsin or papain, or via recombinant DNA techniques, e.g., by using host cells transformed with truncated heavy and/or light chain genes. Heavy and light chain monomers may similarly be produced by treating an intact antibody with a reducing agent such as dithiothreitol or  $\beta$ -mercaptoethanol or by using host cells transformed with DNA encoding either the desired heavy chain or light chain or both.

WO 94/16094 PCT/US94/00266

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-23-

The following examples ar intended to further illustrate certain preferred embodim nts of the invention and are not intended to be limiting in nature. In the following examples, the necessary restriction enzymes, plasmids, and other reagents and materials may be obtained from commercial sources and cloning, ligation and other recombinant DNA methodology may be performed by procedures well-known in the art.

#### Example 1

# Isolation of DNA Sequenc s Encoding Murine Anti-VLA4 Variable Regions

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A. Isolation of the HP1/2 heavy and light chain cDNA

To design a humanized recombinant antibody with
specificity for VLA4, it was first necessary to determine
the sequence of the variable domain of the murine HP1/2
heavy and light chains. The sequence was determined from
heavy and light chain cDNA that had been synthesized from
cytoplasmic RNA according to methods referenced in
Tempest et al., 1991 [5].

### Cells and RNA isolation

Cytoplasmic RNA (~200  $\mu$ g) was prepared by the method of Favaloro et al., 1980 [63], from a semi-confluent 150cm² flask of HP1/2-producing hybridoma cells (about 5 X 10<sup>5</sup> logarithmic phase cells). The cells were pelleted and the supernatant was assayed for the presence of antibody by a solid phase ELISA using an Inno-Lia mouse monoclonal antibody isotyping kit (Innogenetics, Antwerp, Belgium) using both the kappa conjugate and the lambda conjugate. The antibody was confirmed to be  $IgGl/\kappa$  by this method.

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#### 2. <u>cDNA Synthesis</u>

cDNAs were synthesized from the HP1/2 RNA via reverse transcription initiated from primers based on the 5' end of either the murine IgG1 CH<sub>1</sub> or the murine kappa constant domains using approximately 5  $\mu$ g RNA and 25 pmol primer in reverse transcriptase buffer containing 1  $\mu$ 1/50  $\mu$ 1 Pharmacia (Milton Keynes, United Kingdom) RNA Guard<sup>TM</sup> and 250 micromolar dNTPs. The sequence of these primers, CG1FOR and CK2FOR are shown as SEQ ID NO: 1 and SEQ ID NO: 2, respectively. The mixture was heated to 70°C, then allowed to cool slowly to room temperature. Then, 100 units/50  $\mu$ 1 MMLV reverse transcriptase (Life Technologies, Paisley, United Kingdom) was added and the reaction was allowed to proceed at 42°C for one hour.

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#### 3. Amplification of VH and Vx cDNA

Polymerase chain reaction (PCR) of murine MAb variable regions can be achieved using a variety of procedures, for example, anchored PCR or primers based on conserved sequences (see, e.g., Orlandi et al., 1989 [64]). Orlandi et al. [64], Huse et al., 1989 [65] and Jones and Bendig, 1991 [66], have described some variable region primers. We have been unsuccessful, however, in using a number of such primers, particularly those for the light chain PCR of HP1/2 derived  $V_{\rm K}$  sequences.

HP1/2 Ig  $V_H$  and  $V_K$  cDNAs were amplified by PCR as described by Saiki et al., 1988 [67] and Orlandi et al., 1989 [64]. Reactions were carried out using 2.5 units/50 µl Amplitaq™ polymerase (Perkin Elmer Cetus, Norwalk, CT) in 25 cycles of 94°C for 30 seconds followed by 55°C for 30 seconds and 75°C for 45 seconds. The final cycle was followed by five minute incubation at 75°C. The same 3' oligonucleotides used for cDNA synthesis were used in conjunction with appropriate 5' oligonucleotides based on consensus sequences of relatively conserved regions at the 5' end of each V region. V<sub>H</sub> cDNA was successfully amplified using the primers VH1BACK (SEQ ID 3] and CG1FOR [SEQ ID NO: 1] and yielded an amplification product of approximately 400 bp. Vr cDNA was successfully amplified using the primers VK5BACK [SEQ ID NO: 4] and CK2FOR [SEQ ID NO: 2] and yielded an amplification product of approximately 380 bp.

#### 4. Cloning and Sequencing V, DNA

The primers used for the amplification of  $V_H$  DNA, contain the restriction enzyme sites <u>PstI</u> and <u>HindIII</u> which facilitate cloning into sequencing vectors. The general cloning and ligation methodology was as described in <u>Molecular Cloning</u>, <u>A Laboratory Manual</u> 1982, [68]. The

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collection

separate clones.

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PCT/US94/00266 -26amplified DNA was digested with PstI to check for internal PstI sites and an internal PstI site was found. Therefore, the V<sub>H</sub> DNA was cloned as PstI-PstI and PstI-HindIII fragments into M13mp18 and 19. The resulting from two independent CDNA preparations were sequenced by the dideoxy method (Sanger, et al., 1977, [69] using Sequenase™ (United States Biochemicals, Cleveland, Ohio, USA). The sequence of a region of ~100-250 bp was determined from each of 25 Out of more than 4000 nucleotides sequenced, there were three PCR-induced transition mutation in three The HP1/2  $V_H$  DNA sequence and its translated amino acid sequence are set forth in SEQ ID NO: 5 and SEQ ID NO: 6, respectively. It should be noted that the first eight amino acids are dictated by the 5' primer used in the PCR. Computer-assisted comparisons indicate that HP1/2  $V_H$  [SEQ ID NOS: 5 and 6] is a member of family IIC (Kabat et al., 1991, [15]. A comparison between HP1/2  $V_{\rm H}$  [SEQ ID NOS: 5 and 6] and a consensus sequence of family IIC revealed that the only unusual residues are at amino acid positions 80, 98 and 121 (79, 94 and 121 in Kabat numbering). Although Tyr 80 is invariant in subgroup IIC other sequenced murine  $V_{\rm H}$ regions have other aromatic amino acids at this position

although none have Trp. The majority of human and murine V<sub>H</sub>s have an arginine residue at Kabat position 94. presence of Asp 94 in HP1/2  $V_{\rm H}$  is extremely rare; there is only one reported example of a negatively charged residue at this position. Proline at Kabat position 113 is also unusual but is unlikely to be important in the conformation of the CDRs because of its distance from The amino acids making up CDR1 have been found in three other sequenced murine  $V_{\text{H}}$  regions. However, CDR2 and CDR3 are unique to HP1/2 and are not found in any other reported murine V<sub>N</sub>.

WO 94/16094 PCT/US94/00266

#### 5. Cloning and Sequencing V. DNA

The primers used for the amplification of V<sub>K</sub> DNA contain restriction sites for the enzymes <u>Eco</u>RI and <u>HindIII</u>. The PCR products obtained using primers VK1BACK [SEQ ID NO: 7], VK5BACK [SEQ ID NO: 4] and VK7BACK [SEQ ID NO: 8] were purified and cloned into M13. Authentic kappa sequences were obtained only with VK5BACK [SEQ ID NO: 4]. The sequence of a region of ~200-350 bp was determined by the dideoxy method (Sanger et al., 1977, [69] using Sequenase<sup>TM</sup> (United States Biochemicals, Cleveland, Ohio, USA) from each of ten clones from two independent cDNA preparations. Out of more than 2 kb sequenced, there were only two clones which each contained one PCR-induced transition mutation.

The HP1/2  $V_K$  DNA sequence and its translated amino acid sequence are set forth in SEQ ID NO: 9 and SEQ ID NO: 10, respectively. The first four amino acids are dictated by the 5' PCR primer but the rest of the sequence is in total agreement with partial protein sequence data. HP1/2  $V_K$  is a member of Kabat family V (Kabat et al., 1991 [15]) and has no unusual residues. The amino acids of CDR1 and CDR3 are unique. The amino acids making up CDR2 have been reported in one other murine  $V_K$ .

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#### Example 2

#### Design of a CDR-grafted Anti-VLA4 Antibody

To design a CDR-grafted anti-VLA4 antibody, it was necessary to determine which residues of murine HP1/2 comprise the CDRs of the light and heavy chains.

Three regions of hypervariability amid the less variable framework sequences are found on both light and heavy chains (Wu and Kabat, 1970 [16]; Kabat et al., 1991 [15]). In most cases these hypervariable regions correspond to, but may extend beyond, the CDR. The amino acid sequences of the murine  $HP1/2 \ V_H$  and  $V_r$  chains are

PCT/US94/00266

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set forth in SEQ ID NO: 6 and SEQ ID NO: 10, resp ctiv ly. CDRs of murine HP1/2 were elucidated in accordance with Kabat et al., 1991 [15] by alignment with other  $V_H$  and  $V_K$  sequences. The CDRs of murine HP1/2  $V_H$  were identified and correspond to the residues identified in the humanized  $V_H$  sequences disclosed herein as follows:

CDR1  $AA_{31}$ - $AA_{35}$ CDR2  $AA_{50}$ - $AA_{66}$ CDR3  $AA_{90}$ - $AA_{110}$ 

These correspond to  $AA_{31}$ - $AA_{35}$ ,  $AA_{50}$ - $AA_{65}$ , and  $AA_{95}$ - $AA_{102}$ , respectively, in Kabat numbering. The CDRs of murine HP1/2  $V_K$  were identified and correspond to the residues identified in the humanized  $V_K$  sequences disclosed herein as follows:

CDR1 AA<sub>24</sub>-AA<sub>34</sub>
CDR2 AA<sub>50</sub>-AA<sub>56</sub>
CDR3 AA<sub>89</sub>-AA<sub>97</sub>

These correspond to the same numbered amino acids in Kabat numbering. Thus, only the boundaries of the  $V_K$ , but not  $V_H$ , CDRs corresponded to the Kabat CDR residues. The human frameworks chosen to accept the HP1/2 CDRs were NEWM and REI for the heavy and light chains respectively. The NEWM and the REI sequences have been published in Kabat et al., 1991 [15].

An initial stage of the humanization process may comprise the basic CDR grafting with a minimal framework change that might be predicted from the literature. For example, in Riechmann et al., 1988 [4], the MAB CAMPATH-1H was successfully humanized using direct CDR grafting with only one framework change necessary to obtain an antibody with a binding efficiency similar to that of the original murine antibody. This framework change was the substitution of a Phe for a Ser at position 27. However, using the same humanization strategy by CDR grafting and the single framework change disc vered by Riechmann et

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al., 1988 [4] for the preparation of humanized antibodies having other specificities did not yield antibodies with affinities comparable to the murine antibodies from which they were derived. In such cases, the humanization process must necessarily include additional empirical changes to achieve the desired specificity and potency. Such changes may be related to the unique structure and sequence of the starting murine antibody but are not predictable based upon other antibodies of different specificity and sequence. For example, analysis of the murine  $V_{H}$  amino acid sequence from HP1/2 as set forth in SEQ ID NO: 6 as compared with the other known sequences indicated that residues 79, 94 and 113 (Kabat numbering) Of these, only Asp 94 is likely to be were unusual. important in CDR conformation. Most  $\mathbf{V}_{H}$  regions that have been sequenced have an arginine at this position which is able to form a salt bridge with a relatively conserved Asp 101 in CDR3. Because NEWM has an Arg 94 and  $V_{\rm H}$  CDR3 of HP1/2 has an Asp 101, there remains the possibility that a salt bridge would form which would not normally occur. The presence of a negatively charged residue at position 94 is very unusual and therefore it was decided to include the Asp 94 into the putative humanized  $V_{\rm H}$ .

A chimeric (murine V/human IgGl/ $\kappa$ ) HP1/2 antibody may be useful, but not a necessary, intermediate in the initial stages of preparing a CDR grafted construct because (i) its antigen-binding ability may indicate that the correct V regions have been cloned; and (ii) it may act as a useful control in assays of the various humanized antibodies prepared in accordance with the present invention.

For  $V_H$ , an M13 clone containing full-length HP1/2  $V_H$  was amplified using VH1BACK [SEQ ID NO: 3] and VH1FOR [SEQ ID NO: 11] which contain PstI and BstEII sites respectively at the 5' and 3' ends of the  $V_H$  domain. The amplified DNA was cut with BstEII and partially cut with

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PstI, full-1 ngth DNA purified and cl ned into M13VHPCR1 (Orlandi et al., 1989 [64]) which had been cut with PstI and BstEII. For  $V_K$  an M13 clone containing full-length HP1/2  $V_K$  was amplified using VK3BACK [SEQ ID NO: 12] and VK1FOR [SEQ ID NO: 13] to introduce PvuII and BglII sites respectively at the 5' and 3' ends of the  $V_K$  domain. The amplified DNA was cut with PvuII and BglII and cloned into M13VKPCR1 (Orlandi et al., 1989 [64]) which had been cut with PvuII and BclI.

In sum, the 5' primers used for the amplification of the murine  $V_H$  and  $V_K$  regions contain convenient restriction sites for cloning into our expression vectors. The 3' primers used in the PCRs were from the constant regions. Restriction sites at the 3' end of the variable regions were introduced into cloned murine variable region genes with PCR primers which introduced BstII or BglII sites in the heavy and light (kappa) variable regions, respectively. Additionally, the  $V_H$  primer changed Pro 113 to Ser.

The murine  $V_R$  and  $V_K$  DNAs were cloned into vectors containing the gpt and hygromycin resistance genes respectively, such as pSVgpt and pSVhyg as described by Orlandi, et al. [64], and appropriate human IgGl, IgG4 or k constant regions were added, for example, as described by Takahashi et al., 1982 [70], Flanagan and Rabbitts, 1982 [71], and Hieter et al., 1980 [72], respectively. The vectors were cotransfected into the rat myeloma YB2/0. and mycophenolic acid resistant clones screened by ELISA for secretion of chimeric  $IgG/\kappa$  antibody. The YB2/0 cell line was described by Kilmartin et al., 1982 [73] and is available from the American Type Culture Collection (ATCC, Rockville, MD). ELISA positive clones were expanded and antibody purified from culture medium by protein A affinity chromatography. The chimeric antibody purified from the transfected cells was assayed for anti-

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VLA4 antibody activity as described in Example 7 and was found to be equipotent with the murine HP1/2 antibody.

#### Example 3

Transplantation of CDR Sequences and Mutagenesis of Selected Framework Residues

Transplantation of the CDRs into human frameworks was performed using M13 mutagenesis vectors. The human frameworks chosen to accept the CDR sequences outlined in Example 2 were derived from NEWM for  $V_H$  and REI for  $V_K$ , each in an M13 mutagenesis vector. The M13 mutagenesis vectors used for  $V_H$  and  $V_K$ , were M13VHPCR1 and M13VKPCR2, M13VKPCR2 is identical to M13VKPCR1 as respectively. described by Orlandi et al., 1989 [64], except for a single amino acid change from valine (GTG) to glutamine (GAA) in framework 4 of the REI  $V_{K}$  coding sequence. M13VHPCR1 described by Orlandi et al., 1989 [64] is M13 that contains the coding sequence for a  $V_{\rm H}$  region that is an NEWM framework sequence with CDRs derived from an anti-hapten (4-hydroxy-3-nitrophenyl acetyl caproic acid) antibody; the irrelevant VH CDRs are replaced by sitedirected mutagenesis with the CDRs derived from HP1/2  $\rm V_{H}$ as described below. The  $V_H$  region sequence (DNA and amino acid) encoded by M13VHPCR1 is shown as SEQ ID NOS: M13VKPCR2, like M13VKPCR1 described by Orlandi et al. [64], is M13 that contains the coding sequence for a  $V_K$  region that is N-terminal modified REI framework sequence with CDRs derived from an antilysozyme antibody; these irrelevant  $V_K$  CDRs are replaced by site-directed mutagenesis with the CDRs derived from HP1/2  $V_{\mbox{\scriptsize K}}$  as described below. The  $V_{\mbox{\scriptsize K}}$  region sequence (DNA and amino acid) encoded by M13PCR2 is shown as SEQ ID NOS: 16 and 17.

Synthetic oligonucleotides were synthesized containing the HP1/2-derived  $V_H$  and  $V_K$  CDRs flanked by short sequences drawn from NEWM and REI frameworks,

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respectively, and grafted int the human frameworks by olig nucleotide site-dir cted mutagenesis as follows. human V<sub>H</sub> framework, grafting into the mutagenizing oligonucleotides 598 [SEQ ID NO: 18], 599 [SEQ ID NO: 19] and 600 [SEQ ID NO: 20] were used. For the  $v_{\kappa}$ framework, into human grafting mutagenizing oligonucleotides were 605 [SEQ ID NO: 21], 606 [SEQ ID NO: 22] and 607 [SEQ ID NO: 23]. To 5  $\mu g$  of  $V_{H}$  or  $V_{K}$  single-stranded DNA in M13 was added a 2-fold molar excess of each of the three  $V_{H}$  or  $V_{K}$  phosphorylated oligonucleotides together with flanking primers based on M13 sequences, oligo 10 [SEQ ID NO: 24] for  $V_{\rm H}$  and oligo 385 [SEQ ID NO: 25] for  $V_{\kappa}$ . Primers were annealed to the template by heating to 70°C and slowly cooling to 37°C. The annealed DNA was extended and ligated with 2.5 U T7 DNA polymerase (United States Biochemicals) and 1 U T4 DNA ligase (Life Technologies) in 10 mM Tris HCl pH 8.0, 5 mM MgCl<sub>2</sub>, 10 mM DTT, 1 mM ATP, 250  $\mu$ M dNTPs in a reaction volume of 50  $\mu$ l at 16°C for 1-2 hours.

The newly extended mutagenic strand was preferentially amplified using 1 U Vent DNA polymerase (New England Biolabs) and 25 pmol oligo 11 [SEQ ID NO: 26] or oligo 391 [SEQ ID NO: 27] (for  $V_H$  or  $V_K$ , respectively) in 10 mM KCl, 10 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 20 mM Tris HCl pH 8.8, 2 mM MgSO<sub>4</sub>, 0.1% Triton X-100, 25  $\mu$ M dNTPs in a reaction volume of 50  $\mu$ l and subjecting the sample to 30 cycles of 94°, 30s; 50°, 30s; 75°, 90s.

A normal PCR was then performed by adding 25 pmololigo 10 [SEQ ID NO: 24] (for  $V_{\rm H}$ ) or oligo 385 [SEQ ID NO: 25] (for  $V_{\rm K}$ ) with 10 thermal cycles. The product DNAs were digested with <u>HindIII</u> and <u>Bam</u>HI and cloned into M13mp19. Single-stranded DNA was prepared from individual plaques, sequenced and triple mutants were identified.

The resulting Stage 1  $V_{\text{H}}$  construct with the DNA sequence and its translated product set f rth in SEQ ID

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NO: 28 and SEQ ID NO: 29, respectively. In addition to the CDR grafting, the Stage 1  $V_H$  construct contained selected framework changes. Just prior to CDR1, a block of sequences was changed to the murine residues Phe 27, Asn 28, Ile 29 and Lys 30 [compare  $AA_{27}$ - $AA_{30}$  of SEQ ID NO: 29 with that of murine  $V_H$  sequence [SEQ ID NO: 6]]. This included Phe-27 as substituted in the humanization of the rat CAMPATH1-H antibody (Riechmann et al., 1988 [4]), but then also substitutes the next three residues found in the murine sequence. Although these four residues are nominally included in CDR1 (i.e., hypervariable in the Kabat sense), structurally they are a part of the CDR1 loop (i.e., structural loop residues), and therefore included empirically as part of CDR1. addition, the change from Arg to Asp at residue 94 was made based on the rationale discussed in Example 2. alignment of the CDR-grafted Stage 1 framework sequences as compared with the NEWM framework is shown in Table I. The resulting VK1 (DQL) construct with the DNA sequence and its translated product are set forth in SEQ ID NO: 30 and SEQ ID NO: 31, respectively. An alignment of the CDR-grafted VK1 (DQL) framework sequences as compared with the REI framework is shown in Table II.

The CDR replaced  $V_H$  (Stage 1) and  $V_K$  (VK1) genes were cloned in expression vectors according to Orlandi, et al., 1989 [64] to yield the plasmids termed pHuVHHuIgG1, pHuVHHuIgG4 and pHuVKHuCK. For pHuVHHuIgG1 and pHuVHHuIgG4, the Stage 1  $V_H$  gene together with the Ig heavy chain promoter, appropriate splice sites and signal peptide sequences were excised from the M13 mutagenesis vector by digestion with HindIII and BamHI, and cloned into an expression vector such as pSVgpt as described by Orlandi et al. [64], containing the murine Ig heavy chain enhancer, the SV40 promoter, the gpt gene for selection in mammalian cells and genes for replication and selection in  $\underline{E}$ .  $\underline{Coli}$ . A human IgG1 constant region as

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described in Takahashi et al., 1982 [70] was then added as a <u>Bam</u>HI fragment. Alternatively, a human IgG4 construct region as described by Flanagan and Rabbitts, 1982 [71] is added. The construction of the pHuVKHuCK plasmid, using an expression vector such as pSVhyg as described by Orlandi et al. [64], was essentially the same as that of the heavy chain expression vector except that the gpt gene for selection was replaced by the hygromycin resistance gene (hyg) and a human kappa chain constant region as described by Hieter, 1980, [72] was The vectors were cotransfected into the rat myeloma YB2/0 and mycophenolic acid resistant clones screened by ELISA for secretion of human  $IgG/\kappa$  antibody. The YB2/0 cell line was described by Kilmartin et al., 1982 [73] and is available from the American Type Culture Collection (ATCC, Rockville, MD). ELISA positive clones were expanded and antibody purified from culture medium by protein A affinity chromatography. The transfected cells are assayed for anti-VLA4 antibody activity as described in Example 7.

#### Example 4

## Modification of a CDR grafted Antibody

Beyond the stages of design and preparation to yield anti-VLA4 antibodies as described above in Examples 2 and 25 3, additional stages of empirical modifications were used to successfully prepare humanized recombinant anti-VLA4 antibodies. The Stage 1 modifications as described in Example 3 were based on our analysis of primary sequence 30 and experience in attempting to successfully humanize antibodies. The next modifications, designated as Stage 2, were empirical, based in part on our analysis of 3D modelling data. For the  $V_H$ region, further modifications, designated Stage 3, were so-called "scanning" modifications empirically made to correct any 35 remaining defects in affinities or other antibody

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properties. The modifications that were made in these several stages were empirical changes of various blocks of amino acids with the goal of optimizing the affinity and other desired properties of humanized anti-VLA4 antibodies. Not every modification made during the various stages resulted in antibodies with desired properties.

# 1. Additional heavy chain modifications

### a. Stage 2 Modification

An additional empirical change in the VH framework was made with the use of computer modelling, to generate a Stage 2 construct with the DNA sequence and its translated product set forth in SEQ ID NO: 32 and SEQ ID NO: 33, respectively. Using computer modelling of the Stage 1  $V_H$  region, we determined to make a single change in the framework for Stage 2, namely a substitution of a Ser for Lys at position 75 (Kabat numbering), that is position 76 in SEQ ID NO: 33. This determination was in part based on the possibility that Lys-75 might project into CDR1 and alter its conformation. The M13 vector containing the Stage 1 CDR grafted HuVH, as described in Example 3, was used as template for two-step PCR-directed mutagenesis using the overlap/extension method described by Ho et al., 1989 [74]. In the first step, two separate PCRs were set up, one with an end primer, oligo 10, [SEQ ID NO: 24] and a primer containing the desired mutation, 684 [SEQ ID NO: 34], and the other with the opposite end primer, oligo 11 [SEQ ID NO: 26], and a primer, 683 [SEQ ID NO: 35], that is complementary to the first mutagenic primer. The amplification products of this first pair of PCRs were then mixed together and a second PCR step was carried out using only the end primers oligos 10 and 11, SEQ ID NO: 24 and SEQ ID NO: 26, respectively. The mutagenized amplification product of this PCR was then cloned into M13mp19 and

sequenced, and a mutant bearing the Lys to Ser change (Stage 2 or "S mutant") was identified.

This turned out to be a critical change in the humanized heavy chain derived from HP1/2 (see Example 7). However, this critical change in the preparation of humanized recombinant anti-VLA4 antibodies according to the present invention was not similarly critical in the preparation of other humanized antibodies. Specifically, using the same rationalization and analysis as outlined above, a change in that position was not found to be a beneficial change in the humanization of antibodies of 2 different specificities. An alignment of the CDR-grafted Stage 2 framework sequences as compared with the NEWM, as well as Stage 1 sequences, is shown in Table I.

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# b. Stage 3 Modifications

Additional empirical changes were made as Stage 3 In Stage 3, a series of 5 different block changes of amino acids, for largely empirical reasons, were made to try to improve potency. These constructs are designated STAW, KAITAS, SSE, KRS, and AS. contain the position 75 Ser (Kabat numbering) changed in Stage 2 [position 76 of SEQ ID NO: 35], with other changes as noted. Each of these constructs was prepared mutagenesis using the directed two-step PCR overlap/extension method of Ho et al., 1989 [74], as described for the Stage 2 Ser mutant, above. the additional changes were Gln to Thr at position 77, Phe to Ala at position 78 and Ser to Trp at position 79 (Kabat numbering). These changes were accomplished using end primers, oligos 10 [SEQ ID NO: 24] and 11 [SEQ ID NO: 26] in conjunction with mutagenizing primers 713 [SEQ ID NO: 36] and 716 [SEQ ID NO: 37]. The STAW V<sub>H</sub> DNA sequence and its translated amino acid sequence are set forth in SEQ ID NO: 38 and SEQ ID NO: 39, respectively. KAITAS was prepared with additi nal changes of Arg to Lys

(position 66), Val to Ala (67), Met to Ile (69), Leu to Thr (70) and Val to Ala (71) (Kabat numbering), using oligos 10 [SEQ ID NO: 24] and 11 [SEQ ID NO: 26] in conjunction with oligos 706 [SEQ ID NO: 40] and 707 [SEQ 5 The KAITAS  $V_{\rm H}$  DNA sequence and its 41]. translated amino acid sequence are set forth in SEQ ID 42 and SEQ ID NO: 43, respectively. additional changes of Ala to Ser (84) and Ala to Glu (85) (Kabat numbering), effected by oligos 10 and 11 with 10 oligos 768 [SEQ ID NO: 44] and 769 [SEQ ID NO: 45]. The SSE  $V_{\rm H}$  DNA sequence and its translated amino acid sequence are set forth in SEQ ID NO: 46 and SEQ ID NO: 47, respectively. KRS had additional changes of Arg to Lys (38) and Pro to Arg (40) (Kabat numbering), from oligos 10 [SEQ ID NO: 24] and 11 [SEQ ID NO: 26] with 15 oligos 704 [SEQ ID NO: 48] and 705 [SEQ ID NO: 49]. KRS  $V_H$  DNA sequence and its translated amino acid sequence are set forth in SEQ ID NO: 50 and SEQ ID NO: 51, respectively. AS had additional change Val to Ala at position 24 (Kabat numbering) from oligos 10 [SEQ ID NO: 20 24] and 11 [SEQ ID NO: 26] with oligos 745 [SEQ ID NO: 52] and 746 [SEQ ID NO: 53]. The AS  $V_{\rm H}$  DNA sequence and its translated amino acid sequence are set forth in SEQ ID NO: 54 and SEQ ID NO: 55, respectively. An alignment of the CDR-grafted Stage 3 framework sequences with the 25 NEWM, Stage 0 (see below), Stage 1, and Stage 2 sequences is shown in Table I. Importantly, as shown in Example 7, the potency of STAW and AS humanized antibodies were improved, while KAITAS and KRS humanized antibodies were not of better potency. This could not be predicted. 30

# c. Reverse (Stage 0) Modifications

The two blocks of changes made to generate Stage 1 at positions 28-30 (NIK) and 94 (D) were mutated back to the NEWM sequences at positions 28-30 (TFS), 94 (R), or both positions 27-30 (TFS) and 94 (R). These constructs

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were designated Stage 0-A, 0-B and 0-C, respectively. Each of these constructs was prepared by two-step PCR directed mutagenesis using the overlap/extension method of Ho et al., 1989 [74], as described for the Stage 2 Ser mutant, above. Stage 0-A and 0-B were generated from Stage 1; Stage 0-C was generated from Stage 0-A, as For Stage 0-A, the change was from Asp to Arg This change was accomplished using end at position 94. primers, oligos 10 [SEQ ID NO: 24] and 11 [SEQ ID NO: 26] in conjunction with mutagenizing primers 915 [SEQ ID NO: 56] and 917 [SEQ ID NO: 57]. For stage 0-B, the changes were from Asn-Ile-Lys to Thr-Phe-Ser at positions 28-30. These changes were accomplished by using end primers 10 [SEQ ID NO: 24] and 11 [SEQ ID NO: 26] in conjunction with mutagenizing primers 918 [SEQ ID NO: 58] and 919 [SEQ ID NO: 59]. Finally, for stage 0-C, to the change of Asp to Arg at position 94 in Stage 0-A were added the changes were from Asn-Ile-Lys to Thr-Phe-Ser at positions 28-30. These changes were accomplished with the same end primers and mutagenizing primers described above for the Stage 0-B construct.

-39-

### TABLE I

# HEAVY CHAIN SEQUENCES

5			FR1
	NEWM		?VQLXXSGPGLVRPSQTLSLTCTVSGSTFS
10	Humanized	Anti-VLA4	l <b>:</b>
10	STAGE O-A STAGE O-B STAGE O-C		QVQLQEFNIK QVQLQEF
15	STAGE 1		QVQLQEFNIK
	STAGE 2		QVQLQEFNIK
20	STAGE 3	(STAW) (KAITAS) (SSE) (KRS) (AS)	QVQLQEFNIK QVQLQEFNIK QVQLQEFNIK QVQLQEFNIK QVQLQEFNIK
25			
25			
25			FR2
25	NEWM		FR2 WVRQPPGRGLEWIG
30	NEWM Humanized	Anti-VLA4	WVRQPPGRGLEWIG
30		Anti-VLA4	WVRQPPGRGLEWIG
	Humanized STAGE O-A STAGE O-B	Anti-VLA4	WVRQPPGRGLEWIG
30	Humanized STAGE O-A STAGE O-B STAGE O-C	Anti-VLA4	WVRQPPGRGLEWIG :
30	Humanized STAGE O-A STAGE O-B STAGE O-C	(STAW) (KAITAS) (SSE) (KRS) (AS)	WVRQPPGRGLEWIG

-40-

# TABLE I (Cont'd)

# FR3

5	NEWM		RVTMLVDTSKNQFSLRLSSVTAADTAVYYCAR
	Humanized	Anti-VLA4	•
10	STAGE O-A STAGE O-B STAGE O-C		
15	STAGE 2		b
20	STAGE 3	(STAW) (KAITAS) (SSE) (KRS) (AS)	S.TAWD KA.ITASDSSEDSD
25	NEWM		FR4 WGQGSLVTVSS
	Humanized	Anti-VLA4:	
30	STAGE O-A STAGE O-B STAGE O-C		TT TT
35	STAGE 1		TT
40	STAGE 3	(STAW) (KAITAS) (SSE) (KRS) (AS)	TT TT TT
	Note: X de	notes Glx.	, ? denotes Q or E.

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#### Light Chain Modifications 2.

our experience, the humanized light In generally requires few, if any, modifications. However, in the preparation of humanized anti-VLA4 antibodies, it became apparent that the light chain of HP1/2 did require 5 several empirical changes. For example, humanized heavy chain of the Stage 2 construct (the Ser mutant) with murine light chain was about 2.5 fold lower potency than murine HP1/2, while the same humanized heavy chain with humanized light chain was about 4-fold lower potency. 10 The Stage 1 humanized  $V_{K}$  construct was designated  $V_{K1}$ (DQL) and the DNA sequence and its translated amino acid sequence are set forth in SEQ ID NO: 30 and SEQ ID NO: 31, respectively. The DQL mutations arose from the PCR primer used in the initial cloning of the  $V_{\mbox{\scriptsize K}}$  region (see Example 1). Alterations were made in the light chain, generating two mutants, SVMDY and DQMDY (VK2 and VK3, respectively). The SVMDY mutant was prepared from the DQL sequence using oligos 10 [SEQ ID NO: 24] and 11 [SEQ ID NO: 26] for DY sequences with oligos 697 [SEQ ID NO: 60 and 698 [SEQ ID NO: 61] for SVM sequences. The VK2 (SVMDY) DNA sequence and its translated amino acid sequence are set forth in SEQ ID NO: 62 and SEQ ID NO: 63, respectively. The DQMDY sequences were restored to the original REI framework sequences by two-step PCRdirected mutagenesis using end primers 10 [SEQ ID NO: 24] and 11 [SEQ ID NO: 26] with mutagenic primers 803 [SEQ ID NO: 64] and 804 [SEQ ID NO: 65], and using the SVMDY sequence as template. The VK3 (DQMDY) DNA sequence and its translated amino acid sequence are set forth in SEQ ID NO: 66 and SEQ ID NO: 67, respectively. The change in the amino terminus (SVM versus DQM) is not relevant, and relates to the amino terminus of the murine light chain. The other two changes, D and Y, were made to improve potency, and did indeed do so as described in Example 7. An alignment of the CDR-grafted DQL (VK1), SVMDY (VK2)

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and DQMDY (VK3) framework sequences as compared with the REI sequence is shown in Table II.

When the AS mutant heavy chain was combined with the improved light chain (SVMDY), the resulting humanized antibody was equipotent with murine HP1/2 as shown in Table III.

# 3. Alternative Humanized V<sub>H</sub> and V<sub>v</sub> Regions

Alternatively, a humanized  $V_H$  region sequence based on HP1/2  $V_H$  region [SEQ ID NO: 5] may be prepared. One such alternative is designated  $V_H$ -PDLN. The DNA sequence of PDLN  $V_H$  and its translated amino acid sequence are set forth as SEQ ID NO: 68 and SEQ ID NO: 69, respectively.

In addition, an alternative humanized  $V_K$  region sequence based on the HP1/2  $V_K$  region [SEQ ID NO: 9] may be prepared. One such alternative  $V_K$  sequence is designated  $V_K$ -PDLN and its translated amino acid sequence are set forth as SEQ ID NO: 70 and SEQ ID NO: 71, respectively.

The humanized  $V_H$ -PDLN was prepared by ligating 12 oligonucleotides, which together span the entire humanized variable region, and by screening for constructs having the correct sequence. The protocol is described in more detail below.

Oligonucleotides 370-119 through 370-130 (SEQ ID NO:72 through SEQ ID NO:83, respectively) (20 pmoles each) were dried down, and separately resuspended in 20  $\mu$ l 1x Kinase Buffer containing 1 mM ATP and 1  $\mu$ l T4 polynucleotide kinase (10 U/ $\mu$ l). The kinase reaction mixture was incubated for 1 hour at 37°C. The reaction was terminated by incubating at 70°C 5 minutes.

The kinase-treated oligonucleotides were combined with each other (240  $\mu$ l total) and ligated together with 26  $\mu$ l 10 mM ATP and 2  $\mu$ l T4 DNA ligase (10 U/ $\mu$ l), and the reaction mixture was incubated at room temperature for 6 hours. The ligation reaction mixture was extracted

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with phenol:chloroform (1:1) saturated with TE buffer, and then ethanol precipitated and washed 5 times with 70% ethanol.

The dried and washed ethanol precipitate was resuspended in 50  $\mu$ l 1x 150 mM Restriction Enzyme Buffer (10x 150 mM Restriction Enzyme Buffer is 100 mM Tris-HCl, pH 8.0, 1.5 M NaCl, 100 mM MgCl<sub>2</sub>, 1 mg/ml gelatin, 10 mM dithiothreitol) and incubated with restriction enzymes BstE2 and PstI for 16 hours at 37°C. The digestion products were electrophoresed through a 2% agarose gel, and the band corresponding to 330 bp was excised. The fragment was eluted using GENECLEAN II® and the eluate was ethanol precipitated. The ethanol precipitate was resuspended in 20  $\mu$ l TE buffer.

Next, the 330 bp fragment was ligated into vector pLCB7 which was prepared for ligation by digesting with PstI and BstE2, dephosphorylating the 5' ends with calf alkaline phosphatase, fractionating on a low melting temperature agarose (LMA) gel, and excising the pLCB7/PstI/BstE2 LMA fragment. The pLCB7 LMA fragment was then ligated to the 330 bp oligonucleotide fragment encoding the humanized V<sub>H</sub> region using T4 DNA ligase.

The ligation mixture was used to transform  $\underline{E}$ .  $\underline{coli}$  JA221(Iq) to ampicillin resistance. Colonies were grown up and mini-prep DNA was prepared. The recombinant plasmids were screened for the presence of an approximately 413 bp  $\underline{NotI/BstE2}$  fragment. DNA sequence analysis identified vector pMDR1023 as having the designed humanized  $V_H$ -PDLN sequence.

The humanized  $V_K$ -PDLN was prepared by ligating 12 oligonucleotides, which together span the entire humanized  $V_K$ -PDLN variable region, and by screening for constructs having the correct sequence. The protocol is described in more detail below.

Oligonucleotides 370-131 through 370-142 (SEQ ID NO:84 through SEQ ID NO:95, respectively)

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(20 pmoles each) were dried down, and separately resuspended in 20  $\mu$ l 1x Kinase Buffer containing 1 mM ATP and 1  $\mu$ l T4 polynucleotide kinase (10 U/ $\mu$ l). The kinase reaction mixture was incubated for 1 hour at 37°C. The reaction was terminated by incubating at 70°C for 5 minutes.

The kinase-treated oligonucleotides were combined with each other (240  $\mu$ l total) and ligated together with 26  $\mu$ l 10 mM ATP and 2  $\mu$ l T4 DNA ligase (10 U/ $\mu$ l), and the reaction mixture was incubated at room temperature for 6 hours. The ligation reaction mixture was extracted with phenol:chloroform (1:1) saturated with TE buffer, and then ethanol precipitated and washed 5 times with 70% ethanol.

The dried and washed ethanol precipitate was resuspended in 40  $\mu$ l TE, then electrophoresed through a 2% agarose gel, and the band corresponding to 380 bp was excised. The fragment was eluted using GENECLEAN II® and the eluate was ethanol precipitated. The ethanol precipitate was resuspended in 20  $\mu$ l TE buffer.

Next, the 380 bp fragment was ligated into vector pNN03, which was prepared for ligation by linearizing with HindIII and BamHI, dephosphorylating the 5' ends with calf alkaline phosphatase, fractionating on a low melting temperature agarose gel, and excising the band corresponding to linearized pNN03 (2.7 kb). The linearized, dephosphorylated pNN03 was then ligated to the 380 bp oligonucleotide fragment encoding the humanized  $V_{\rm K}$  region using T4 DNA ligase.

The ligation mixture was used to transform <u>E. coli</u>

JA221(Iq) to ampicillin resistance. Colonies were grown
up and mini-prep DNA was prepared. The recombinant
plasmids were screened for the presence of the variable

WO 94/16094

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region fragment. DNA sequence analysis identified vector pMDR1025 as having the designed humanized  $V_K\text{-PDLN}$  sequence.

When an antibody with a  $V_H$ -PDLN containing heavy chain and with a  $V_K$ -PDLN containing light chain was assayed for potency according to Example 7, the resulting humanized antibody was approximately equipotent with the murine HP1/2 antibody.

-46-

# TABLE II LIGHT CHAIN SEQUENCES

5		FR1
	REI Humanized Anti-VLA4:	DIQMTQSPSSLSASVGDRVTITC
10	Construct VK1 (DQL) Construct VK2 (SVMDY) Construct VK3 (DQMDY)	
	•	
15		FR2
10	REI Humanized Anti-VLA4:	WYQQTPGKAPKLLIY
	VK1 (DQL)	K
	VK2 (SVMDY)	K
20	VK3 (DQMDY)	K
	·	FR3
25	REI GVP: Humanized Anti-VLA4:	SRFSGSGSGTDYTFTISSLQPEDIATYYC
		· · · · · · · · · · · · · · · · · · ·
	VK2 (SVMDY)	DYF
30	VK3 (DQMDY)1	DYF
		FR4
	REI	FGQGTKLQIT
35	Humanized Anti-VLA4:	
	VK1 (DQL)	VE.K
	VK2 (SVMDY)	VE.K
	VK3 (DQMDY)	VE.K

WO 94/16094 PCT/US94/00266

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-47-

### Example 5

### Expression of Rec mbinant Anti-VLA4 Antibodies

Each of the  $V_H$  region sequences and  $V_K$  region sequences prepared according to Examples 1-4, transferred into expression vectors with constant region sequences, and the vectors are transfected, preferably via electroporation, into mammalian cells. The heavy and light chain sequences may be encoded on separate vectors and co-transfected into the cells or alternatively heavy and light chain sequences may be encoded by and transfected as a single vector. Such a single vector will contain 3 expression cassettes: one for Iq heavy chain, one for Ig light chain and one for a selection marker. Expression levels of antibody are measured following transfection, as described below, described in Example 7.

 $V_H$  and  $V_K$  region sequences as described in Example 4, were inserted into various cloning and expression vectors. For the anti-VLA4  $V_H$  region sequences, plasmids containing such sequences [as described in Examples 1-4] were digested with PstI and BstE2. The plasmid DNA after digestion with PstI and BstE2, was dephosphorylated and electrophoresed through 2% agarose gel. The band for ligation was excised and the DNA elected using the GENECLEAN technique (Bio101 Inc., LaJolla, California), ethanol precipitated and resuspended in 20  $\mu$ l TE buffer (10mM Tris-HCl, 1mM Na<sub>2</sub> EDTA). Then, 10  $\mu$ l of the resuspended DNA was used for ligation with the PstI/BstE2 digested  $V_H$  region sequence.

The ligation mixture was used to transform  $\underline{E}$ .  $\underline{coli}$  K 12 JA221 (Iq) to ampicillin resistance.  $\underline{E}$ .  $\underline{coli}$  K12 JA221 (Iq) cells have been deposited with the ATCC (accession number 68845). Recombinant colonies were screened for the presence of the  $V_H$  insert. Some of the plasmids containing such fragments were sequenced. The

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V<sub>H</sub>-containing plasmids were designated pBAG 184 STAW), pBAG 183 ( $V_H$ -KAITAS), pBAG 185 ( $V_H$ -KRS), pBAG 207 ( $V_H$ -SSE) and pBAG 195 ( $V_H$ -AS), and were deposited in E. coli K12 J221 (Iq) cells with the ATCC as accession nos. 69110, 69109, 69111, 69116 and 69113, respectively. plasmid containing alternative V<sub>H</sub>-PDLN Was designated pMDR1023.

For the  $V_K$  region sequences, the DNA encoding these sequences were amplified for cloning and transformation using PCR. Prior to amplification, 20 pmoles of each of the  $V_{\mbox{\scriptsize K}}$  chain primers were kinased by incubation with  $T_4$ polynucleotide kinase at 37°C for 60 minutes by a conventional protocol. The kinase reactions were stopped by heating at 70°C for 10 minutes.

The PCR reactions each contained 10  $\mu$ l 10X PCR buffer (10X PCR buffer is 100 mM Tris/HCl, pH 8.3, 500 mM  $\,$ KCl, 15 mM MgCl<sub>2</sub>, 0.01% gelatin, 20 pmoles each of the appropriate kinased primers, 20  $\mu$ l cDNA, 0.5  $\mu$ l Tag polymerase (5 U/ $\mu$ l, Perkin Elmer-Cetus) and 49.5  $\mu$ l  $H_2O$ . The PCR conditions were 30 cycles of incubation for: 1 minute at 94°C; 2 minutes at 40°C (for heavy chain PCR) or at 55°C (for light chain PCR); and 2 minutes at 72°C. For VK1-DQL, primers were 370-247 [SEQ ID NO: 96] and 370-210 [SEQ ID NO: 97]. For VK2-SVMDY, primers were 370-269 [SEQ ID NO: 98] and 370-210 [SEQ ID NO: 97]. For VK3-DQMDY, primers were 370-268 [SEQ ID NO: 99] and 370-210 [SEQ ID NO: 97].

The reaction mixtures were electrophoresed through 2% agarose gel, and the bands corresponding to the expected sizes of the light chain variable region (-330 bp) were excised with AgeI and BamHI. The DNA in those bands were eluted using the GENECLEAN™ technique (Bio101 Inc., LaJolla, California), ethanol precipitated and subsequently each resuspended in 20  $\mu$ l TE buffer (10 mM Tris-HCl, 1 mM Na2EDTA).

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Klenow fragment of DNA polymerase (New England Biolabs, 5 U/ $\mu$ l) (1  $\mu$ l) was added to the purified PCR fragments in a reaction volume of 25  $\mu$ l containing 1x ligation buffer (10x ligation buffer is 0.5 M Tris/HCl, pH 7.5, 100 mM MgCl<sub>2</sub> and 40 mM DTT) and 0.125 mM each of dXTPs and the reaction incubated at room temperature for 15 minutes. The reaction was terminated by incubation at 70°C for 5 minutes, and then stored on ice.

The fragment from each PCR reaction is ligated to a plasmid such as pNN03 or a plasmid derived from pNN03 such as pLCB7, which had been previously linearized by EcoRV, dephosphorylated and fractionated through low temperature melting agarose. Such plasmids, including pNN03 and pLCB7 have been described in co-pending and co-assigned (Burkly et al., U.S. Ser. No. 07/916,098, filed July 24, 1992 [75]).

The ligation mixture was used to transform E.coli K12 JA221(Iq) to ampicillin resistance. E.coli K12 JA221(Iq) cells are deposited with American Type Culture Collection (accession number 68845). colonies were screened for the presence of the  $\boldsymbol{V}_{\boldsymbol{K}}$  insert. Some of the plasmids containing such fragments were sequenced. The  $V_{\kappa}$ -containing plasmids were designated pBAG 190 (VK1-DQL), pBAG 198 (VK2-SVMDY) and pBAG 197 (VK3-DQMDY), and were deposited in  $\underline{E}$ .  $\underline{coli}$  K12 JA 221 (Iq) cells with the ATCC as accession nos. 69112, 69115 and 69114, respectively. The plasmid containing the alternative  $V_K$  (PDLN) region was designated pMDR 1025.

In a series of experiments, the expression vectors encoding recombinant anti-VLA4 heavy and light chains are transfected via electroporation and the cells are then cultured for 48 hours. After 48 hours of culture, the cells are radiolabelled using <sup>35</sup>S-cysteine overnight and then the cell extracts and conditioned media are immunoprecipitated by incubation with protein A-Sepharose. The protein A-Sepharose is washed and the

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bound proteins are elut d with SDS-PAGE loading buffer. The samples are analyzed via electrophoresis through 10% SDS-PAGE gels under reducing conditions. In this way, light chain expression is detected only as a consequence of the light chains being associated with the heavy chains. The expected sizes of the heavy and light chains as visualized in the 10% gels are 50 kD and 25 kD, respectively.

Since recombinant anti-VLA4 antibody molecules, prepared as described in Examples 1-4, may be stably expressed in a variety of mammalian cell lines, it is possible to express recombinant antibody nonsecreting myeloma or hybridoma cell lines under the control of Ig-gene promoters and enhancers or in nonlymphoid cells, such as Chinese hamster ovary (CHO) cells, in conjunction with vector amplification using DHFR selection. Recently, Bebbington et al., 1992 [76] have described a method for the high-level expression of a recombinant antibody from myeloma cells using a glutamine synthetase gene as an amplifiable marker. This GS expression system is most preferred for the production of recombinant anti-VLA4 antibody molecules according to the present invention. The methods, vectors with hCMV promoters and with 5' untranslated sequences from the hCMV-MIE genes including cell lines (most preferably NSO) and media for GS expression of recombinant antibodies is described in detail in Bebbington et al., 1992 [76], WO86/05807 [77], WO87/04462 [78], WO89/01036 [79] and WO89/10404 [80].

In accordance with the teachings of these publications, NSO cells were transfected with a heavy chain sequence having the VH-AS region sequence [SEQ ID NO: 54] and a light chain sequence having the VK-SVMDY sequence [SEQ ID NO: 66] to obtain a stable cell line secreting a humanized recombinant anti-VLA4 antibody with high potency comparable to the murine HP1/2 antibody.

-51-

This cell line has been deposited with the ATCC on November 3, 1992 and given accession no. CRL 11175. The AS/SVMDY humanized antibody is at least equipotent with or perhaps more potent than the murine HP1/2 antibody.

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### Example 6

# Purification of MAbs from Conditioned Media for Assay

To obtain accurate values for half-maximal binding or inhibition, stock solutions of purified antibodies are needed at known concentrations. Stable cell lines secreting the antibodies of interest were made and the humanized recombinant anti-VLA4 antibodies were purified from conditioned medium using conventional protein A chromatography. The concentration of the purified antibodies is assessed by their absorption coefficient at 280 nm, which is well established for antibodies.

A cell line producing a humanized anti-VLA4 antibody is grown in roller bottles in Dulbecco's modified Eagle medium containing 10% fetal bovine serum. A 2 liter batch of conditioned medium is used for each purification run. Cells are removed from the medium by centrifugation in a RC-3B preparative centrifuge (4K, 30 minutes, H4000 rotor) and the supernatant is filtered first through a 0.45  $\mu$  membrane and then through a 0.22  $\mu$  membrane. The medium is stored at 4°C until it can be processed.

Two liters of conditioned medium is concentrated to 220 ml in a spiral ultrafiltration unit (Amicon, Corp., Cherry Hill Drive, Danvers, MA 01923) that is equipped with an S1Y30 (YM30) Diaflo cartridge. The concentrate is diluted with 400 ml of protein A binding buffer (3M NaCl, 1.5M glycine pH 8.9) and again concentrated to The concentrate is treated in batch with 0.5 ml Fast Flow Protein A Sepharose 4 (Pharmacia, Inc., 800 Centennial Avenue, Piscataway, NJ 08854) using a raised stir bar to agitate the mixture. After an overnight incubation at 4°C. the resin is collected centrifugation (5 minutes, 50 g), washed twice with 20 volumes of protein A binding buffer (using centrifugation to recover the resin), and transferred to a column for subsequent treatment. The column is washed four times

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with 0.5 ml of protein A binding buffer, two times with 0.25 ml of PBS, and the IgG is eluted with Pierce IgG elution buffer (Pierce Chemical Co., Rockford, IL. 61105 Cat No. 21004Y or 21009Y). 180  $\mu$ l fractions are collected, which are neutralized with 20  $\mu$ l of 1M HEPES 5 pH 7.5. Fractions are analyzed for absorbance at 280 nm and by SDS-PAGE. The gel is stained with Coomassie blue. Peak fractions are pooled. 100  $\mu$ l (14 ml/ml) is diluted with 100  $\mu$ l of PBS and subjected to gel filtration on a Superose 6 FPLC column (Pharmacia, Inc., 800 Centennial Avenue, Piscataway, NJ 08854 ) in PBS. The column is run at 20 ml/hour and 1.5 minute fractions are collected. Peak column fractions are pooled, aliquoted, frozen on dry ice, and stored at -70°C. SDS-polyacrylamide gel profile of the final product is obtained under reducing and non-reducing conditions. In some cases when the sample is analyzed under non-reducing conditions, about 10% of the product is not an intact antibody. Studies in these cases indicate that this product is a heavy-light chain dimer. This has been previously recognized as a problem with IgG4 antibodies.

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### Example 7

# Determination of Relative Binding Affinities of Humanized Recombinant Anti-VLA4 Antibodies

Recombinant antibodies according to the present invention are purified, as described in Example 6, and are assayed to determine their specificity for VLA4 and their binding affinity or potency. In particular, the potency of a recombinant anti-VLA4 antibody was assessed by calculating the half-maximal binding constant (reported as ng/ml or  $\mu$ g/ml of purified antibody) using two different assays described as follows.

Inhibition of VLA4-dependent adhesion to VCAM1 The critical function of an anti-VLA4 antibody is defined by the ability to inhibit the VCAM1/VLA4 adhesion It has been previously shown (Lobb et al., 1991a, [81]) that purified recombinant soluble VCAM1 (rsVCAM1) can be immobilized on plastic and functional adhesion molecule. Immobilized rsVCAM1 binds VLA4-expressing cells such as the human B cell line Ramos, and this binding can be inhibited by MAbs to VCAM1, such as 4B9 or MAbs to VLA4, such as HP1/2. assay provides a reproducible method to assess the potency of any humanized recombinant antibody. Briefly, the antibody solution is diluted, and the serial antibody dilutions are incubated with Ramos cells, which are then incubated with rsVCAM1-coated plates. The Ramos cells are fluorescently labelled as described by Lobb, 1991b [82], and binding assessed by fluorescence in 96 well cluster plates according to the following protocol.

Recombinant soluble VCAM1 was prepared and purified essentially as described by Lobb et al., 1991a [81]. Soluble VCAM is diluted to 10  $\mu$ g/ml in 0.05 M NaHCO<sub>3</sub>, (15mM NaHCO<sub>3</sub>, 35mM Na<sub>2</sub>CO<sub>3</sub>) pH 9.2. Then 50  $\mu$ l/well is added into a Linbro Titertek polystyrene 96 well plate, flat bottom, Flow Labs catalog #76-231-05. The plate is incubat d at 4°C overnight.

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Following this incubation, the contents of the wells are rem ved by inverting and blotting the plate. To the empty wells, 100  $\mu$ l/well of 1% of BSA in PBS, 0.02% NaN, is added for 1 hour or longer at room temperature. If the plate is not to be used immediately, it can be blocked and stored for one week at 4°C. BSA is added to some wells to assess non-specific binding.

For binding quantitation, VLA4 presenting cells, preferably Ramos cells, should be prelabelled. cells may be radiolabelled or fluorescently labelled. 10 For radiolabelling, prelabelling of the cells may be done using overnight <sup>3</sup>H-thymidine  $(0.5 \mu Ci/ml)$ . Alternatively, and preferably, the cells are preincubated with BCECF-AM (chemical name: 2',7'-bis-(2-carboxyethyl)-5(and -6) carboxyfluorescein, acetoxymethyl 15 Molecular Probes Inc., Eugene, Oregon, catalog #B-1150). For this method, cells are suspended to 5 x  $10^6/\mathrm{ml}$ , 2  $\mu\mathrm{M}$ BCECF-AM is added and the mixture is incubated for 30 minutes at 37°C. Following either method, the cells are washed with RPMI, 2% FBS, pH 7.4. RPMI with 1% FBS may 20 also be used.

For the binding study, 2-4 x 10 $^6$  cells/ml in RPMI, 2% FBS are resuspended, then 50  $\mu l$  of labelled cells are added per well for 10 minutes of binding at room temperature.

After the 10 minute incubation, the contents of the wells are removed by inversion and the plates washed 1-2 times gently with RPMI, 2% FBS. When examined under a light microscope, BSA blank wells should have very few cells bound. A brief inverted spin may be included to remove cells not firmly attached and the plates may be washed again 1-2 times.

For the BCECF-AM method, 100  $\mu$ l of 1% NP40 is added to each well to solubilize the cells and then the plate is read on a fluorescence plate scanner. (If the radiolabelling method is used, 100  $\mu$ l of 0.1% NaOH is

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added to each well and then the contents of each well are transferred to scintillation vials containing cocktail).

A volume of 50  $\mu$ l of labelled cells should be counted to obtain a total known value added to each well. Then the 50  $\mu$ l of labelled cells are added to either a well containing only 100  $\mu$ l of 1% NP40 or to a scintillation vial depending on the method used.

For antibody blocking studies, 100  $\mu$ l/well of murine HP1/2 MAb (anti-VLA4) typically at 10  $\mu$ g/ml in RPMI, 2% FBS are added to the rsVCAM1 coated plates and incubated for 30 minutes at room temperature prior to cell binding as described above. MAb HP1/2 (anti-VLA4) or any recombinant humanized anti-VLA4 antibody prepared as described herein must be preincubated with labelled cells for 30 minutes at room temperature prior to the cell binding. Concentrations of the antibodies preincubated will vary, but generally concentrations were in the range of about 1  $\mu$ g/ml.

In these adhesion assays, murine HP1/2 inhibits Ramos cell binding completely at about 40 ng/ml, and half 20 maximally at about 15 ng/ml (10  $\mu$ M). The results of adhesion assays as represented by the calculated halfmaximal binding constants using humanized recombinant anti-VLA4 antibodies made according to the present 25 invention are shown in Table III. The number (n) of experiments performed for each value is indicated for the recombinant humanized antibodies. As discussed below, these results generally compare well with the results obtained with the FACS binding assay.

The potency of recombinant Stage 0, Stage 1, Stage 2 and Stage 3 antibodies having the VK1 (DQL) light chain that had been purified from stably transfected YB2/0 cell lines was measured in the adhesion assay, as shown in Table III. The results showed that there was no inhibition detected in concentrations up to 1 μg/ml (1000 ng/ml) with the Stage 0-B and 0-C humanized antibodies.

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The results with the recombinant Stage 3 antibodies STAW and AS having the improved VK2 (SVMDY) light chain showed that the AS/SVMDY antibody was at least equipotent and perhaps more potent than the murine HP1/2 antibody. Certain Stage 2 and Stage 3 constructs showed potencies of about 20% to about 100% of the potency of the murine HP1/2 antibody.

### 2. FACS Assays

The binding of humanized recombinant antibodies to the cell surface can be assessed directly by fluorescence activated cell sorter (FACS) analysis, using fluorescently labelled antibodies. This is a standard technique that also provides half-maximal binding information following dose response measurements. The FACS methods are described in Lobb et al., 1991b [82].

Briefly, 25  $\mu$ l cells (4 x 10 $^6/m$ l in FACS buffer (PBS 2% FBS, 0.1% NaN<sub>3</sub>) on ice are added to 5  $\mu$ l of 5  $\mu$ g/ml FITC or phycoerythrin (PE) conjugated antibody in FACS buffer, and incubated in V-bottomed microtiter wells on ice for 30 minutes. To the wells, 125  $\mu$ l of FACS buffer is added, the plates are centrifuged at 350  $\times$  g for 5 minutes, and the supernatant is shaken off. To each well added 125  $\mu$ l FACS buffer, then the cells are transferred to 12 x 75 mm Falcon polystyrene tubes and resuspended to a final volume of 250  $\mu$ l in FACS buffer. The mixture is analyzed on a Becton Dickinson FACStar. The results of the FACS assays as represented by the half-maximal calculated binding constructs humanized recombinant anti-VLA4 antibodies made according to the present invention are shown in Table III and the number (n) of experiments performed for each value is indicated for the humanized antibodies. Table III also shows the potency calculated from the combined adhesion and FACS assays. Murine HP1/2 binds half-maximally to Ramos cells at 15 ng/ml. The AS/SVMDY humanized antibody binds half-maximally to Ramos cells at 12 ng/ml.

the two assays (i.e., adhesion and FACS assays) show an excellent correlation for the murine antibody and the humanized AS/SVMDY antibody.

TABLE III

SUMMARY OF HALF-MAXIMAL BINDING CONSTANTS FOR HUMANIZED RECOMBINANT ANTI-VLA4 ANTIBODIES

			_ <del>_</del>	
Antibody	Adhesion Assay	FACS Assay	Combination	
Murine HP1/2	15 ng/ml	15 ng/ml	15 ng/ml	
Stage 0 (Humanized heavy chain)	>1000 ng/ml (n=3)		- Ig/mi	
Stage 1 (Humanized heavy chain)	228 ng/ml (n=6)	-	228 ng/ml (n=6)	
Stage 2 (Ser mutant)	56 ng/ml (n=14)	47 ng/ml (n=6)	60 ng/ml (n=20)	
Stage 3		·	(20)	
(STAW)	30 ng/ml (n=3)	33 ng/ml (n=3)	32 ng/ml (n=6)	
(KAITAS)	85 ng/ml (n=2)	100 ng/ml (n=1)	90 ng/ml (n=3)	
(SSE)	100 ng/ml (n=2)	40 ng/ml (n=1)	80 ng/ml (n=3)	
(KRS)	50 ng/ml (n=2)	70 ng/ml (n=1)	57 ng/ml (n=3)	
(AS)	28 ng/ml (n=2)	14 ng/ml (n=2)	21 ng/ml (n=4)	
Constructs with improved light chain				
STAW/SVMDY	25 ng/ml (n=4)	35 ng/ml (n=3)	29 ng/ml (n=7)	
AS/SVMDY		12 ng/ml (n=2)	12 ng/ml (n=4)	

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### <u>Deposits</u>

The following plasmids in <u>E. coli</u> K12 J221 (Iq) cells were deposited under the Budapest Treaty with American Type Culture Collection (ATCC), Rockville, Maryland (USA) on October 30, 1992. The deposits are identified as follows:

	<u>Plasmid</u>		Accession No.
	pBAG 184	(V <sub>H</sub> -STAW)	69110
10	pBAG 183	(V <sub>H</sub> -KAITAS)	69109
	pBAG 185	(V <sub>H</sub> -KRS)	69111
	<b>pBAG</b> 207	(V <sub>H</sub> -SSE)	69116
	pBAG 195	(V <sub>H</sub> -AS)	69113
15	pBAG 190	(VK1-DQL)	69112
	pBAG 198	(VK2-SVMDY)	69115
	<b>pBAG</b> 197	(VK3-DQMDY)	69114

In addition, an NSO cell line producing humanized recombinant anti-VLA4 antibody was deposited under the Budapest Treaty with American Type Culture Collection (ATCC), Rockville, Maryland (USA) on November 3, 1992. The deposit was given ATCC accession no. CRL 11175.

### 25 <u>Sequences</u>

The following is a summary of the sequences set forth in the Sequence Listing:

	SEQ ID NO:1	DNA sequence of CG1FOR primer
	SEQ ID NO:2	DNA sequence of CK2FOR primer
30	SEQ ID NO:3	DNA sequence of VH1BACK primer
	SEQ ID NO:4	DNA sequence of VH5BACK primer
	SEQ ID NO:5	DNA sequence of HP1/2 heavy chain variable region
35	SEQ ID NO:6	Amino acid sequence of HP1/2 heavy chain variable region
	SEQ ID NO:7	DNA sequence of VK1BACK primer

WO 94/16094 PCT/US94/00266

-60-

	SEQ ID NO:8	DNA sequ nce of VK7BACK primer
-	SEQ ID NO:9	DNA sequence of HP1/2 light chain variable region
5	SEQ ID NO:10	Amino acid sequence of HP1/2 light chain variable region
• •	SEQ ID NO:11	DNA sequence of VH1FOR primer
10	SEQ ID NO:12	DNA sequence of VK3BACK primer
	SEQ ID NO:13	DNA sequence of VK1FOR primer
15	SEQ ID NO:14	DNA sequence of VH insert in M13VHPCR1
	SEQ ID NO:15	Amino acid sequence of VH insert in M13VHPCR1
20	SEQ ID NO:16	DNA sequence of VK insert in M13VKPCR2
	SEQ ID NO:17	Amino acid sequence of VK insert in M13VKPCR2
25	SEQ ID NO:18	DNA sequence of OLIGO598
	SEQ ID NO:19	DNA sequence of OLIGO599
30	SEQ ID NO:20	DNA sequence of OLIGO600
30	SEQ ID NO:21	DNA sequence of OLIGO605
	SEQ ID NO:22	DNA sequence of OLIGO606
35	SEQ ID NO:23	DNA sequence of OLIGO607
	SEQ ID NO:24	DNA sequence of OLIGO10
40	SEQ ID NO:25	DNA sequence of OLIGO385
40	SEQ ID NO:26	DNA sequence of OLIGO11
. •	SEQ ID NO:27	DNA sequence of OLIGO391
45	SEQ ID NO:28	DNA sequence of Stage 1 heavy chain variable region
50	SEQ ID NO:29	Amino acid sequence of Stage 1 heavy chain variable region
50	SEQ ID NO:30	DNA sequence of VK1 (DQL) light chain variable region

	SEQ ID NO:31	Amino acid sequence of VK1 (DQL) light chain variable region
5	SEQ ID NO:32	DNA sequence of Stage 2 heavy chain variable region
	SEQ ID NO:33	Amino acid sequence of Stage 2 heavy chain variable region
10	SEQ ID NO:34	DNA sequence of OLIGO684
	SEQ ID NO:35	DNA sequence of OLIGO683
15	SEQ ID NO:36	DNA sequence of OLIGO713
	SEQ ID NO:37	DNA sequence of OLIGO716
20	SEQ ID NO:38	DNA sequence of STAW heavy chain variable region
	SEQ ID NO:39	Amino acid sequence of STAW heavy chain variable region
25	SEQ ID NO:40	DNA sequence of OLIGO706
23	SEQ ID NO:41	DNA sequence of OLIGO707
30	SEQ ID NO:42	DNA sequence of KAITAS heavy chain variable region
	SEQ ID NO:43	Amino acid sequence of KAITAS heavy chain variable region
35	SEQ ID NO:44	DNA sequence of OLIGO768
	SEQ ID NO:45	DNA sequence of OLIGO769
40	SEQ ID NO:46	DNA sequence of SSE heavy chain variable region
40	SEQ ID NO:47	Amino acid sequence of SSE heavy chain variable region
45	SEQ ID NO:48	DNA sequence of OLIGO704
43	SEQ ID NO:49	DNA sequence of OLIGO705
50	SEQ ID NO:50	DNA sequence of KRS heavy chain variable region
30	SEQ ID NO:51	Amino acid sequence of KRS heavy chain variable region
	SEQ ID NO:52	DNA sequence of OLIGO745

	SEQ ID NO:53	DNA sequence of OLIGO746
5	SEQ ID NO:54	DNA sequence of AS heavy chain variable region
_	SEQ ID NO:55	Amino acid sequence of AS heavy chain variable region
10	SEQ ID NO:56	DNA sequence of OLIGO915
	SEQ ID NO:57	DNA sequence of OLIGO917
	SEQ ID NO:58	DNA sequence of OLIGO918
15	SEQ ID NO:59	DNA sequence of OLIOG919
	SEQ ID NO:60	DNA sequence of OLIGO697
20	SEQ ID NO:61	DNA sequence of OLIGO698
	SEQ ID NO:62	DNA sequence of VK2 (SVMDY) light chain variable region
25	SEQ ID NO:63	Amino acid sequence of VK2 (SVMDY) light chain variable region
	SEQ ID NO:64	DNA sequence of OLIGO803
30	SEQ ID NO:65	DNA sequence of OLIGO804
30	SEQ ID NO:66	DNA sequence of VK3 (DQMDY) light chain variable region
35	SEQ ID NO:67	Amino acid sequence of VK3 (DQMDY) light chain variable region
	SEQ ID NO:68	DNA sequence of PDLN heavy chain variable region
40	SEQ ID NO:69	Amino acid sequence of PDLN heavy chain variable region
45	SEQ ID NO:70	DNA sequence of PDLN light chain variable region
45	SEQ ID NO:71	Amino acid sequence of PDLN light chain variable region
50	SEQ ID NO:72	DNA sequence of Oligo 370-119
	SEQ ID NO:73	DNA sequence of Oligo 370-120
	SEQ ID NO:74	DNA sequence of Oligo 370-121

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	SEQ ID NO:75	DNA sequencé of Oligo 370-122
	SEQ ID NO:76	DNA sequence of Oligo 370-123
5	SEQ ID NO:77	DNA sequence of Oligo 370-124
	SEQ ID NO:78	DNA sequence of Oligo 370-125
10	SEQ ID NO:79	DNA sequence of Oligo 370-126
	SEQ ID NO:80	DNA sequence of Oligo 370-127
	SEQ ID NO:81	DNA sequence of Oligo 370-128
15	SEQ ID NO:82	DNA sequence of Oligo 370-129
	SEQ ID NO:83	DNA sequence of Oligo 370-130
20	SEQ ID NO:84	DNA sequence of Oligo 370-131
	SEQ ID NO:85	DNA sequence of Oligo 370-132
	SEQ ID NO:86	DNA sequence of Oligo 370-133
25	SEQ ID NO:87	DNA sequence of Oligo 370-134
	SEQ ID NO:88	DNA sequence of Oligo 370-135
30	SEQ ID NO:89	DNA sequence of Oligo 370-136
	SEQ ID NO:90	DNA sequence of Oligo 370-137
	SEQ ID NO:91	DNA sequence of Oligo 370-138
35	SEQ ID NO:92	DNA sequence of Oligo 370-139
	SEQ ID NO:93	DNA sequence of Oligo 370-140
40	SEQ ID NO:94	DNA sequence of Oligo 370-141
	SEQ ID NO:95	DNA sequence of Oligo 370-142
	SEQ ID NO:96	DNA sequence of VK1-DQL primer 370-247
45	SEQ ID NO:97	DNA sequence of VK1-DQL primer 370-210
	SEQ ID NO:98	DNA sequence of VK2-SVMDY primer 370-269
50	SEQ ID NO:99	DNA sequence of VK3-DQMDY primer 370-268

While we have hereinbefore described a number of embodim nts of this invention, it is apparent that our basic embodiments can be altered to provide other

embodiments that utilize the compositions and processes of this invention. Therefore, it will be appreciated that the scope of this invention includes all alternative embodiments and variations which are defined in the foregoing specification and by the claims appended hereto; and the invention is not to be limited by the specific embodiments that have been presented herein by way of example.

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10		binding to vascular cell adhesion molecule 1
		and endothelial leukocyte adhesion molecule
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		Endothelial Cells: Contributions of
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	[01]	12, 1992
	((2)	·
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		Overlap Extension Using The Polymerase Chain
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		Characterization of a Soluble Form of
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Each of the above-listed references is hereby incorporated by reference in its entirety.

#### SEQUENCE LISTING

- (1) GENERAL INFORMATION:
  - (i) APPLICANT: Lobb, Roy R.; Carr, Frank J.; Tempest, Philip R.
  - (ii) TITLE OF INVENTION: Recombinant Anti-VLA4 Antibody Molecules
  - (iii) NUMBER OF SEQUENCES: 99
  - (iv) CORRESPONDENCE ADDRESS:
    - (A) ADDRESSEE: Allegretti & Witcoff, Ltd.
    - (B) STREET: 10 South Wacker Drive, Suite 3000
    - (C) CITY: Chicago

    - (D) STATE: IL (E) COUNTRY: US
    - (F) ZIP: 60606
  - (v) COMPUTER READABLE FORM:
    - (A) MEDIUM TYPE: Floppy disk
    - (B) COMPUTER: IBM PC compatible
    - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
    - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
  - (vi) CURRENT APPLICATION DATA:
    - (A) APPLICATION NUMBER: US
    - (B) FILING DATE:
    - (C) CLASSIFICATION:
  - (viii) ATTORNEY/AGENT INFORMATION:
    - (A) NAME: McNicholas, Janet M.
    - (B) REGISTRATION NUMBER: 32,918
    - (C) REFERENCE/DOCKET NUMBER: 92,445/D012 US
    - (ix) TELECOMMUNICATION INFORMATION:
      - (A) TELEPHONE: 312-715-1000
      - (B) TELEFAX: 312-715-1234
- (2) INFORMATION FOR SEQ ID NO:1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 31 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note = "CG1FOR PCR primer"

		-74-	
( <b>x</b> i)	SEC	UENCE DESCRIPTION: SEQ ID NO:1:	
GAAGCT	TAG A	CAGATGGGG GTGTCGTTTT G	3
) INF	ORMAT	ION FOR SEQ ID NO:2:	
(Ţ)			
(ii)	` ,		
		❖	
(ix)			
	(A)	NAME/KEY: misc_feature	
	(0)	OTHER INFORMATION: /note= "CK2FOR PCR primer"	
(xi)	SEQ	JENCE DESCRIPTION: SEQ ID NO:2:	
AAGCTI	GA AC	SATGGATAC AGTTGGTGCA GC	32
TMPC	DVST	CON BOD CTO TO NO. 3	,
INFC	MINIT.	ON FOR SEQ ID NO:3:	-
(i)			
	(D)	TOPOLOGY: linear	
(ii)	MOLE	CULE TYPE: cDNA	
(ix)	FEAT	URE:	
	(A)	NAME/KEY: misc_feature	
	(B)	LOCATION: 1	
	(D)	OTHER INFORMATION: /note= "VHIBACK PCR primer"	
(xi)	SEQU	ENCE DESCRIPTION: SEQ ID NO:3:	
TSMAR	CT GC.	AGSAGTCW GG	22
INFO	RMATI(	ON FOR SEQ ID NO:4:	
(i)	SEQUI	ENCE CHARACTERISTICS:	·
• •			
	(B)	TYPE: nucleic acid	
	CAAGCT (ii) (iii) (ix)  AAGCTT INFO (ii) (ix)  (xi) (xi) (xi) (xi) INFOI	CAAGCTTAG A  (i) SEQUICAN  (ii) MOLE  (iii) MOLE  (iii) MOLE  (iii) SEQUICAN  (iii) MOLE  (iiii) MOLE  (iiii) MOLE  (iiiiiiiiiiiiiiiiiiiiiiiiiiiiiiiiiii	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 32 base pairs  (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear  (ii) MOLECULE TYPE: cDNA  (ix) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1 (D) OTHER INFORMATION: /note= "CK2FOR PCR primer"  (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:  AAGCTTGA AGATGGATAC AGTTGGTGCA GC  INFORMATION FOR SEQ ID NO:3:  (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 22 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear  (ii) MOLECULE TYPE: cDNA  (ix) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1 (D) OTHER INFORMATION: /note= "VHIBACK PCR primer"  (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:  TSMARCT GCAGSAGTCW GG  INFORMATION FOR SEQ ID NO:4:  (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 32 base pairs

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

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(A) NAME/KEY: misc_feature (B) LOCATION: 1 (D) OTHER INFORMATI N: /note= "VKSBACK PCR primer"	
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TTGAATTCGG TGCCAGAKCW SAHATYGTKA TG	32
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(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 360 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
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(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "pBAG159 insert: HP1/2 heavy chain variable region; 1, 3 & 6 = E, Q & E in HP1/2	, ,
(ix) FEATURE:	
(A) NAME/KEY: CDS	
(B) LOCATION: 1360	
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GTC AAA CTG CAG CAG TCT GGG GCA GAG CTT GTG AAG CCA GGG GCC TCA	48
Val Lys Leu Gln Gln Ser Gly Ala Glu Leu Val Lys Pro Gly Ala Ser	40
2 6 11 16	
CTC AAC TTC TCC TCC ACC ACC ACC ACC	
GTC AAG TTG TCC TGC ACA GCT TCT GGC TTC AAC ATT AAA GAC ACC TAT	96
Val Lys Leu Ser Cys Thr Ala Ser Gly Phe Asn Ile Lys Asp Thr Tyr 21 26 31	
26 31	
ATG CAC TGG GTG AAG CAG AGG CCT GAA CAG GGC CTG GAG TGG ATT GGA	144
Met His Trp Val Lys Gln Arg Pro Glu Gln Gly Leu Glu Trp Ile Gly	144
36 41 46	
100 100 010 000 000	
AGG ATT GAT CCT GCG AGT GGC GAT ACT AAA TAT GAC CCG AAG TTC CAG	192
Arg Ile Asp Pro Ala Ser Gly Asp Thr Lys Tyr Asp Pro Lys Phe Gln 51	
. 51 56 61	
GTC AAG GCC ACT ATT ACA GCG GAC ACG TCC TCC AAC ACA GCC TGG CTG	240
Val Lys Ala Thr Ile Thr Ala Asp Thr Ser Ser Asn Thr Ala Trp Leu	240
71 76 81	

				GAG Glu				 	288
				GGA Gly					336
 	ACG Thr 116	 	 						360

- (2) INFORMATION FOR SEQ ID NO:6:
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    - (A) LENGTH: 120 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Val Lys Leu Gln Gln Ser Gly Ala Glu Leu Val Lys Pro Gly Ala Ser 2 6 11 16

Val Lys Leu Ser Cys Thr Ala Ser Gly Phe Asn Ile Lys Asp Thr Tyr 21 26 31

Met His Trp Val Lys Gln Arg Pro Glu Gln Gly Leu Glu Trp Ile Gly 36 41 46

Arg Ile Asp Pro Ala Ser Gly Asp Thr Lys Tyr Asp Pro Lys Phe Gln 51 56 61

Val Lys Ala Thr Ile Thr Ala Asp Thr Ser Ser Asn Thr Ala Trp Leu 66 71 76 81

Gln Leu Ser Ser Leu Thr Ser Glu Asp Thr Ala Val Tyr Tyr Cys Ala 86 91 96

Asp Gly Met Trp Val Ser Thr Gly Tyr Ala Leu Asp Phe Trp Gly Gln
101 106 111

Gly Thr Thr Val Thr Val Ser Ser 116 121

- (2) INFORMATION FOR SEQ ID NO:7:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 24 base pairs
    - (B) TYPE: nucleic acid

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1
  - (D) OTHER INFORMATION: /note = "VKIBACK PCR primer"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

#### GACATTCAGC TGACCCAGTC TCCA

24

- (2) INFORMATION FOR SEQ ID NO:8:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 32 base pairs

    - (B) TYPE: nucleic acid (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "VK7BACK PCR primer"
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## TTGAATTCGG AGTTGATGGG AACATTGTAA TG

- (2) INFORMATION FOR SEQ ID NO:9:
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    - (B) TYPE: nucleic acid
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  - (ii) MOLECULE TYPE: cDNA
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    - (B) LOCATION: 1..318
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- (D) OTHER INFORMATION: /note= "pBAG172 insert: HP1/2 light chain variable region"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

AGI Ser 1	TATI	C GTO	S ATO	ACC Thr	CAG Gln	ACI Thr	Pro	Lys	TTC Phe 10	CTG Leu	CTT Leu	GTT Val	TCA Ser	GCA Ala 15	GGA Gly	48
GAC Asp	AGG Arg	GT1	Thr 20	ATA Ile	ACC	TGC Cys	AAG Lys	GCC Ala 25	AGT Ser	CAG Gln	AGT Ser	GTG Val	ACT Thr 30	AAT Asn	GAT Asp	96
GTA Val	GCT Ala	TGG Trp 35	TAC Tyr	CAA Gln	CAG Gln	AAG Lys	CCA Pro 40	GGG Gly	CAG Gln	TCT Ser	CCT Pro	AAA Lys 45	CTG Leu	CTG Leu	ATA Ile	144
TAT Tyr	TAT Tyr 50	GCA Ala	TCC Ser	AAT Asn	CGC Arg	TAC Tyr 55	ACT Thr	GGA Gly	GTC Val	CCT Pro	GAT Asp 60	CGC Arg	TTC Phe	ACT Thr	GGC Gly	192
AGT Ser 65	GGA Gly	TAT Tyr	GGG Gly	ACG Thr	GAT Asp 70	TTC Phe	ACT Thr	TTC Phe	ACC Thr	ATC Ile 75	AGC Ser	ACT Thr	GTG Val	CAG Gln	GCT Ala 80	240
GAA Glu	GAC Asp	CTG Leu	GCA Ala	GTT Val 85	TAT Tyr	TTC Phe	тст Сув	CAG Gln	CAG Gln 90	GAT Asp	TAT Tyr	AGC Ser	TCT Ser	CCG Pro 95	TAC Tyr	288

ACG TTC GGA GGG GGG ACC AAG CTG GAG ATC Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile 100

- (2) INFORMATION FOR SEQ ID NO:10:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 106 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Ser Ile Val Met Thr Gln Thr Pro Lys Phe Leu Leu Val Ser Ala Gly
1 5 10 15

Asp Arg Val Thr Ile Thr Cys Lys Ala Ser Gln Ser Val Thr Asn Asp 20 25 30

Val Ala Trp Tyr Gln Gln Lys Pro Gly Gln Ser Pro Lys Leu Leu Ile 35 40 45

Tyr Tyr Ala Ser Asn Arg Tyr Thr Gly Val Pro Asp Arg Phe Thr Gly

Ser Gly Tyr ly Thr Asp Phe Thr Phe Thr Ile S r Thr Val Gln Ala 70

Glu Asp Leu Ala Val Tyr Phe Cys Gln Gln Asp Tyr Ser Ser Pro Tyr

Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile

- (2) INFORMATION FOR SEQ ID NO:11:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 34 base pairs

    - (B) TYPE: nucleic acid(C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_1
    - (B) LOCATION: 1
    - (D) OTHER INFORMATIO.

R primer"

(xi) SEQUENCE DESCRIPTION:

TGAGGAGACG GTGACCGTGG TCCCTTGGC

34

- (2) INFORMATION FOR SEQ ID NO:12
  - (i) SEQUENCE CHARACTERISTICS
    - (A) LENGTH: 17 base pai

    - (B) TYPE: nucleic acid
      (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /no-

imer"

(xi) SEQUENCE DESCRIPTION: SEQ II

GACATTCAGC TGACCCA

12276

17

(2) INFORMATION FOR SEQ ID NO:13:

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(i) SEQUENCE CHARACTERISTICS:
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- (A) LENGTH: 22 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1
  - (D) OTHER INFORMATION: /note= "VK1FOR PCR primer"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

#### GTTAGATCTC CAGCTTGGTC CC

22

C

G

Α·

- (2) INFORMATION FOR SEQ ID NO:14:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 823 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc feature
    - (B) LOCATION: 1..621
    - (D) OTHER INFORMATION: /note= "VH insert in M13 VHPCR1"
  - (ix) FEATURE:
    - (A) NAME/KEY: exon
    - (B) LOCATION: 261..621
  - (ix) FEATURE:
    - (A) NAME/KEY: sig\_peptide
    - (B) LOCATION: join(122..167, 250..260)
  - (ix) FEATURE:
    - (A) NAME/KEY: mat\_peptide
    - (B) LOCATION: 261..621
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: join(122..167, 250..621)
  - (ix) FEATURE:
    - (A) NAME/KEY: TATA\_signal
    - (B) LOCATION: 38..45

(ix) FEATURE:

		E/KEY: misc_:			
		ATION: 35136			
	(D) OTH	ER INFORMATIO	ON: /note= "C	DR1"	
(ix)	FEATURE:				
` ,		KEY: misc_	feature		
		TION: 40845			
	(D) OTHE	R INFORMATIO	ON: /note= "C	DR2"	
(ix)	FEATURE:			·	
	(A) NAME	KEY: misc_i	eature		
	• •	TION: 55558			
	(D) OTHE	R INFORMATIO	N: /note= "C	DR3"	
(ix)	FEATURE:				
	(A) NAME	/KEY: misc_f	eature		
		TION: (621^62			
	(D) OTHE	R INFORMATIO	N: /note= "s	plice to constant region	n"
(xi)	SEQUENCE	DESCRIPTION:	SEQ ID NO:14	l:	
AAGCTTATG	A ATATCCA		TC T1 C1 TCCT1	A ATATAGGTTT GTCTATACCA	
					60
CAAACAGAA	A AACATGA	GAT CACAGTTC	TC TCTACAGTT	A CTGAGCACAC AGGACCTCAC	120
				STA GCA ACA GCT ACA G	167
				/al Ala Thr Ala Thr	
-19		15	-10	-5	
GTAAGGGGC	T CACAGTA	GCA GGCTTGAG	GT CTGGACATA1	ATATGGGTGA CAATGACATC	227
CACTTTGCC	T TTCTCTC	TAC AG GT G	TO CAG TOO C	AG GTC CAA CTG CAG GAG	
00111000	1 1101010			In Val Gln Leu Gln Glu	278
			-3	1 S	
			•	1	
AGC GGT C	CA GGT CT1	GTG AGA CC	T AGC CAG ACC	CTG AGC CTG ACC TGC	326
Ser Gly P	co Gly Leu	Val Arg Pro	Ser Gln Thr	Leu Ser Leu Thr Cys	320
-	10		15	20	
				20	
ACC GTG TO	T GGC AGC	ACC TTC AG	AGC TAC TGG	ATG CAC TGG GTG AGA	374
Thr Val Se	er Gly Ser	Thr Phe Sea	Ser Tyr Tro	Met His Trp Val Arg	3/4
2	25	30	,	35	
				•	
CAG CCA CC	T GGA CGA	GGT CTT GAG	TGG ATT GGA	AGG ATT GAT CCT AAT	422
Gln Pro Pr	o Gly Arg	Gly Leu Glu	Trp Ile Gly	Arg Ile Asp Pro Asn	
40		45	- <b>-</b>	50	
AGT GGT GG	T ACT AAG	TAC AAT GAG	AAG TTC AAG	AGC AGA GTG ACA ATG	470
				THE THE PART OF THE PARTY IN THE	
Ser Gly Gl	y Thr Lvs	Tyr Asn Glu	LVS Phe Lvs	Ser Arg Val Thr Mot	470
Ser Gly Gl	y Thr Lys	Tyr Asn Glu	Lys Phe Lys 65	Ser Arg Val Thr Met	470

CTG Leu	GTA Val	GAC Asp	ACC Thr	AGC Ser 75	AAG Lys	AAC Asn	CAG Gln	TTC Phe	AGC Ser 80	CTG Leu	AGA Arg	CTC Leu	AGC Ser	AGC Ser 85	GTG Val	518
ACA Thr	GCC Ala	GCC Ala	GAC Asp 90	ACC Thr	GCG Ala	GTC Val	TAT Tyr	TAT Tyr 95	TGT Cys	GCA Ala	AGA Arg	TAC Tyr	GAT Asp 100	TAC Tyr	TAC Tyr	566
GGT Gly	AGT Ser	AGC Ser 105	TAC Tyr	TTT Phe	GAC Asp	TAC Tyr	TGG Trp 110	GGC Gly	CAA Gln	GGG Gly	ACC Thr	ACG Thr 115	GTC Val	ACC Thr	GTC Val	614
	TCA Ser 120	G														621
(2)	INFO	RMAT	NOI	FOR	SEQ	ID N	10:15	:								
	(	i) s		LEN TYF	GTH: PE: 8	ACTE 139 mino	ami aci	ino a id		3						
	(i	i) ł	OLEC	CULE	TYPE	: pr	otei	in '								
	(x	i) S	EQUE	NCE	DESC	RIPT	: NOI	SEÇ	Z ID	NO:	5:					
Met -19	Gly	Trp	Ser	Сув -15	Ile	Ile	Leu	Phe	Leu -10	Val	Ala	Thr	Ala	Thr -5		
Val	His	Ser	Gln 1	Val	Gln	Leu	Gln 5	Glu	Ser	Gly	Pro	Gly 10	Leu	Val	Arg	
Pro	Ser 15	Gln	Thr	Leu	Ser	Leu 20	Thr	Сув	Thr	Val	Ser 25	Gly	Ser	Thr	Phe	
Ser 30	Ser	Tyr	Trp	Met	His 35	Trp	Val	Arg	Gln	Pro 40	Pro	Gly	Arg	Gly	Leu 45	
Glu	Trp	Ile	GJA	Arg 50	Ile	Asp	Pro	Asn	Ser 55	Gly	Gly	Thr	Lys	Tyr 60	Asn	
Glu	Lys	Phe	Lys ·65	Ser	Arg	Val	Thr	Met 70	Leu	Val	увр	Thr	Ser 75	Lys	Asn	٠
Gln	Phe	Ser 80	Leu	Arg	Leu	Ser	Ser 85	Val	Thr	Ala	Ala	<b>Asp</b> 90	Thr	Ala	Val	
Tyr	Tyr 95	Сув	Ala	Arg	Tyr	Asp 100		Tyr	Gly	Ser	Ser 105	Tyr	Phe	Asp	Tyr	

Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser 110 120

- (2) INFORMATION FOR SEQ ID NO:16:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 594 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
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  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
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    - (B) LOCATION: 273..594
  - (ix) FEATURE:
    - (A) NAME/KEY: sig\_peptide
    - (B) LOCATION: join(134..179, 262..272)
  - (ix) FEATURE:
    - (A) NAME/KEY: mat peptide
    - (B) LOCATION: 273..594
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: join(134..179, 262..594)
  - (ix) FEATURE:
    - (A) NAME/KEY: TATA\_signal
    - (B) LOCATION: 50..57
  - (ix) FEATURE:
    - (A) NAME/KEY: misc feature
    - (B) LOCATION: 342..374
    - (D) OTHER INFORMATION: /note = "CDR1"
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 420..440
    - (D) OTHER INFORMATION: /note= "CDR2"
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 537..563
    - (D) OTHER INFORMATION: /note= "CDR3"

/ i - \	FEATURE	•
	FEMIURE	2

- (A) NAME/KEY: misc\_feature
  (B) LOCATION: (594^595)
- (D) OTHER INFORMATION: /note= "splice to constant region"

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CT	CTTA	AACT	TCA	AGCT	TAT :	GAAT	ATGC	AA A	CCT	CTGA	A TC	raca:	iggi	AAAT	TATAGG	T 60
TT	GTCT	ATAC	CAC	AAAC	AGA :	AAAA	CATG	AG A	CAC	GTT	TC	CTAC	CAGT	TACT	rgagca	ic 120
AC	AGGA	CCTC	ACC		Gly		AGC Ser									169
		T AC		GTA	AGGGG	CT (	CACAG	TAGO	A GG	CTTG	aggt	CTG	GACA	TAT		219
ATĀ	TGG	STGA	CAA	rgaci	ATC C	CACTI	TGCC	T TI	CTCT	CCAC			GTC Val -3			272
GAC Asp I	ATC Ile	CAC Glr	CTC	ACC Thr	CAG Gln	AGC Ser	Pro	AGC Ser	AGC Ser 10	CTG Leu	AGC Ser	GCC Ala	AGC Ser	GTG Val 15	GGT Gly	320
GAC Asp	AGA Arg	GTG Val	ACC Thr 20	Ile	ACC Thr	TGT Cys	AGA Arg	GCC Ala 25	AGC Ser	GCT	AAC Asn	ATC Ile	CAC His	AAC Asn	TAC Tyr	368
CTG Leu	GCT Ala	TGG Trp 35	TAC	CAG Gln	CAG Gln	AAG Lys	CCA Pro 40	GGT Gly	AAG Lys	GCT Ala	CCA Pro	AAG Lys 45	CTG Leu	CTG Leu	ATC Ile	416
TAC Tyr	TAC Tyr 50	ACC Thr	ACC Thr	ACC Thr	CTG Leu	GCT Ala 55	GAC Asp	GGT Gly	GTG Val	CCA Pro	AGC Ser 60	AGA Arg	TTC Phe	AGC Ser	GGT Gly	464
AGC Ser 65	GGT Gly	AGC Ser	GGT Gly	ACC Thr	GAC Asp 70	TTC Phe	ACC Thr	TTC Phe	ACC Thr	ATC Ile 75	AGC Ser	AGC Ser	CTC Leu	CAG Gln	CCA Pro 80	512
AG lu	GAC Asp	ATC Ile	GCC Ala	ACC Thr 85	TAC Tyr	TAC Tyr	TGC Cys	CAG Gln	CAC Him 90	TTC Phe	TGG Trp	AGC Ser	ACC Thr	CCA Pro 95	AGG Arg	. 560
CG hr	Phe	Gly	CAA Gln 100	Gly	ACC Thr	AAG Lys	GTG Val	GAA Glu	ATC Ile	AAA Lys	С					594

- (2) INFORMATION FOR SEQ ID NO:17:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 126 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Met Gly Trp Ser Cys Ile Ile Leu Phe Leu Val Ala Thr Ala Thr Gly
-19 -15 -10 -5

Val His Ser Asp Ile Gln Leu Thr Gln Ser Pro Ser Ser Leu Ser Ala

1 5 10

Ser Val Gly Asp Arg Val Thr Ile Thr Cys Arg Ala Ser Gly Asn Ile
15 20 25

His Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys 30 40 45

Leu Leu Ile Tyr Tyr Thr Thr Thr Leu Ala Asp Gly Val Pro Ser Arg
50 55 60

Phe Ser Gly Ser Gly Thr Asp Phe Thr Phe Thr Ile Ser Ser 65 70 75

Leu Gln Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln His Phe Trp Ser 80 85 90

Thr Pro Arg Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys 95 100 ' 105

- (2) INFORMATION FOR SEQ ID NO:18:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 54 base pairs
      - (B) TYPE: nucleic acid
      - (C) STRANDEDNESS: Bingle
      - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "DNA sequence of 598 oligonucleotide"

	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:18:	
TG	TCTCA	CCC AGTGCATATA GGTGTCTTTA ATGTTGAAGC CAGACACGCT GCAG	54
(2)	INF	ORMATION FOR SEQ ID NO:19:	
	(i)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 71 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(īī)	HOLECULE TYPE: CDNA	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "DNA sequence of 599 oligonucleotide"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:19:	
CAG	CATTG	TC ACTCTGACCT GGAACTTCGG GTCATATTTA GTATCGCCAC TCGCAGGATC	60
AAT	CCTTC	CA A	71
(2)	INFO	RMATION FOR SEQ ID NO:20:	,,
(2)	11120	MATION FOR SEQ ID NO:20:	
	(i)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 70 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "DNA sequence of 600 oligonucleotide"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:20:	
GGTC	CCTTG	G CCCCAGAAGT CCAGAGCATA TCCCGTTGAT ACCCACATTC CGTCTGCACA	60
ATAA	TAGAC	<b>:c</b>	70
(2)	INFOR	MATION FOR SEQ ID NO:21:	
		SECUENCE CHARACTERISTICS.	

(A) LENGTH: 51 base pairs

(ii) MOLECULE TYPE: cDNA

(B) LOCATION: 1

(A) NAME/KEY: misc\_feature

oligonucleotide\*

(D) OTHER INFORMATION: /note = "DNA sequence of 607

(ix) FEATURE:

			<b>-87</b> -	
		(B)	TYPE: nucleic acid	
			STRANDEDNESS: single	
			TOPOLOGY: linear	
	(ii)	MOL:	ECULE TYPE: cDNA	
	(ix)	FEA:	TURE:	
		(A)	NAME/KEY: misc_feature	
			LOCATION: 1	
		(D)	OTHER INFORMATION: /note= "DNA sequence of 605 oligonucleotide"	
	(xi)	SEQU	JENCE DESCRIPTION: SEQ ID NO:21:	
TCC	CTTG	GCC GA	LACGTGTAC GGAGAGCTAT AATCCTGCTG GCAGTAGTAG G	51
(2)	INFO	RMATI	ON FOR SEQ ID NO:22:	
	(i)	SEQU	ENCE CHARACTERISTICS:	
			LENGTH: 52 base pairs	
			TYPE: nucleic acid	
			STRANDEDNESS: single	
		(D)	TOPOLOGY: linear	
	(ii)	MOLE	CULE TYPE: CDNA	
	(ix)	FEAT		
		(A)	NAME/KEY: misc_feature	
			LOCATION: 1	
		(D)	OTHER INFORMATION: /note= "DNA sequence of 606	
			oligonucleotide"	
	(xi)	SEQUI	ENCE DESCRIPTION: SEQ ID NO:22:	
ATC	rgctt	GG GCI	ACACCAGT GTAGCGATTG GATGCATAGT AGATCAGCAG CT	52
(2)	INFO	CITAMS	ON FOR SEQ ID NO:23:	
	(i)	SEQUE	ENCE CHARACTERISTICS:	
		(A)	LENGTH: 61 base pairs	
		(B)	TYPE: nucleic acid	
			STRANDEDNESS: single	
		(D)	TOPOLOGY: linear	

	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:23:	
TC	TGCTG	STA CCAAGCTACA TCATTAGTCA CACTCTGACT GGCCTTACAG GTGATGGTCA	60
С			61
(2)	INFO	DRMATION FOR SEQ ID NO:24:	
<b>()</b>			
	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 17 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "DNA sequence OLIGO 10 oligonucleotide"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:24:	
GTA	AAACG	AC GGCCAGT	17
(2)	INFO	RMATION FOR SEQ ID NO:25:	
	(i)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 22 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "DNA sequence of OLIGO 385 oligonucleotide"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:25:	
GCG	GGCCT	CT TCGCTATTACGC	22
(2)	INFO	RMATION FOR SEQ ID NO:26:	
	(i)	SEQUENCE CHARACTERISTICS:	
	\-/	(A) LENGTH: 16 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	

•	(D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: cDNA	
(ix)	FEATURE:  (A) NAME/KEY: misc_feature  (B) LOCATION: 1  (D) OTHER INFORMATION: /note= "DNA sequence of OLIGO 11 oligonucleotide"	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:26:	
AACAGCTA	TG ACCATG	16
(2) INFO	RMATION FOR SEQ ID NO:27:	
(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 22 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: cDNA	
(ix)	<pre>FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1 (D) OTHER INFORMATION: /note= "DNA sequence of OLIGO 391</pre>	
( <b>x</b> i)	SEQUENCE DESCRIPTION: SEQ ID NO:27:	
CTCTCTCAC	GG GCCAGGCGGT GA	22
(2) INFO	RMATION FOR SEQ ID NO:28:	
(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 429 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: cDNA	
(ix)	FEATURE:  (A) NAME/KEY: sig_peptide  (B) LOCATION: 157	
(ix)	FEATURE:  (A) NAME/KEY: mat_peptide  (B) LOCATION: 58.429	

(ix) FEAT	URE:
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- (A) NAME/KEY: CDS
- (B) LOCATION: 1..429

#### (ix) FEATURE:

- (A) NAME/KEY: misc\_feature
- (B) LOCATION: 1
- (D) OTHER INFORMATION: /note= "pMDR1019 insert: Stage 1 heavy chain variable region "

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

	. Asi		ACC Thr		Arg					Leu					Gly	48
GCC Ala	CAC His	Ser	CAG Gln 1	GTC Val	CAA Gln	CTG Le	CAG Glm 5	GAG Glu	AGC Ser	GGT	CCA Pro	GGT Gly 10	CTT Leu	GTG Val	AGA Arg	96
			ACC Thr													144
AAA Lys 30	GAC Asp	ACC Thr	TAT Tyr	ATG Met	CAC His 35	TGG Trp	GTG Val	AGA Arg	CAG Gln	CCA Pro 40	CCT Pro	GGA Gly	CGA Arg	GGT Gly	CTT Leu 45	192
GAG Glu	TGG Trp	ATT Ile	GGA Gly	AGG Arg 50	ATT Ile	GAT Asp	CCT Pro	GCG Ala	AGT Ser 55	GGC Gly	GAT Asp	ACT Thr	AAA Lys	TAT Tyr 60	GAC Asp	240
CCG Pro	AAG Lys	TTC Phe	CAG Gln 65	GTC Val	AGA Arg	GTG Val	ACA Thr	ATG Met 70	CTG Leu	GTA Val	GAC Asp	ACC Thr	AGC Ser 75	AAG Lys	AAC Asn	288
CAG Gln	TTC Phe	AGC Ser 80	CTG Leu	AGA Arg	CTC Leu	AGC Ser	AGC Ser 85	GTG Val	ACA Thr	GCC Ala	GCC Ala	GAC Asp 90	ACC Thr	GCG Ala	GTC Val	336
TAT Tyr	TAT Tyr 95	TGT Cys	GCA Ala	GAC Asp	GGA Gly	ATG Met 100	TGG Trp	GTA Val	TCA Ser	ACG Thr	GGA Gly 105	TAT Tyr	GCT Ala	CTG Leu	GAC Asp	384
TTC Phe 110	TGG Trp	GGC Gly	CAA Gln	Gly	ACC Thr 115	ACG :	GTC Val	ACC Thr	GTC Val	TCC Ser 120	TCA Ser	GGT Gly	GAG Glu	TCC Ser		429

# (2) INFORMATION FOR SEQ ID NO:29:

### (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 143 amino acids

- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

Met Asp Trp Thr Trp Arg Val Phe Cys Leu Leu Ala Val Ala Pro Gly
-19 -15 -10 -5

Ala His Ser Gln Val Gln Leu Gln Glu Ser Gly Pro Gly Leu Val Arg

1 5 10

Pro Ser Gln Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Phe Asn Ile
15 20 25

Lys Asp Thr Tyr Met His Trp Val Arg Gln Pro Pro Gly Arg Gly Leu 30 40 45

Glu Trp Ile Gly Arg Ile Asp Pro Ala Ser Gly Asp Thr Lys Tyr Asp
50 55 60

Pro Lys Phe Gln Val Arg Val Thr Met Leu Val Asp Thr Ser Lys Asn 65 70 75

Gln Phe Ser Leu Arg Leu Ser Ser Val Thr Ala Ala Asp Thr Ala Val 80 85 90

Tyr Tyr Cys Ala Asp Gly Met Trp Val Ser Thr Gly Tyr Ala Leu Asp 95 100 105

Phe Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Gly Glu Ser 110 115 120

- (2) INFORMATION FOR SEQ ID NO:30:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 386 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: sig\_peptide
    - (B) LOCATION: 1..57
  - (ix) FEATURE:
    - (A) NAME/KEY: mat\_peptide
    - (B) LOCATION: 58..386

										-9	2 –					
	(i:	() F	EATU	RE:												
			(A)	NAME	KEY:	CD	s									
				LOCA:												
	(ix	) F	EATU	RE:												
		(	(A) 1	NAME/	KEY:	mi	sc fe	Batu	ce c							
		(	(B) 1	LOCAT	CION	: 1										
							TION	₹: /n	ote=	· "ni	RAGIO	ю .			(1 (DQ1	
				11	.ght	chai	n va	riat	le r	ecic	) n =	0 11	IBEL I	-: A1	ci (DÖI	_)
											•••					
	(xi	) SE	EQUEN	ice d	ESCR	IPTI	ON:	SEQ	ID N	10:30	:	-				
ATG	GGI	TGG	TCC	TGC	ATC	ATC	CTG	TTC	CTG	CTT	CCT	* ACC			GGT	
Met	Gly	Trp	Ser	Cys	Ile	Ile	Leu	Phe	Leu	Val	Ala	The	. GC1	ACC	Gly	
19	)			-15	•				-10	)	*****		VIA	THE		
GTT	CAC	TCC	GAC	ATC	CAG	CTG	ACC	CAG	AGC	CCA	AGC	AGC	CTG	AGC	GCC	
Val	His	Ser	Asp	Ile	Gln	Leu	Thr	Gln	Ser	Pro	Ser	Ser	Leu	Ser	Ala	
			1				5					10		JEI	VIG	
AGC	GTG	GGT	GAC	AGA	GTG	ACC	ATC	ACC	TGT	AAG	GCC	AGT	CAG	AGT	GTG	
Ser	447	Gly	Asp	Arg	Val	Thr	Ile	Thr	Сув	Lys	Ala	Ser	Gln	Ser	Val	
	15					20				_	25					
HCT Bloom	AAT	GAT	GTA	GCT	TGG	TAC	CAG	CAG	AAG	CCA	GGT	AAG	GCT	CCA	AAG	
inr	ABN	ASD	Val	Ala	T	Tree	C1-	~1-	•			_				

-1	9			-1	5				-1	0				-	_ '	Y	
GT1 Val	CA(	C TCC	GAG As <sub>1</sub>	P Ile	C CA	G CTC	ACC Thi	C CAC Glr	AG0 Ser	C CCI	A AGO Ser	Ser 10	Leu	G AGO	C GCC	C A	96
AGC Ser	Val	GIY	GAC Asp	AGP Arg	GTC Val	ACC Thr 20	ATC Ile	ACC Thr	TG1	AAG Lys	GCC Ala 25	AGT Ser	CAG Gln	AG1 Ser	GTC Val	<b>;</b>	144
ACT Thr 30	AAT Asn	GAT Asp	GTA Val	GCT Ala	TGG Trp 35	Tyr	CAG Gln	CAG Gln	AAG Lys	CCA Pro 40	GGT Gly	AAG Lys	GCT Ala	CCA Pro	AAG Lys 45	;	192
CTG Leu	CTG Leu	ATC Ile	TAC Tyr	TAT Tyr 50	GCA Ala	TCC Ser	AAT Asn	CGC Arg	TAC Tyr 55	ACT Thr	GGT Gly	GTG Val	CCA Pro	AGC Ser 60	AGA Arg		240
TTC Phe	AGC Ser	GGT Gly	AGC Ser 65	GGT Gly	AGC Ser	GGT Gly	ACC Thr	GAC Asp 70	TTC Phe	ACC Thr	TTC Phe	ACC Thr	ATC Ile 75	AGC Ser	AGC Ser		288
CTC Leu	CAG Gln	CCA Pro 80	GAG Glu	GAC Asp	ATC Ile	GCC Ala	ACC Thr 85	TAC Tyr	TAC Tyr	TGC Cys	CAG Gln	CAG Gln 90	GAT Asp	TAT Tyr	AGC Ser		336
TCT (	Pro 95	TAC . Tyr	ACG Thr	TTC Phe	GCC Gly	CAA Gln 100	GCG	ACC Thr	AAG Lys	Val	GAA Glu 105	ATC :	AAA Lys	CGT Arg	AAG Lys	TG	386

# (2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 128 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

Met Gly Trp Ser Cys Ile Ile Leu Phe Leu Val Ala Thr Ala Thr Gly

Val His Ser Asp Ile Gln Leu Thr Gln Ser Pro Ser Ser Leu Ser Ala

Ser Val Gly Asp Arg Val Thr Ile Thr Cys Lys Ala Ser Gln Ser Val

Thr Asn Asp Val Ala Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys

Leu Leu Ile Tyr Tyr Ala Ser Asn Arg Tyr Thr Gly Val Pro Ser Arg

Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Phe Thr Ile Ser Ser

Leu Gln Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln Gln Asp Tyr Ser

Ser Pro Tyr Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg Lys

- (2) INFORMATION FOR SEQ ID NO:32:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 429 base pairs

    - (B) TYPE: nucleic acid(C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:

0

36

- (A) NAME/KEY: sig\_peptide
- (B) LOCATION: 1..57
- (ix) FEATURE:
  - (A) NAME/KEY: mat\_peptide
  - (B) LOCATION: 58..429
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 1..429
- (ix) FEATURE:
  - (A) NAME/KEY: misc feature

- (B) LOCATION: 1
- (D) OTHER INFORMATION: /note= "pMDR1028 insert: Stage 2 heavy chain variable region"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

	Asp	ACC Thr		Arg			Leu			Gly	48
		CAG Gln 1									96
		ACC Thr							 		144
		TAT Tyr									192
		GGA Gly									240
		CAG Gln 65									288
		CTG Leu									336
		GCA Ala									384
TTC Phe 110			Gly								429

- (2) INFORMATION FOR SEQ ID NO:33:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 143 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

Met Asp Trp Thr Trp Arg Val Phe Cys Leu Leu Ala Val Ala Pro Gly
-19
-15
-5

Ala His Ser Gln Val Gln Leu Gln Glu Ser Gly Pro Gly Leu Val Arg

Pro Ser Gln Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Phe Asn Ile 15 20 25

Lys Asp Thr Tyr Met His Trp Val Arg Gln Pro Pro Gly Arg Gly Leu 30 40 45

Glu Trp Ile Gly Arg Ile Asp Pro Ala Ser Gly Asp Thr Lys Tyr Asp 50 55 60 .

Pro Lys Phe Gln Val Arg Val Thr Met Leu Val Asp Thr Ser Ser Asn 65 70 75

Gln Phe Ser Leu Arg Leu Ser Ser Val Thr Ala Ala Asp Thr Ala Val 80 85 90

Tyr Tyr Cys Ala Asp Gly Met Trp Val Ser Thr Gly Tyr Ala Leu Asp 95 100 105

Phe Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Gly Glu Ser 110 120

- (2) INFORMATION FOR SEQ ID NO:34:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 24 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "DNA sequence of 684 oligonucleotide"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

AGACACCAGC AGCAACCAGT TCAG

- (2) INFORMATION FOR SEQ ID NO:35:
  - (i) SEQUENCE CHARACTERISTICS:

			-90-	
		(A) 1	LENGTH: 24 base pairs	
		(B) 2	TYPE: nucleic acid	
		(c) s	STRANDEDNESS: single	
		(D) 2	ropology: linear	
	(ii)	MOLECT	JLE TYPE: CDNA	
	(4)	FEATUR	DP.	
	(LX)		NAME/KEY: misc_feature	
			LOCATION: 1	
		(a)	OTHER INFORMATION: /note= "DNA sequence of 683	
			oligonucleotide"	
	(xi)	SEQUEN	HCE DESCRIPTION: SEQ ID NO:35:	
GA	ACTGG	TT GCT	GCTGGTG TCTA	24
2)	INFO	RMATION	FOR SEQ ID NO:36:	
	(i)	SEQUEN	NCE CHARACTERISTICS:	
	(-)		LENGTH: 37 base pairs	
			TYPE: nucleic acid	
			STRANDEDNESS: single	
		(D) 1	TOPOLOGY: linear	
	(ii)	MOLECU	JLE TYPE: cDNA	
	(ix)	FEATUR	æ:	
		(A) h	NAME/KEY: misc_feature	
			LOCATION: 1	
		(D) C	OTHER INFORMATION: /note = "DNA sequence of 713 oligonucleotide"	
	(xi)	SEQUEN	NCE DESCRIPTION: SEQ ID NO:36:	-
.CC	AGCAG	CA ACAC	CAGCCTG GCTGAGACTC AGCAGCG	37
(2)	INFO	RMATION	N FOR SEQ ID NO:37:	
	(i)	SEQUEN	NCE CHARACTERISTICS:	
		(A) I	LENGTH: 38 base pairs	
			TYPE: nucleic acid	
			STRANDEDNESS: single	
		(D) 7	TOPOLOGY: linear	
	(ii)	MOLECU	JLE TYPE: CDNA	
	(ix)	FEATUR	æ:	
			NAME/KEY: misc_feature	
			LOCATION: 1	
		(D) (	THER INFORMATION: /note= "DNA sequence of 716	

oligonucleotide"

192

										-97-	-						
	(xi)	SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0:37	•						
GCI	GAGT	CTC I	AGCC	AGGC	TG T	GTTG	CTGC	T GG	TGTC	GA							3
(2)	INF	ORMAT	rion	FOR	SEQ	ID	NO:3	8:									
	<b>(i)</b>	(A (B (C	) L ) T	engt Ype: Tran	HARA H: 4: nuc DEDN OGY:	29 b leic ESS:	ase aci sin	pair d	8								
	(ii)	MOI	LECU	LE T	YPE:	CDN	A										
	(ix)		) N	AME/F	ŒY: ION:			tide									
	(ix)	٠,	) N	ame/f	ŒY: ION:	•		tide									
	(ix)	` '	) N2	ame/r	EY: ION:		29										
	(ix)	(A) (B)	L	AME/R OCAT: THER	ION: INFO	l ORMA'	- TION	atur :/nc reg	te=	"pBI	AG184	ine	sert:	: ST	A₩ he	<b>av</b> y	
	(xi)	SEQ	UENC	CE DI	ESCR	PTI	эи:	SEQ	ID NO	38:							
		TGG . Trp													Gly		4
		TCC (															96
		CAG :															14

AAA GAC ACC TAT ATG CAC TGG GTG AGA CAG CCA CCT GGA CGA GGT CTT

Lys Asp Thr Tyr Met His Trp Val Arg Gln Pro Pro Gly Arg Gly Leu

										<b>-98</b> -	•				
						<b></b>			200		C2.T	3 CT		ThT	GAC
GAG	TGG	ATT	GGA	AGG	ATT	GAT	Pro	GCG Ala	Ser	Glv	ARD	Thr	Lvs	Tvr	Asp
GIU	IIP	116	Gry	50					55	,			-4-	60	
				_											
CCG	AAG	TTC	CAG	GTC	AGA	GTG	ACA	ATG	CTG	GTA	GAC	ACC	AGC	AGC	AAC
Pro	Lys	Phe		Val	Arg	Val	Thr	Met 70	Leu	Val	Asp	Thr	Ser 75	ser	ASn
			65					70							
ACA	GCC	TGG	CTG	AGA	CTC	AGC	AGC	GTG	λCλ	GCC	GCC	GAC	ACC	GCG	GTC
Thr	Ala		Leu	Arg	Leu	Ser		Val	Thr	Ala	Ala		Thr	Ala	Val
		80					85					90			
ТАТ	TAT	TGT	GCA	GAC	GGA	ATG	TGG	GTA	TCA	ACG	GGA	TAT	GCT	CTG	GAC
Tyr	Tyr	Сув	Ala	Asp	Gly	Met	Trp	Val	Ser	Thr	Gly	Tyr	Ala	Leu	Asp
	95					100					105				
mmc	TCC	ccc	CAA	ccc	ACC	ACG.	GTC	ACC	GTC	TCC	TCA	GGT	GAG	TCC	
Phe	Trp	Gly	Gln	Gly	Thr	Thr	Val	Thr	Val	Ser	Ser	Gly	Glu	Ser	
110	•	•		•	115					120					
40.								١.							
(2)	INF	ORMA:	LION	FOR	SEQ	י עד	NO: 39	•							
	(	(i)						rics							
			7.7					ino a	acid	5					
			(B) (D)				o ac: linea								
			(0)	101											
	(i	i) 1	OLE	CULE	TYPI	E: pı	rote	in							
					220	70 T D	TON	. ce/	. TD	NO.	20.				
	( <b>x</b>	i) :	SEQUI	ENCE	DESC	JRIP.	TION	: SE(	מז ב	NO:	) 7 :		-		
Met	Asp	Trp	Thr	Trp	Arg	Val	Phe	Сув	Leu	Leu	Ala	Val	Ala	Pro	Gly
-19	_			-15	•				-10					-5	
•••	•••		<b>-</b> 01-	**- 1	<b>61</b> -	T	c1-	<b>~</b> 1	<b>5</b> 0-	C1**	Dro	Gly	TAU	V=1	Ara
ATE	HIB	Ser	GIn 1	ATT	GIN	reu	5	Glu	Ser	Gry	FLO	10	Dea	. 441	nry
			_				_								
Pro	Ser	Gln	Thr	Leu	Ser		Thr	Cys	Thr	Val		Gly	Phe	Asn	Ile
	15					20					25				
Tue	Aen	Th~	ጥነታም	Met	Hie	Tro	Val	Arg	Gln	Pro	Pro	Glv	Ara	Glv	Leu
30	vaħ	****	-1-	1.46	35	2	***	9		40		1	9	1	45

Glu Trp Ile Gly Arg Ile Asp Pro Ala Ser Gly Asp Thr Lys Tyr Asp

Pro Lys Phe Gln Val Arg Val Thr Met Leu Val Asp Thr Ser Ser Asn

Thr Ala Trp Leu Arg Leu Ser Ser Val Thr Ala Ala Asp Thr Ala Val

Tyr Tyr Cys Ala Asp Gly Met Trp Val Sor Thr Gly Tyr Ala Leu Asp 95 100 105

Phe Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Gly Glu Ser 110 120

- (2) INFORMATION FOR SEQ ID NO:40:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 40 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "DNA sequence of 706 oligonucleotide"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

### AGTTCCAGGT CAAAGCGACA ATTACGGCAG ACACCAGCAA

40

- (2) INFORMATION FOR SEQ ID NO:41:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 40 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "DNA sequence of 707 oligonucleotide"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

# CTTGCTGGTG TCTGCCGTAA TTGTCGCTTT GACCTGGAAC

- (2) INFORMATION FOR SEQ ID NO:42:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 429 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single

240

288

336

15

30

	(D) TOPOLOGY: linear															
		([	) T	OPOL	OGY:	line	ar									
	(ii)	MO	LECU	LE T	YPE:	CDN	A									
	(ix)	FE	ATURI	E:												
		(A	) N	AME/K	EY:	sig	_pept	tide								
		(E	) L	OCAT:	ION:	157	7									
	(ix)	FE	ATURI	E:												
	•	(A	) N	AME/K	EY:	mat	pept	ide								
						584										
	(ix)	FE	ATURI	E:												
	•		) N		EY:	CDS										•
						142	9									
	(ix)	FEATURE:														
	• •	(A	) NI	AME/R	EY:	misc	: fea	ature	3							
			) L				_									
						ORMAT	CION	: /no	te=	"pB	AG183	in	ert:	KA:	TAS	
		•				chair										
	(x1)	SE	QUENC	CE DI	ESCR	PTIC	on: S	SEQ 1	D NO	2:42:			•			
ATG	GAC	TGG	ACC	TGG	AGG	GTC	TTC	TGC	TTG	CTG	GCT	GTA	GCA	CCA	GGT	48
						Val										
-19		•		-15	•			-	-10					-5	-	
GCC	CAC	TCC	CAG	GTC	CAA	CTG	CAG	GAG	AGC	GGT	CCA	GGT	CTT	GTG	AGA	96
						Leu										
nia	UIB	261	1	Val	GIII	Dea	5	014	501	<b></b> 3		10			•9	
			1				,									
						CTG										144
Pro	Ser	Gln	Thr	Leu	Ser	Leu	Thr	Сув	Thr	Val	Ser	Gly	Phe	Asn	Ile	

20

50

65

80

AAA GAC ACC TAT ATG CAC TGG GTG AGA CAG CCA CCT GGA CGA GGT CTT

Lys Asp Thr Tyr Met His Trp Val Arg Gln Pro Pro Gly Arg Gly Leu

GAG TGG ATT GGA AGG ATT GAT CCT GCG AGT GGC GAT ACT AAA TAT GAC

Glu Trp Ile Gly Arg Ile Asp Pro Ala Ser Gly Asp Thr Lys Tyr Asp

CCG AAG TTC CAG GTC AAA GCG ACA ATT ACG GCA GAC ACC AGC AGC AAC

Pro Lys Phe Gln Val Lys Ala Thr Ile Thr Ala Asp Thr Ser Ser Asn

CAG TTC AGC CTG AGA CTC AGC AGC GTG ACA GCC GCC GAC ACC GCG GTC

Gln Phe Ser Leu Arg Leu Ser Ser Val Thr Ala Ala Asp Thr Ala Val

85

55

38

36

-101-

			TGG Trp				GAC Asp	384
			GTC Val					429

- (2) INFORMATION FOR SEQ ID NO:43:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 143 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

Met Asp Trp Thr Trp Arg Val Phe Cys Leu Leu Ala Val Ala Pro Gly -19 -15 -10 -5

Ala His Ser Gln Val Gln Leu Gln Glu Ser Gly Pro Gly Leu Val Arg
1 5 10

Pro Ser Gln Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Phe Asn Ile
15 20 25

Lys Asp Thr Tyr Met His Trp Val Arg Gln Pro Pro Gly Arg Gly Leu 30 45

Glu Trp Ile Gly Arg Ile Asp Pro Ala Ser Gly Asp Thr Lys Tyr Asp
50 55 60

Pro Lys Phe Gln Val Lys Ala Thr Ile Thr Ala Asp Thr Ser Ser Asn 65 70 75

Gln Phe Ser Leu Arg Leu Ser Ser Val Thr Ala Ala Asp Thr Ala Val 80 85 90

Tyr Tyr Cys Ala Asp Gly Met Trp Val Ser Thr Gly Tyr Ala Leu Asp 95 100 105

Phe Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Gly Glu Ser 110 120

- (2) INFORMATION FOR SEQ ID NO:44:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 36 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single

(D) TOPOLOGY: line	a.I
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- (ii) MOLECULE TYPE: CDNA
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1
  - (D) OTHER INFORMATION: /note= "DNA sequence of 768 oligonucleotide"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

# CTCAGCAGCG TGACATCTGA GGACACCGCG GTCTAT

36

- (2) INFORMATION FOR SEQ ID NO:45:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 36 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: CDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "DNA sequence of 769 oligonucleotide"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

ATAGACCGCG GTGTCCTCAG ATGTCACGCT GCTGAG

- (2) INFORMATION FOR SEQ ID NO:46:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 372 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
    - (ii) MOLECULE TYPE: cDNA
    - (ix) FEATURE:
      - (A) NAME/REY: misc\_feature
      - (B) LOCATION: 1..372
      - (D) OTHER INFORMATION: /note= "pBAG207 insert: SSE heavy chain variable region\*
    - (ix) FEATURE:
      - (A) NAME/KEY: mat\_peptide

(B	) LOCATION: 1	l372

### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..372

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

CAC Gl:	G GTC	CAI L Glr	A CTG	CAC Glr	GAG Glu	AGC Ser	GGT	Pro	GGT Gly 10	CTT Leu	GTG Val	AGA Arg	CCT	AGC Ser 15	CAG Gln	48
ACC Thr	CTC	AGC Ser	CTG Leu 20	ACC Thr	TGC Cys	ACC Thr	GTG Val	Ser 25	GCC	TTC Phe	AAC Asn	ATT Ile	AAA Lys 30	GAC Asp	ACC Thr	<b>9</b> 6
TAT Tyr	ATG Het	CAC His	TGG	GTG Val	AGA Arg	CAG Gln	CCA Pro 40	CCT Pro	GGA Gly	CGA Arg	GGT Gly	CTT Leu 45	GAG Glu	TGG Trp	ATT Ile	144
GGA Gly	AGG Arg 50	ATT	GAT Asp	CCT Pro	GCG Ala	AGT Ser 55	GGC Gly	GAT Asp	ACT Thr	AAA Lys	TAT Tyr 60	GAC Asp	CCG Pro	AAG Lys	TTC Phe	192
CAG Gln 65	GTC Val	AGA Arg	GTG Val	ACA Thr	ATG Met 70	CTG Leu	GTA Val	GAC Asp	ACC Thr	AGC Ser 75	AGC Ser	AAC Asn	CAG Gln	TTC Phe	AGC Ser 80	240
CTG Leu	AGA Arg	CTC Leu	AGC Ser	AGC Ser 85	GTG Val	ACA Thr	TCT Ser	GAG Glu	GAC Asp 90	ACC Thr	GCG Ala	GTC Val	TAT Tyr	TAT Tyr 95	TGT Cys	288
GCA Ala	GAC Asp	GGA Gly	ATG Met 100	TGG Trp	GTA Val	TCA Ser	ACG Thr	GGA Gly 105	TAT Tyr	GCT Ala	CTG Leu	GAC Asp	TTC Phe 110	TGG Trp	GGC Gly	336
						GTC Val										372

# (2) INFORMATION FOR SEQ ID NO:47:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 124 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(~1) SE	OUENCE	DESCRIPTION:	SEQ	ID	NO:47:
---------	--------	--------------	-----	----	--------

Gln Val Gln Leu Gln Glu Ser Gly Pro Gly Leu Val Arg Pro Ser Gln
1 5 10 15

Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Phe Asn Ile Lys Asp Thr 20 25 30

Tyr Met His Trp Val Arg Gln Pro Pro Gly Arg Gly Leu Glu Trp Ile 35 40 45

Gly Arg Ile Asp Pro Ala Ser Gly Asp Thr Lys Tyr Asp Pro Lys Phe 50 55 60

Gln Val Arg Val Thr Met Leu Val Asp Thr Ser Ser Asn Gln Phe Ser 65 70 75 80

Leu Arg Leu Ser Ser Val Thr Ser Glu Asp Thr Ala Val Tyr Tyr Cys
85 90 95

Ala Asp Gly Met Trp Val Ser Thr Gly Tyr Ala Leu Asp Phe Trp Gly 100 105 110

Gln Gly Thr Thr Val Thr Val Ser Ser Gly Glu Ser 115 120

### (2) INFORMATION FOR SEQ ID NO:48:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 31 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1
  - (D) OTHER INFORMATION: /note= "DNA sequence of 704 oligonucleotide"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

#### TGCACTGGGT GAAACAGCGA CCTGGACGAG G

(2) INFORMATION FOR SEQ ID NO:49:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 31 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single

WO 94/16094

	(D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: cDNA	
(ix)	FEATURE:  (A) NAME/KEY: misc_feature  (B) LOCATION: 1  (D) OTHER INFORMATION: /note= "DNA sequence of 705 oligonucleotide"	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:49:	
CCTCGTCC	CAG GTCGCTGTTT CACCCAGTGC A	31
(2) INFO	RMATION FOR SEQ ID NO:50:	
(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 429 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: cDNA	
, ,	FEATURE:  (A) NAME/KEY: sig_peptide  (B) LOCATION: 157  FEATURE:  (A) NAME/KEY: mat_peptide  (B) LOCATION: 58429	
(ix)	FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1429	
(ix)	FEATURE:  (A) NAME/KEY: misc_feature  (B) LOCATION: 1  (D) OTHER INFORMATION: /note= "pBAG185 insert: KRS heavy chain variable region"	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:50:	
	rgg ACC Tgg Agg GTC TTC TgC TTG CTG GCT GTA GCA CCA GGT Frp Thr Trp Arg Val Phe Cys Leu Leu Ala Val Ala Pro Gly -15 -10 -5	48
	TCC CAG GTC CAA CTG CAG GAG AGC GGT CCA GGT CTT GTG AGA Ser Gln Val Gln Leu Gln Glu Ser Gly Pro Gly Leu Val Arg 1 5 10	96

Pro	Ser 15	Gln	Thr	Leu	Ser	Leu 20	TNF	Сув	THE	VAI	25	<b>U</b> I,	TTC Phe			144
AAA Lys 30	GAC Asp	ACC Thr	TAT Tyr	ATG Met	CAC His 35	TGG Trp	GTG Val	AAA Lys	CAG Gln	CGA Arg 40	CCT Pro	GGA Gly	CGA Arg	GGT	CTT Leu 45	192
GAG Glu	TGG Trp	ATT Ile	GGA Gly	AGG Arg 50	ATT Ile	GAT Asp	CCT Pro	GCG Ala	AGT Ser 55	GGC Gly	GAT Asp	ACT	AAA Lys	TAT Tyr 60	GAC Asp	240
CCG Pro	AAG Lys	TTC Phe	CAG Gln 65	GTC Val	AGA Arg	GTG Val	ACA Thr	ATG Met 70	CTG Leu	GTA Val	GAC Asp	ACC Thr	AGC Ser 75	AGC Ser	AAC Asn	288
CAG Gln	TTC Phe	AGC Ser 80	CTG Leu	AGA Arg	CTC Leu	AGC Ser	AGC Ser 85	GTG Val	ACA Thr	GCC Ala	GCC Ala	GAC Asp 90	ACC Thr	GCG Ala	GTC Val	336
TAT Tyr	TAT Tyr 95	TGT Cys	GCA Ala	GAC Asp	GGA Gly	ATG Het 100	TGG Trp	GTA Val	TCA Ser	ACG Thr	GGA Gly 105	TAT Tyr	GCT Ala	CTG Leu	GAC Asp	384
TTC Phe 110	TGG Trp	GGC	CAA Gln	GGG	ACC Thr 115	ACG Thr	GTC Val	ACC	GTC Val	Ser 120	TCA Ser	GGI Gly	GAG	Ser	:	42'

- (2) INFORMATION FOR SEQ ID NO:51:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 143 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:

Met Asp Trp Thr Trp Arg Val Phe Cys Leu Leu Ala Val Ala Pro Gly
-19 -15 -5

Ala His Ser Gln Val Gln Leu Gln Glu Ser Gly Pro Gly Leu Val Arg

Pro Ser Gln Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Phe Asn Ile 15 20 25

Lys Asp Thr Tyr Met His Trp Val Lys Gln Arg Pro Gly Arg Gly Leu 30 45

WO 94/16094 PCT/US94/00266

-107-

Glu Trp Ile Gly Arg Ile Asp Pro Ala Ser Gly Asp Thr Lys Tyr Asp
50 55 60

Pro Lys Phe Gln Val Arg Val Thr Met Leu Val Asp Thr Ser Ser Asn 65 70 75

Gln Phe Ser Leu Arg Leu Ser Ser Val Thr Ala Ala Asp Thr Ala Val 80 85 90

Tyr Tyr Cys Ala Asp Gly Met Trp Val Ser Thr Gly Tyr Ala Leu Asp 95 100 105

Phe Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Gly Glu Ser 110 115 120

- (2) INFORMATION FOR SEQ ID NO:52:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 26 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "DNA sequence of 745 oligonucleotide"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:

TGACCTGCAC CGCGTCTGGC TTCAAC

- (2) INFORMATION FOR SEQ ID NO:53:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 26 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
    - (ii) MOLECULE TYPE: cDNA
    - (ix) FEATURE:
      - (A) NAME/KEY: misc feature
      - (B) LOCATION: 1
      - (D) OTHER INFORMATION: /note= "DNA sequence of 746 oligonucleotide"
    - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:

TTGA	TTGAAGCCAG ACGCGGTGCA GGTCAG (2) INFORMATION FOR SEQ ID NO:54:														26
(2)	INFO	RMATI	ON FOR	SEQ	ID N	10:54	:								
	(Ţ)	(A) (B) (C)	LENGT TYPE: STRAN	H: 42 nucl DEDNI	9 ba Leic ESS:	se p acid sing	airs l								
	(ii)	MOLE	CULE T	YPE:	CDNA										
	(ix)	(B)  FEAT (A) (B)  FEAT (A) (B)  FEAT (A) (B)	NAME/I LOCAT TURE: NAME/I LOCAT TURE: NAME/I LOCAT TURE: NAME/I LOCAT OTHER	ION:  KEY: ION:  KEY: ION:	157  mat 584  CDS 142	pept 29 9 c_fea	ide ature	te=	*p8/	AG 195	ins	sert:	: AS	heavy	
	(xi)	SEQU	JENCE D	ESCR:	[PTIC	on: S	SEQ I	D NC	:54:			·			
ATG Met -19	GAC Asp	TGG A	ACC TGG Thr Trp -15	Arg	GTC Val	TTC Phe	TGC Cys	TTG Leu -10	CTG Leu	GCT Ala	GTA Val	Ala	CCA Pro -5	Gly	48
GCC Ala	CAC His	TCC C	CAG GTO Sin Val	CAA Gln	CTG Leu	CAG Gln 5	GAG Glu	AGC Ser	GGT Gly	CCA Pro	GGT Gly 10	CTT Leu	GTG Val	AGA Arg	96
CCT Pro	AGC Ser 15	CAG A	ACC CTG	AGC Ser	CTG Leu 20	ACC Thr	TGC Cys	ACC Thr	GCG Ala	TCT Ser 25	GCC	TTC Phe	AAC Asn	ATT Ile	144
AAA Lys	GAC Asp	ACC T	TAT ATO	CAC His	TGG Trp	GTG Val	AGA Arg	CAG Gln	CCA Pro	CCT Pro	GGA Gly	CGA Arg	GGT Gly	CTT	192

GAG TGG ATT GGA AGG ATT GAT CCT GCG AGT GGC GAT ACT AAA TAT GAC

Glu Trp Ile Gly Arg Ile Asp Pro Ala Ser Gly Asp Thr Lys Tyr Asp

50

336

384

429

Pro	G AA	G TTO	CAG Gln 65	Val	AGA Arg	GTG Val	ACA Thr	Met 70	Leu	GTA Val	. Asp	ACC Thr	Ser 75	Ser	AAC Aan
CA(	TTO Pho	C AGO Ser 80	Leu	AGA Arg	CTC Leu	AGC Ser	AGC Ser 85	GTG Val	ACA Thr	GCC Ala	GCC Ala	GAC Asp 90	ACC	GCG Ala	GTC Val
TAT	TAT	Cys	GCA Ala	GAC Asp	GGA Gly	ATG Met 100	TGG Trp	GTA Val	TCA Ser	ACG Thr	GGA Gly 105	TAT	GCT Ala	CTG Leu	GAC Asp
TTC Phe 110	TGG Trp	GGC Gly	CAA Gln	GGG Gly	ACC Thr 115	ACG Thr	GTC Val	ACC Thr	GTC Val	TCC Ser 120	TCA Ser	GGT Gly	GAG Glu	TCC Ser	
(2)	INF	ORMA	TION	FOR	SEQ	ID N	10 : 55	:							
		(i)	SEQUI (A) (B) (D)	LEN TYP	CHAR GTH: E: a OLOG	143 mino	ami aci	.no a		3					
	(i	.i) 1	HOLEC	ULE	TYPE	: pr	otei	.n							
	K)	:i) \$	SEQUE	NCE	DESC	RIPT	'ION:	SEC	ID	NO:5	55:				
Met -19	•		SEQUE Thr									Val	Ala	Pro -5	Gly
19	<b>Хвр</b>	Trp		Trp . -15	Arg '	Val	Phe	Сув	Leu -10	Leu	Ala			-5	
19 Ala	Asp His	Trp	Thr	Trp . -15	Arg '	Val Leu	Phe Gln 5	Cys Glu	Leu -10 Ser	Leu Gly	Ala Pro	Gly 10	Leu	-5 Val	Arg
-19 Ala Pro	Asp His Ser 15	Trp Ser Gln	Thr Gln 1	Trp . -15 Val (	Arg v	Val Leu ( Leu ( 20	Phe Gln 5	Cys Glu Cys	Leu -10 Ser	Leu Gly Ala	Ala Pro Ser 25	Gly 10 Gly	Leu Phe	-5 Val Asn	Arg
-19 Ala Pro Lys 30	Asp His Ser 15	Trp Ser Gln Thr	Thr Gln l	Trp15	Arg 'Gln 1 Ser 1 His 1	Val Leu ( 20	Phe Gln 5 Thr	Cys Glu Cys Arg	Leu -10 Ser Thr	Gly Ala Pro 40	Ala Pro Ser 25 Pro	Gly 10 Gly	Leu Phe Arg	-5 Val Asn Gly	Arg Ile Leu 45
-19 Ala Pro Lys 30 Glu	Asp His Ser 15 Asp	Trp Ser Gln Thr	Thr Gln l Thr	Trp -15 Val ( Leu : Met I Arg 1	Arg Ser I	Val Leu ( 20 Trp (	Phe  Gln  5  Thr  Val	Cys Glu Cys Arg	Leu -10 Ser Thr Gln Ser 55	Gly Ala Pro 40 Gly	Ala Pro Ser 25 Pro	Gly 10 Gly Gly	Leu Phe Arg	-5 Val Asn Gly Tyr 60	Arg Ile Leu 45

Tyr Tyr Cys Ala Asp Gly Met Trp Val Ser Thr Gly Tyr Ala Leu Asp 95 100 105

-110-

Phe	Trp	Gly	Gln	Gly	Thr	Thr	Val	Thr	Val	Ser	Ser	Gly	Glu	Ser
110	_	_		_	115					120				

- (2) INFORMATION FOR SEQ ID NO:56:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 29 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "DNA sequence of 915 oligonucleotide"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:

### TATTATTGTG CAAGAGGAAT GTGGGTATC

29

- (2) INFORMATION FOR SEQ ID NO:57:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 29 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "DNA sequence of 917 oligonucleotide"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

## ATACCCACAT TCCTCTTGCA CAATAATAG

- (2) INFORMATION FOR SEQ ID NO:58:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 41 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA

		(A) NAME/REI: MISC_ICACATO	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "DNA sequence of 918	
		oligonucleotide"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:58:	
			41
CTG	CACCG	TG TCTGGCTTCA CCTTCAGCGA CACCTATATG C	••
		NO	
(2)	INFO	RMATION FOR SEQ ID NO:59:	
	(I)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 41 base pairs	
		(B) TYPE: nucleic acid	
,	•	(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
		·	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1 (D) OTHER INFORMATION: /note= "DNA sequence of 919	
		(D) OTHER INFORMATION: Mode - Dan Beddenso of Fig.	
		oligonucleotide"	•
		SEQUENCE DESCRIPTION: SEQ ID NO:59:	
	(x1)	SEQUENCE DESCRIPTION: SEQ 15 NO.331	
		GT GTCGCTGAAG GTGAAGCCAG ACACGGTGCA G	41
GCA	TATAG	GT GTCGCTGAAG GTGAAGCCAG ACACCCTTG. C	
(2)	TNEC	RMATION FOR SEQ ID NO:60:	
(2)	INFC	AMAIION FOR DEG ID NOTOTI	
	(i)	SEQUENCE CHARACTERISTICS:	
	(+)	(A) LENGTH: 31 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
		(b) 10102001 2:00-2	
	633	MOLECULE TYPE: CDNA	
	(11)	NOBBOOKS ::: CV	
	(i =)	FEATURE:	
	(14)	(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note = "DNA sequence of 697	
		oligonucleotide"	
		0219011011011	
	(-1)	SEQUENCE DESCRIPTION: SEQ ID NO:60:	
	(XI)	andanuas analyses seems and an inches	
cc	rcarco:	ACT CCAGCATCGT GATGACCCAG A	41
GG		ici cenacuites, automatem	
(2)	TNE	DRMATION FOR SEQ ID NO:61:	
(4)	TUL	MANITAN FAN ARE DE NATAL	

	(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 31 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: CDNA	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "DNA sequence of 698 oligonucleotide"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:61:	
TCT	GGGTC	CAT CACGATGCTG GAGTGGACAC C	41
(2)	INFO	ORMATION FOR SEQ ID NO:62:	
	(i)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 386 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
	(ix)	FEATURE:	
		(A) NAME/KEY: sig_peptide	
		(B) LOCATION: 157	
	(ix)	FEATURE:	
		(A) NAME/KEY: mat_peptide	
		(B) LOCATION: 58386	
	(ix)	FEATURE:	
		(A) NAME/KEY: CDS	
		(B) LOCATION: 1386	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "pBAG198 insert: VK2 (SVMDY)	
		light chain variable region"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:62:	
		TGG TCC TGC ATC ATC CTG TTC CTG GTT GCT ACC GCT ACC GGT	48
Met	Gly '	Trp Ser Cys Ile Ile Leu Phe Leu Val Ala Thr Ala Thr Gly	
-19		-15 -10 -5	

-113-

GTC Val	CAC His	TCC Ser	AGC Ser	ATC Ile	GTG Val	ATG Met	ACC Thr 5	CAG Gln	AGC Ser	CCA Pro	AGC Ser	AGC Ser 10	CTG Leu	AGC Ser	GCC Ala		96
AGC Ser	GTG Val	GGT Gly	GAC Asp	AGA Arg	GTG Val	Thr	ATC Ile	ACC Thr	TGT Cys	AAG Lys	Ala	AGT Ser	CAG Gln	AGT Ser	GTG Val		144
ACT	15 AAT	GAT	GTA	GCT	TGG	20 TAC	CAG	CAG	AAG	CCA	25 GGT	AAG	GCT	CCA	AAG		192
Thr 30	Asn	Asp	Val	Ala	Trp 35	Tyr	Gln	Gln	Lys	Pro 40	Gly	Lys	Ala	Pro	Lys 45		
CTG Leu	CTG Leu	ATC Ile	TAC Tyr	TAT Tyr 50	GCA Ala	TCC Ser	AAT Asn	CGC Arg	TAC Tyr 55	ACT Thr	GGT Gly	GTG Val	CCA Pro	GAT Asp 60	AGA Arg		240
TTC Phe	AGC Ser	GGT Gly	AGC Ser 65	GGT Gly	TAT Tyr	GGT Gly	ACC Thr	GAC Asp 70	TTC Phe	ACC Thr	TTC Phe	ACC Thr	ATC Ile 75	AGC Ser	AGC Ser		288
								TAC Tyr									336
TCT Ser	CCG Pro 95	TAC Tyr	ACG Thr	TTC Phe	GGC Gly	CAA Gln 100	GGG Gly	ACC Thr	AAG Lys	GTG Val	GAA Glu 105	ATC Ile	AAA Lys	CGT Arg	AAG Lys	TG	386
(2)	INFO	TEMBO	MOT	FOR	SEO	TD B	vo: 63	١:									

- (2) INFORMATION FOR SEQ ID
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 128 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:

Met Gly Trp Ser Cys Ile Ile Leu Phe Leu Val Ala Thr Ala Thr Gly -19 -15

Val His Ser Ser Ile Val Met Thr Gln Ser Pro Ser Ser Leu Ser Ala

Ser Val Gly Asp Arg Val Thr Ile Thr Cys Lys Ala Ser Gln Ser Val

Thr Asn Asp Val Ala Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys 30 35

Leu Leu Ile Tyr Tyr Ala Ser Asn Arg Tyr Thr Gly Val Pro Asp Arg
50 55 60

Phe Ser Gly Ser Gly Tyr Gly Thr Asp Phe Thr Phe Thr Ile Ser Ser 65 70 75

Leu Gln Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln Gln Asp Tyr Ser

Ser Pro Tyr Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg Lys 95 100 105

- (2) INFORMATION FOR SEQ ID NO:64:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 32 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note = "DNA sequence of 803 oligonucleotide"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:64:

GGTGTCCACT CCGACATCCA GATGACCCAG AG

32

- (2) INFORMATION FOR SEQ ID NO:65:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 32 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
    - (ii) MOLECULE TYPE: cDNA
    - (ix) FEATURE:
      - (A) NAME/KEY: misc feature
      - (B) LOCATION: 1
      - (D) OTHER INFORMATION: /note= "DNA sequence of 804 oligonucleotide"
    - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:65:

CTCTGGGTCA TCTGGATGTC GGAGTGGACA CC

(2) INFORMATION FOR SEQ ID NO.0	(2)	INFORMATION	FOR	SEQ	ID	NO:66
---------------------------------	-----	-------------	-----	-----	----	-------

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 386 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: sig peptide
  - (B) LOCATION: 1..57
- (ix) FEATURE:
  - (A) NAME/KEY: mat\_peptide
  - (B) LOCATION: 58..386
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 1..386
- (ix) FEATURE:
  - (A) NAME/KEY: misc feature
  - (B) LOCATION: 1
  - (D) OTHER INFORMATION: /note= "pBAG197 insert: VK3 (DQMDY) light chain variable region"
- (x1) SEQUENCE DESCRIPTION: SEQ ID NO:66:
- ATG GGT TGG TCC TGC ATC CTG TTC CTG GTT GCT ACC GCT ACC GGT

  Met Gly Trp Ser Cys Ile Ile Leu Phe Leu Val Ala Thr Ala Thr Gly

  -19

  -10

  -5
- GTC CAC TCC GAC ATC CAG ATG ACC CAG AGC CCA AGC AGC CTG AGC GCC

  Val His Ser Asp Ile Gln Met Thr Gln Ser Pro Ser Ser Leu Ser Ala

  1 5 10
- AGC GTG GGT GAC AGA GTG ACC ATC ACC TGT AAG GCC AGT CAG AGT GTG

  Ser Val Gly Asp Arg Val Thr Ile Thr Cys Lys Ala Ser Gln Ser Val

  15 20 25
- ACT AAT GAT GTA GCT TGG TAC CAG CAG AAG CCA GGT AAG GCT CCA AAG

  Thr Asn Asp Val Ala Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys

  30 35 40 45
- CTG CTG ATC TAC TAT GCA TCC AAT CGC TAC ACT GGT GTG CCA GAT AGA
  Leu Leu Ile Tyr Tyr Ala Ser Asn Arg Tyr Thr Gly Val Pro Asp Arg
  50 55 60

#### -116-

TTC Phe	AGC Ser	GGT Gly	AGC Ser 65	GGT Gly	TAT Tyr	GGT Gly	ACC Thr	GAC Asp 70	TTC Phe	ACC Thr	TTC Phe	ACC Thr	ATC Ile 75	AGC Ser	AGC Ser		288
CTC Leu	CAG Gln	CCA Pro 80	GAG Glu	GAC Asp	ATC Ile	GCC Ala	ACC Thr 85	TAC Tyr	TAC Tyr	TGC Cys	CAG Gln	CAG Gln 90	GAT Asp	TAT Tyr	AGC Ser		336
TCT Ser	CCG Pro 95	TAC Tyr	ACG Thr	TTC Phe	GGC Gly	CAA Gln 100	GGG Gly	ACC Thr	AAG Lys	GTG Val	GAA Glu 105	ATC Ile	AAA Lys	CGT Arg	AAG Lys	TG	386

- (2) INFORMATION FOR SEQ ID NO:67:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 128 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:

Met Gly Trp Ser Cys Ile Ile Leu Phe Leu Val Ala Thr Ala Thr Gly
-19 -15 -10 -5

Val His Ser Asp Ile Gln Met Thr Gln Ser Pro Ser Ser Leu Ser Ala
1 5 10

Ser Val Gly Asp Arg Val Thr Ile Thr Cys Lys Ala Ser Gln Ser Val

Thr Asn Asp Val Ala Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys 30 40 45

Leu Leu Ile Tyr Tyr Ala Ser Asn Arg Tyr Thr Gly Val Pro Asp Arg
50 55 60

Phe Ser Gly Ser Gly Tyr Gly Thr Asp Phe Thr Phe Thr Ile Ser Ser 65 70 75

Leu Gln Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln Gln Asp Tyr Ser

Ser Pro Tyr Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg Lys 95 100 105

- (2) INFORMATION FOR SEQ ID NO:68:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 429 base pairs
    - (B) TYPE: nucleic acid

(C)	STRANDEDNESS:	single

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1..429
  - (D) OTHER INFORMATION: /note= "pMDR1023 insert: PDLN heavy chain variable region"
- (ix) FEATURE:
  - (A) NAME/KEY: sig\_peptide
  - (B) LOCATION: 1..57
- (ix) FEATURE:
  - (A) NAME/KEY: mat\_peptide
  - (B) LOCATION: 58..429
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 1..429
- (xi) SEQUENCE DESCRIPTION: SEQ. ID NO:68:

ATG	GAC	TGG	ACC	TGG	AGG	GTC	TTC	TGC	TTG	CTG	GCT	GTA	GCA	CCA	GGT	48
Met	Asp	Trp	Thr	Trp	Arg	Val	Phe	Сув	Leu	Leu	Ala	Val	Ala	Pro	Gly	
-19				-15					-10					-5		
666	~~	maa	<b>C</b>	CEC	~~~	CTG	C1.C	C3.C	<b>***</b>			C				06
																. 96
MIG	ura	ser	GIN	ÁŒI	GIN	Leu	GIN	GIU	ser	GIY	MIG	10	VAI	ATT	Lys	
			1				3					10				
CCG	GGT	TCC	TCC	GTT	AAA	CTG	TCC	TGC	AAA	GCT	TCC	GGT	TTC	AAC	ATC	144
Pro	Gly	Ser	Ser	Val	Lys	Leu	Ser	Cys	Lys	Ala	Ser	Gly	Phe	Asn	Ile	
	15				•	20		•	•		25	•				
AAA	GAC	ACC	TAC	ATG	CAC	TGG	GTT	AAA	CAG	CGT	CCG	GGT	CAG	GGT	CTG	192
Lys	Asp	Thr	Tyr	Met	His	Trp	Val	Lys	Gln	Arg	Pro	Gly	Gln	Gly	Leu	
30					35					40			•		45	
GAA	TGG	ATC	GGT	CGT	ATC	GAC	CCG	GCT	TCC	CCT	GAC	ACC	AAA	TAC	GAC	240
Glu	Trp	Ile	Gly	Arg	Ile	Asp	Pro	Ala	Ser	Gly	Хвр	Thr	Lys	Tyr	Asp	
				50					55					60		
						GCT										288
Pro	Lys	Phe		Val	Lys	Ala	Thr		Thr	Ala	Asp	Glu		Thr	Ser	
			65					70					75			
N.C.C	C CTT	T. C.	CTC	CAA	~~~	TCC	TCC	CTC	CCT	TCC	CAA	CAC	» CC	COT	C TOTAL	336
						Ser										336
THE	VIG	80	Leu	GIU	Leu	Jer	85	reu	arg	361	GIU	90	Int	VIG	AGI	
		<del></del>					6.7					70				

						GCT Ala		384
						GAG Glu		429

- (2) INFORMATION FOR SEQ ID NO:69:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 143 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:69:

Met Asp Trp Thr Trp Arg Val Phe Cys Leu Leu Ala Val Ala Pro Gly -19 -15 -10 -5

Ala His Ser Gln Val Gln Leu Gln Glu Ser Gly Ala Glu Val Val Lys
1 5 10

Pro Gly Ser Ser Val Lys Leu Ser Cys Lys Ala Ser Gly Phe Asn Ile 15 20 25

Lys Asp Thr Tyr Met His Trp Val Lys Gln Arg Pro Gly Gln Gly Leu 30 45

Glu Trp Ile Gly Arg Ile Asp Pro Ala Ser Gly Asp Thr Lys Tyr Asp
50 55 60

Pro Lys Phe Gln Val Lys Ala Thr Ile Thr Ala Asp Glu Ser Thr Ser
65 70 75

Thr Ala Tyr Leu Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Val

Tyr Tyr Cys Ala Asp Gly Met Trp Val Ser Thr Gly Tyr Ala Leu Asp 95 100 105

Phe Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Gly Glu Ser 110 120

- (2) INFORMATION FOR SEQ ID NO:70:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 383 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single

		(D)	TOP	OLOG	Y: 1	inea	r									
	(ii)	MOLI	ECULE	TYP	PE: C	DNA										
	(ix)		IAN	Œ/KE CATIO ER	N: I	nisc_ 1376 RMATI ariab	ON:	/not	e= " on"	'pMDF	11025	ins	ert:	PDI	N light	
*	(ix)	FEA (A) (B)	TURE: NAI LO	KE/KE	Y:	sig_I 157	p <b>ep</b> t:	ide								
	(ix)	FEA (A) (B)	NA	ME/KI	Y: :	mat_j 5837	pept. 16	ide			-					
	(ix)	FEA (A) (B)	NA.	ME/KI	EY: ON:	CDS 1376	5									
		SEC														40
ATG Met -19	GGT Gly	TGG Trp	TCC Ser	TGC Cys -15	ATC Ile	ATC Ile	CTG Leu	TTC Phe	CTG Leu -10	GTT Val	GCT Ala	ACC (	GCT Ala	ACC Thr -5	GGT Gly	48
GTI Val	CAC His	TCC Ser	ATC Ile	GTT Val	ATG Met	ACC Thr	CAG Gln 5	TCC Ser	CCG Pro	GAC Asp	TCC Ser	CTG Leu 10	GCT Ala	GTT Val	TCC Ser	96
CTC	GGT Gly 15	GAA Glu	CGT Arg	GTT Val	ACC Thr	ATC Ile 20	AAC Asn	TGC Cys	AAA Lys	GCT Ala	TCC Ser 25	CAG Gln	TCC Ser	GTT Val	ACC Thr	144
AA( As:	GAC Asp	GTT Val	GCT Ala	TGG Trp	TAC Tyr 35	CAG Gln	CAG Gln	AAA Lys	CCG Pro	GGT Gly 40	CAG Gln	TCC Ser	CCG Pro	AAA Lys	CTG Leu 45	192
CT(	; ATC	TAC Tyr	TAC Tyr	GCT Ala 50	TCC Ser	AAC Asn	CGT Arg	TAC Tyr	ACC Thr 55	GGT	GTT Val	CCG Pro	GAC Asp	Arg 60	TTC Phe	240
TC: Se	c GGT r Gly	TCC Ser	GGT Gly 65	TAC	GGT Gly	ACC Thr	GAC Asp	TTC Phe 70	ACC	TTC Phe	ACC Thr	ATC Ile	Ser 75	: TCC : Ser	GTT Val	288
CA G1	G GC1 n Ala	GAA Glu 80	Asp	GTT Val	GCI	GTT Val	TAC Tyr 85	Tyr	Cys	CAG	CAG Glr	GAC Asp 90	TAC	C TCC	C TCC	336

WO 94/16094 PCT/US94/00266

-120-

CCG TAC ACC TTC GGT GGT ACC AAA CTG GAG ATC TAA GGA TCC TC

Pro Tyr Thr Phe ly Gly Gly Thr Lys Leu Glu Ile \*

95 100 105

- (2) INFORMATION FOR SEQ ID NO:71:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 124 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:71:

Met Gly Trp Ser Cys Ile Ile Leu Phe Leu Val Ala Thr Ala Thr Gly
-19 -15 -10 -5

Val His Ser Ile Val Met Thr Gln Ser Pro Asp Ser Leu Ala Val Ser
1 5 10

Leu Gly Glu Arg Val Thr Ile Asn Cys Lys Ala Ser Gln Ser Val Thr
15 20 25

Asn Asp Val Ala Trp Tyr Gln Gln Lys Pro Gly Gln Ser Pro Lys Leu 30 40 45

Leu Ile Tyr Tyr Ala Ser Asn Arg Tyr Thr Gly Val Pro Asp Arg Phe
50 55 60

Ser Gly Ser Gly Tyr Gly Thr Asp Phe Thr Phe Thr Ile Ser Ser Val 65 70 75

Gln Ala Glu Asp Val Ala Val Tyr Tyr Cys Gln Gln Asp Tyr Ser Ser 80 85 90

Pro Tyr Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile 95 100 105

- (2) INFORMATION FOR SEQ ID NO:72:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 60 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1

- (D) OTHER INFORMATION: /note= "Oligo 370-119 corresponding to 58-117 VH-PDLN"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:72:

CAGGITCAGC TGCAGGAGIC CGGTGCTGAA GITGITAAAC CGGGTTCCTC CGTTAAACTG 60

- (2) INFORMATION FOR SEQ ID NO:73:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 60 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: CDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "Oligo 370-120 corresponds to 118-177 VH-PDLN"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:73:

TCCTGCARAG CTTCCGGTTT CARCATCARA GACACCTACA TGCACTGGGT TARACAGCGT 60

- (2) INFORMATION FOR SEQ ID NO:74:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 60 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "Oligo 370-121 corresponds to 178-237 VH-PDLN"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:74:

CCGGGTCAGG GTCTGGAATG GATCGGTCGT ATCGACCCGG CTTCCGGTGA CACCAAATAC 60

- (2) INFORMATION FOR SEQ ID NO:75:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 66 base pairs
    - (B) TYPE: nucleic acid

-122-

		(C) STRANDEDNESS: single (D) TOPOLOGY: linear		
	(ii) MOLECULE TYPE: cDNA			
	(ix)	FEATURE:  (A) NAME/KEY: misc_feature  (B) LOCATION: 1  (D) OTHER INFORMATION: /note= "Oligo 370-122 corresponds to 238-303 VH-PDLN"		
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:75:		
GAC	CCGAP	AAT TCCAGGTTAA AGCTACCATC ACCGCTGACG AATCCACCTC CACCGCTTAC	60	
CTG	GAA		66	
(2)	INFO	ORMATION FOR SEQ ID NO:76:		
	(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 63 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear		
	(ii)	MOLECULE TYPE: cDNA		
	(ix)	FEATURE:  (A) NAME/KEY: misc_feature  (B) LOCATION: 1  (D) OTHER INFORMATION: /note= "Oligo 370-123 corresponds to 304-366 VH-PDLN"		
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:76:		
CTG	TCCTC	CC TGCGTTCCGA AGACACCGCT GTTTACTACT GCGCTGACGG TATGTGGGTT	60	
TCC			63	
(2)	INFO	RMATION FOR SEQ ID NO:77:		
	(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 54 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	-	
	(ii)	MOLECULE TYPE: CDNA		
	(ix)	FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1		

		(D) OTHER INFORMATION: /note= "Oligo 370-124 corresponds to 367-420 VH-PDLN"	
	(xi)	) SEQUENCE DESCRIPTION: SEQ ID NO:77:	
AC	CGGTT	ACG CTCTGGACTT CTGGGGTCAG GGTACCACGG TCACCGTTTC CTCC	54
(2)	INF	ORMATION FOR SEQ ID NO:78:	
	(i)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 63 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
	(iv)	ANTI-SENSE: YES	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "Oligo 370-125 corresponds to reverse VH-PDLN 420-358"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:78:	
GGA	GGAAA	CG GTGACCGTGG TACCCTGACC CCAGAAGTCC AGAGCGTAAC CGGTGGAAAC	60
CCA			63
			63
(2)	INFO	RMATION FOR SEQ ID NO:79:	
	(i)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 47 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: CDNA	
	(iv)	ANTI-SENSE: YES	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note = "Oligo 370-126 corresponds	
		to reverse VH-PDLN 357-311"	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:79:

CATACCGTCA GCGCAGTAGT AAACAGCGGT GTCTTCGGAA CGCAGGG

(2)	INFO	ORMATION FOR SEQ 1D NO:80:	
	(i)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 67 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	HOLECULE TYPE: cDNA	
	(iv)	ANTI-SENSE: YES	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "Oligo 370-127 corresponds to reverse VH-PDLN 310-244"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:80:	
AGG	ACAGT	TC CAGGTAAGCG GTGGAGGTGG ATTCGTCAGC GGTGATGGTA GCTTTAACCT	60
GGA	ATTT		67
(2)	INFO	RMATION FOR SEQ ID NO:81:	
	(i)	SEQUENCE CHARACTERISTICS:	
	<b>\-/</b>	(A) LENGTH: 60 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
	(iv)	ANTI-SENSE: YES	
	(ix)	FEATURE:	
	•	(A) NAME/KEY: misc_feature	
	-	(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "Oligo 370-128 corresponds to reverse VH-PDLN 243-186"	1
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:81:	
CGG	GTCGT	AT TTGGTGTCAC CGGAAGCCGG GTCGATACGA CCGATCCATT CCAGACCCTG	60
(2)	INFO	RMATION FOR SEQ ID NO:82:	
	(i)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 60 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		- ·	

(D) TOPOLOGY: linear

	(ii)	MOLECULE TYPE: cDNA				
	(iv)	ANTI-SENSE: YES				
	(ix)	FEATURE:  (A) NAME/KEY: misc_feature  (B) LOCATION: 1  (D) OTHER INFORMATION: /note= "Oligo 370-129 corresponds to reverse VH-PDLN 185-124"				
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:82:				
AC	CCGGAC	CGC TGTTTAACCC AGTGCATGTA GGTGTCTTTG ATGTTGAAAC CGGAAGCTTT	60			
(2)	INFO	PRMATION FOR SEQ ID NO:83:				
	(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 66 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear				
	(ii)	MOLECULE TYPE: cDNA				
	(iv)	ANTI-SENSE: YES				
	(ix)	FEATURE:  (A) NAME/KEY: misc_feature  (B) LOCATION: 1  (D) OTHER INFORMATION: /note= "Oligo 370-130 corresponds to reverse VH-PDLN 123-58"				
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:83:				
GCA	GGACA	GT TTAACGGAGG AACCCGGTTT AACAACTTCA GCACCGGACT CCTGCAGCTG	60			
AAC	CTG		66			
(2)	INFO	RMATION FOR SEQ ID NO:84:				
	(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 66 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	÷ .			
	(ii)	MOLECULE TYPE: cDNA				
	(ix)	FEATURE: (A) NAME/KEY: misc_feature				

(B) LOCATION: 1

		to 1-58 VK-PDLN"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:84:	
AGC	TTACC	AT GGGTTGGTCC TGCATCATCC TGTTCCTGGT TGCTACCGCT ACCGGTGTTC	60
ACI	CCA		66
(2)	INFO	RMATION FOR SEQ ID NO:85:	
	<b>(1)</b>	SEQUENCE CHARACTERISTICS:	
	(-)	(A) LENGTH: 66 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
	(ix)	FEATURE:	
	()	(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= *Oligo 370-132 corresponds	
		to 59-124 VK-PDLN"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:85:	
TCG	TTATG	AC CCAGTCCCCG GACTCCCTGG CTGTTTCCCT GGGTGAACGT GTTACCATCA	60
ACT	GCA		66
(2)	INFO	RMATION FOR SEQ ID NO:86:	
	(i)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 66 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	•
	(ii)	MOLECULE TYPE: cDNA	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "Oligo 370-133 corresponds	
		to 125-190 VK-PDLN"	

	(X1)	SEQUENCE DESCRIPTION: SEQ ID NO:86:	
AA	GCTTC	CCA GTCCGTTACC AACGACGTTG CTTGGTACCA GCAGAAACCG GGTCAGTCCC	60
CG	AAAC		<b>6</b> 6
(2)	INF	ORMATION FOR SEQ ID NO:87:	
	(i)	SEQUENCE CHARACTERISTICS:	
	ν-,	(A) LENGTH: 66 base pairs	
	•	(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
	(ix)	FEATURE:	
		(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "Oligo 370-134 corresponds	
		to 191-256 VK-PDLN"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:87:	
TGC	TGATO	TA CTACGCTTCC AACCGTTACA CCGGTGTTCC GGACCGTTTC TCCGGTTCCG	60
GTI	ACG		66
(2)	INFC	RMATION FOR SEQ ID NO:88:	
	(i)		
		(A) LENGTH: 66 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: CDNA	
	(ix)	FEATURE:	
	` ,	(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "Oligo 370-135 corresponds	
		to 257-322 VK-PDLN*	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:88:	-
GTA	CCGAC	TT CACCTTCACC ATCTCCTCCG TTCAGGCTGA AGACGTTGCT GTTTACTACT	60
GCC	AGC		66
(2)	INFO	RMATION FOR SEQ ID NO:89:	

		THE COURSE OF CO	
	(Ŧ)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 54 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
		ALLE SERVE SERVE ARVE	
	(ii)	MOLECULE TYPE: cDNA	
	(i v)	FEATURE:	
	(,	(A) NAME/KEY: misc_feature	
		(R) IOCATION: 1	
		(D) OTHER INFORMATION: /note= "Oligo 370-136 corresponds	
		to 323-376 VK-PDLN"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:89:	
	. ~~ ~	TC CTCCCCGTAC ACCTTCGGTG GTGGTACCAA ACTGGAGATC TAAG	54
AGG	ACTAC	IC CICCOGIAC ACCITOGIA CICCOGIA	
(2)	TNFO	RMATION FOR SEQ ID NO:90:	
(-)			
	(i)	SEQUENCE CHARACTERISTICS:	
	, ,	(A) LENGTH: 63 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
		WAT DAVIT D. MVDP. ADVA	
	(11)	MOLECULE TYPE: CDNA	
	(iv)	ANTI-SENSE: YES	
	(,		
	(ix)	FEATURE:	
	` '	(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "Oligo 370-137 corresponds	
		to reverse VK-PDLN 380-318"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:90:	
		GA TCTCCAGTTT GGTACCACCA CCGAAGGTGT ACGGGGAGGA GTAGTCCTGC	60
GAT	CCTTA	GA TCTCCAGTTT GGTACCACCA CCGAAGGTGT AGGGGGTGT	
TGG			63
166			
(2)	INFO	RMATION FOR SEQ ID NO:91:	-
ζ- /			-
	(i)	SEQUENCE CHARACTERISTICS:	
		(A) LENGTH: 66 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	

	(= +)		
	(ix)	FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "Oligo 370-138 corresponds to reverse VK-PDLN 317-252"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:91:	
CAG	TAGTA	AA CAGCAACGTC TTCAGCCTGA ACGGAGGAGA TGGTGAAGGT GAAGTCGGTA	60
CCG	TAA		66
(2)	INFO	RMATION FOR SEQ ID NO:92:	
	(i)	SEQUENCE CHARACTERISTICS:	
	(-)	(A) LENGTH: 66 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
	(iv)	ANTI-SENSE: YES	•
	(ix)	FEATURE:	
	(/	(A) NAME/KEY: misc_feature	
		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "Oligo 370-139 corresponds to reverse VK-PDLN 251-186"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:92:	
CCG	GAACC	GG AGAAACGGTC CGGAACACCG GTGTAACGGT TGGAAGCGTA GTAGATCAGC	60
agt	TTC		66
<b>(2)</b>	INFO	RMATION FOR SEQ ID NO:93:	
	(i)	SEQUENCE CHARACTERISTICS:	
	<b>\</b>	(A) LENGTH: 66 base pairs	
		(B) TYPE: nucleic acid	
		(C) STRANDEDNESS: single	
		(D) TOPOLOGY: linear	-
	(ii)	MOLECULE TYPE: cDNA	
	(iv)	ANTI-SENSE: YES	
	(ix)	FEATURE:	
	\/	(A) NAME/KEY: misc feature	

		(B) LOCATION: 1	
		(D) OTHER INFORMATION: /note= "Oligo 370-140 corresponds to r verse VK-PDLN 185-120"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:93:	
GGC	GACTG	AC CCGGTTTCTG CTGGTACCAA GCAACGTCGT TGGTAACGGA CTGGGAAGCT	60
TTC	CAG		66
(2)	INFO	RMATION FOR SEQ ID NO:94:	
	<b>(i)</b>	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 66 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
	(iv)	ANTI-SENSE: YES	
	(ix)	FEATURE:  (A) NAME/KEY: misc_feature  (B) LOCATION: 1  (D) OTHER INFORMATION: /note= "Oligo 370-141 corresponds to reverse VK-PDLN 119-54"	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:94:	
TTG	ATGGT.	AA CACGTTCACC CAGGGAAACA GCCAGGGAGT CCGGGGACTG GGTCATAACG	60
ATG	GAG		66
(2)	INFO	RMATION FOR SEQ ID NO:95:	
	(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 57 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
	(ii)	MOLECULE TYPE: cDNA	
	(iv)	ANTI - SENSE: YES	
	(ix)	FEATURE:	

(D) OTHER INFORMATION: /note= "Oligo 370-142 corresponds

(A) NAME/KEY: misc\_feature

to reverse VK-PDLN 53-1"

(B) LOCATION: 1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:95:

TG	AACAC	CGG TA	GCGGTAGC AACCAGGAAC AGGATGATGC AGGACCAACC CATGGTA	5
(2)	INFO	RHATI	ON FOR SEQ ID NO:96:	
	(i)	SEOU	ENCE CHARACTERISTICS:	
	(-/	_	LENGTH: 51 base pairs	
			TYPE: nucleic acid	
			STRANDEDNESS: single	
		(D)	TOPOLOGY: linear	
	(11)	HOLE	CULE TYPE: cDNA	
	(ix)	FEAT		
			NAME/KEY: misc_feature	
			LOCATION: 1	
		(D)	OTHER INFORMATION: /note= "DNA sequence of VK1-DQL primer 307-247"	
	(xi)	SEQU	ENCE DESCRIPTION: SEQ ID NO:96:	
ACC	GCTAC	CG GT	GTTCACTC CGACATCCAG CTGACCCAGA GCCCAAGCAG C	51
(2)	INFO	RMATI	ON FOR SEQ ID NO:97:	
	(i)	SEQUI	ENCE CHARACTERISTICS:	
		(A)	LENGTH: 56 base pairs	
		(B)	TYPE: nucleic acid	
		(C)	STRANDEDNESS: single	
		(D)	TOPOLOGY: linear	
	(ii)	MOLE	CULE TYPE: cDNA	
	(ix)	FEAT	JRE:	
		(A)	NAME/KEY: misc_feature	
		(B)	LOCATION: 1	
		(D)	OTHER INFORMATION: /note= "DNA sequence of VK1-DQL primer 370-210"	
	(xi)	SEQUE	ENCE DESCRIPTION: SEQ ID NO:97:	
CTG	AGGAT	CC AGA	NAAGTGCA CTTACGTTTG ATTTCCACCT TGGTCCCTTG GCCGAA	56
(2)	INFO	CITAMS	ON FOR SEQ ID NO:98:	
	(i)	_	ENCE CHARACTERISTICS:	
			LENGTH: 51 base pairs	
			TYPE: nucleic acid	
			STRANDEDNESS: single	
		(D)	TOPOLOGY: linear	
			•	

- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1
  - (D) OTHER INFORMATION: /note= "DNA sequence of VK2-SVMDY primer 370-269"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:98:

# CTCTCCACCG GTGTCCACTC CAGCATCGTG ATGACCCAGA GCCCAAGCAG C

51

- (2) INFORMATION FOR SEQ ID NO:99:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 51 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: misc\_feature
    - (B) LOCATION: 1
    - (D) OTHER INFORMATION: /note= "DNA sequence of VK3-DQMDY primer 370-268"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:99:

CTCTCCACCG GTGTCCACTC CGACATCCAG ATGACCCAGA GCCCAAGCAG C

## WHAT IS CLAIMED IS:

- 1. A recombinant antibody molecule comprising antigen binding regions derived from the heavy or light chain variable regions of an anti-VLA4 antibody.
- 2. A humanized recombinant antibody molecule having specificity for VLA4 and having an antigen binding site wherein at least one of the complementarity determining regions (CDR) of the variable regions are derived from a non-human anti-VLA4 antibody.

- 3. A humanized recombinant heavy chain according to claim 2 comprising non-human CDRs at positions 31-35 (CDR1), 50-65 (CDR2) and 95-102 (CDR3) (Kabat numbering).
- 4. A humanized recombinant heavy chain according to claim 3 comprising non-human residues at framework positions 27-30 (Kabat numbering).
- 5. A humanized recombinant heavy chain according to claim 4 comprising additional non-human residues at framework position 75 (Kabat numbering).
- 6. A humanized recombinant heavy chain according to claim 5 comprising additional non-human residues at framework position(s) 77-79 or 66-67 and 69-71 or 84-85 or 38 and 40 or 24.
- 7. A humanized recombinant light chain according to claim 2 comprising non-human CDRs at positions 24-34 (CDR1), 50-56 (CDR2) and 89-97 (CDR3).
- 8. A humanized recombinant light chain according to claim 7 comprising non-human residues at framework positions 60 and 67.

- 9. A humanized recombinant antibody molecule c mprising at least one antibody heavy chain according to claim 3 and at least one antibody light chain according to claim 7.
- 10. A humanized recombinant antibody molecule according to claim 7 wherein the non-human CDRs are derived from the HP1/2 murine monoclonal antibody.
- 11. DNA encoding an antibody heavy chain according to claim 3.
- 12. DNA encoding an antibody light chain according to claim 7.
- 13. DNA encoding an antibody molecule according to claim 10.
  - 14. A vector comprising DNA according to claim 11.
  - 15. A vector comprising DNA according to claim 12.
  - 16. A vector comprising DNA according to claim 13.
- 17. An expression vector comprising DNA encoding an antibody heavy chain according to claim 3 in operative combination with DNA encoding an antibody light chain according to claim 7.
- 18. An expression vector comprising DNA encoding an antibody molecule according to claim 10.
- 19. Host cells transformed with a vector according to claim 14 and a vector according to claim 15.

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- 20. Host cells transformed with a vector according to claim 16.
- 21. A process for the production of a humanized recombinant anti-VLA4 antibody comprising:
  - (a) producing an expression vector comprising an operon having a DNA sequence encoding an antibody heavy or light chain wherein at least one of the CDRs of the variable domain are derived from a non-human anti-VLA4 antibody and the remaining immunoglobulin-derived parts of the antibody chain are derived from a human immunoglobulin;
- (b) producing an expression vector comprising an operon having a DNA sequence encoding a complementary antibody light or heavy chain wherein at least one of the CDRs of the variable domain are derived from a non-human anti-VLA4 antibody and the remaining immunoglobulinderived parts of the antibody chain are derived from a human immunoglobulin;
  - (c) transfecting a host cell with each vector; and
- (d) culturing the transfected cell line to produce the humanized recombinant anti-VLA4 antibody 25 molecule.
  - 22. A process according to claim 21 wherein the DNA sequence encoding the heavy chain and the light chain comprise the same vector.
  - 23. A therapeutic composition comprising an antibody molecule, or a fragment thereof, according to claim 1 in combination with a pharmaceutically acceptable diluent, excipient or carrier.

WO 94/16094 PCT/US94/00266

- 24. A diagnostic composition c mprising an antibody molecule, or a fragment thereof, according to claim 1 in a detectably labelled form.
- 25. A method of treatment comprising administering an effective therapeutic amount of an antibody according to claim 1 to a human or animal subject.
- from a response of a specific defense system in a mammalian subject which comprises providing to a subject in need of such treatment an amount of an anti-inflammatory agent sufficient to suppress the inflammation, wherein the anti-inflammatory agent is an antibody according to claim 1.

15

- 27. A humanized recombinant anti-VLA4 antibody molecule having the characteristics of an antibody which comprises a humanized heavy chain comprising a variable heavy chain region selected from the group consisting of  $V_H$  STAW (SEQ ID NO:39),  $V_H$  KAITAS (SEQ ID NO:43),  $V_H$  SSE (SEQ ID NO:47),  $V_H$  KRS (SEQ ID NO:51), and  $V_H$  AS (SEQ ID NO: 55), in combination with a humanized light chain comprising a light chain variable region selected from the group consisting of VK DQL (SEQ ID NO: 31), VK2 SVMDY (SEQ ID NO: 63), and VK3 DQMDY (SEQ ID NO: 67).
- 28. DNA encoding the humanized heavy chain and the humanized light chain according to claim 27.
  - 29. A vector comprising DNA according to claim 28.
- 30. An expression vector comprising DNA encoding an antibody molecule according to claim 27.



- 31. Host cells transf rmed with a vector according to claim 29.
- 32. Host cells transformed with a vector according to claim 30.
- 33. Host cells according to claim 32 that are ATCC CRL 11175.
- 34. A humanized recombinant anti-VLA4 antibody molecule having a potency from about 20% to about 100% of the potency of an antibody which comprises a humanized heavy chain comprising a variable heavy chain region of V<sub>H</sub> AS (SEQ ID NO: 55), in combination with a humanized light chain comprising a light chain variable region of VK2 SVMDY (SEQ ID NO: 63).
  - 35. A therapeutic composition comprising an antibody molecule, or a fragment thereof, according to claim 27 or 34 in combination with a pharmaceutically acceptable diluent, excipient or carrier.
  - 36. A diagnostic composition comprising an antibody molecule, or a fragment thereof, according to claim 27 or 34 in a detectably labelled form.
  - 37. A method of treatment comprising administering an effective amount of an antibody according to claim 27 or 34 to a human or animal subject.

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- 38. A method for treating inflammation resulting from a response of a sp cific defense system in a mammalian subject which comprises providing to a subject in need of such treatment an amount of an anti-inflammatory agent sufficient to suppress the inflammation, wherein the anti-inflammatory agent is an antibody according to claim 27 or 34.
- 39. A humanized recombinant anti-VLA4 antibody molecule that is the antibody produced by ATCC CRL 11175 or an antibody having the characteristics of the antibody produced by ATCC CRL 11175.
- 40. A humanized recombinant anti-VLA4 antibody molecule that has a potency from about 20% to about 100% of the potency of the antibody produced by ATCC CRL 11175.
- 41. A humanized recombinant anti-VLA4 antibody molecule that has a potency from about 20% to about 100% of the potency of the antibody produced by the murine monoclonal antibody HP1/2.